
生命分子システムにおける 動的秩序形成と高次機能発現

領域番号 2501

平成25年度～平成29年度

科学研究費助成事業（科学研究費補助金）

新学術領域研究（研究領域提案型）研究成果報告書

2019年6月

領域代表者 加藤 晃一

自然科学研究機構 生命創成探究センター・教授

目次

はしがき	2
研究組織	3
交付決定額（配分額）	9
研究発表	10
(1) 雑誌論文（査読あり。共同研究の場合、重複あり。）	10
(2) 学会発表	79
(3) 図書	121
研究成果による産業財産権の出願・取得状況	129
研究成果	130
研究領域の目的及び概要	130
主な研究成果	135

はしがき

本研究成果報告書は、平成 25 年度から 29 年度の期間において、科学研究費補助金新学術領域研究（研究領域提案型）により行われた「生命分子システムにおける動的秩序形成と高次機能発現」（領域番号 2501）に関する成果をまとめたものである。

生命現象の特徴は、複雑な柔構造を有する多様な生命分子素子が動的な集合体を形成することにより、自律的に秩序あるシステムを創出することにある。こうしたシステムの形成原理を解明することは生命現象の本質的理解につながるはずである。本研究領域は、生命分子システムを構成する多数の素子がダイナミックな離合集散を通じて動的秩序を形成し、それが時間発展して高次機能を発現する仕組みを分子科学の観点から解き明かすことを目指す。そのために物理化学に基盤をおく実験と理論の融合研究を展開する。さらに、生命分子科学と超分子化学のアプローチを発展的に統合することによって、生命分子システムの特質を具現化した動的秩序系を人工的に構築することを目指す。この目的を実現するために、生命分子科学を基軸に、生物物理学、理論・計算科学、合成化学、構造・システム生物学、さらには医学・薬学・工学・環境科学等への応用を見据えた研究ネットワークを組織する。こうした国際的にも類例のない学際的な研究体制を構築することにより、生命の本質的理解に向けた先端的な学術領域を創成する。その成果は、創薬をはじめとする産業応用の進展に資するとともに、生命科学一般の深化と分子科学におけるパラダイムシフトをもたらし、人工的な生命システムを設計・創生するための指導原理を導き出すことが期待される。

本研究領域では、動的秩序の探査 (A01)・創生 (A02)・展開 (A03) の 3 つの研究項目を研究の柱とし、各項目内において化学・物理学・生物学の分野横断的研究を実施するとともに、これらの研究項目の枠組みを越えた共同研究を活発に行なって順調に成果を上げることができた。A01 と A02 の連携により、人工錯体の分子集合過程で生じる過渡的化学種を実験的に観測する方法を開発し、それらの遷移ダイナミクスの全容を捉える独創的な分子理論を打ち立てることに成功した。A03 の研究で初めて明らかとなった生体超分子の形成機構とあわせて、人工分子系と生命分子系に底通する自己集合の原理を浮き彫りにすることができた。さらに、アミロイド線維や時計タンパク質などを対象に、非平衡系における生体分子の離合集散過程について、実験と理論の両面から詳細に解き明し、それらの特質を備えた人工分子系の創生も実現している(A01、A02、A03 の連携)。特に、分子集合のエネルギーランドスケープにおける速度論支配と熱力学支配を制御することで超分子形成の時間発展のプログラミングが可能となり、同調的に周期的律動をする人工高分子ゲルや生体分子と人工超分子のハイブリッド化による高次機能の創出にも成果をおさめている。成果発信はもとより、若手育成と国際連携も成功裡に達成することができた。このように、本研究領域の活動を通じて、人工分子系と生命分子系を貫く動的秩序形成機構の普遍性が初めて見出され、人工的な生命システムを創生する指針を導き出すことができた。

研究組織

(総：総括班、支：国際活動支援班、計：総括班及び国際活動支援班以外の計画研究、公：公募研究)

【計画研究】

研究項目	課題番号 研究課題名	代表者氏名	所属機関 部局 職	研究分担者・研究連携者（研究者番号）
X00 総	25102001 生命分子システムにおける動的秩序形成と高次機能発現の研究に関する総括	加藤 晃一	自然科学研究機構・岡崎 統合バイオサイエンスセ ンター・教授	佐藤啓文 上久保裕生 寺嶋正秀 平岡秀一 芳坂貴弘 佐藤宗太 岡本祐幸 稻垣直之 (いずれも計画班員)
Y00 支	15K21708 生命分子システムにおける動的秩序形成と高次機能発現の研究推進のための国際活動支援	加藤 晃一	自然科学研究機構・岡崎 統合バイオサイエンスセ ンター・教授	佐藤啓文 上久保裕生 寺嶋正秀 平岡秀一 芳坂貴弘 佐藤宗太 岡本祐幸 稻垣直之 (いずれも計画班員)
A01 計	25102002 分子集積と秩序形成の分子理論	佐藤 啓文	京都大学・大学院工学研 究科・教授	山本武志 (30397583)
A01 計	25102003 機能を生み出す単位生体分子集団（機能モジュール）の動的秩序の探査	上久保 裕生	奈良先端科学技術大学院 大学・物質創成科学研 究科・教授	片岡幹雄 (30150254)
A01 計	25102004 動的秩序・崩壊のダイナミクスから観る高次機能発現の分子機構解明	寺嶋 正秀	京都大学・大学院理学研 究科・教授	
A02 計	25102005 分子表面の精密デザインに基づく人工系における自己組織化制御	平岡 秀一	東京大学・大学院総合文 化研究科・教授	
A02 計	25102006 生命分子システムの有機化学的拡張による動的秩序の創出	芳坂 貴弘	北陸先端科学技術大学院 大学・マテリアルサイエ ンス研究科・教授	
A02 計	25102007 生体分子系を模倣した動的秩序をもつ人工分子の開発	佐藤 宗太	東京大学・大学院理学系 研究科（理学部）・准教授	
A03 計	25102008 生命分子の動的秩序形成におけるミクロ-マクロ相関の探査と設計原理の探求	加藤 晃一	自然科学研究機構・岡崎 統合バイオサイエンスセ ンター・教授	山口拓実 (60522430) 佐藤匡史 (80532100) 栗原顕輔 (80740919) 矢木真穂 (40608999) 谷中冴子 (80722777)

A03 計	25102009 生体分子集団および人工分子集団の相互作用と大規模構造転換	岡本 祐幸	名古屋大学・大学院理学研究科・教授	
A03 計	25102010 生体分子素子の自己組織化による細胞の動的秩序形成	稻垣 直之	奈良先端科学技術大学院大学・バイオサイエンス研究科・教授	

【公募研究】

平成 26 年度～27 年度

研究項目	課題番号 研究課題名	代表者氏名	所属機関 部局 職	研究分担者・研究連携者（研究者番号）
A01 公	26102501 膜孔形成毒素の動的な作用機構の解明	田中 良和	北海道大学・大学院先端生命科学研究科（研究院）・准教授	
A01 公	26102511 $sHsp$ の動的秩序制御による機能発現の分子機構解明	養王田 正文	東京農工大学・大学院工学研究院・教授	
A01 公	26102512 トポロジー変換可能な新規超分子ポリマーの合成と組織化・機能制御	高田 十志和	東京工業大学・大学院理工学研究科・教授	
A01 公	26102514 ヒトカルシトニンのアミロイド線維形成および阻害の分子機構の解明	内藤 晶	横浜国立大学・大学院工学研究院・教授	川村 出 (20452047)
A01 公	26102515 高速AFMを用いたKaiタンパク質の複合体形成過程のダイナミクス観察	内橋 貴之	金沢大学・数物科学系・教授	
A01 公	26102525（廃止） キュミュラント粗視化力学によるタンパク質動的秩序形成過程の理論研究	重田 育照	筑波大学・大学院数理物質科学研究科・教授	
A01 公	26102526 細菌の細胞分裂ダイナミクスの構造機能相関解析	松村 浩由	立命館大学・生命科学部・教授	
A01 公	26102527 脂質ラフトにおける脂質分子の動的秩序解析	松森 信明	九州大学・大学院理学研究院・教授	木下祥尚 (40529517)
A02 公	26102532（廃止） タンパク質分泌システムの精密探査を可能とする新しい再構成系の構築	塚崎 智也	奈良先端科学技術大学院大学・バイオサイエンス研究科・准教授	
A01 公	26102533 多価カチオンによって媒介される酸性蛋白質間引力の制御と動的秩序構造	秋山 良	九州大学・大学院理学研究院・准教授	
A01 公	26102534 超分子複合系ソフトマターの秩序構造とダイナミクス	安中 雅彦	九州大学・大学院理学研究院・教授	
A01 公	26102538 生細胞内の秩序構造が誘起する蛋白質立体構造の安定性	池谷 鉄兵	首都大学東京・理工学研究科・助教	伊藤隆 (80261147) Peter Güntert (首都大学東京・理工学研究科・客員教授)

A01 公	26102539 量子シミュレーション手法の深化による超分子および生体分子の自己集合機構の解明	立川 仁典	横浜市立大学・大学院生命ナノシステム科学研究所・教授	
A01 公	26102541 人工脂質二重膜におけるドメイン構造の実験的探究	岩田 耕一	学習院大学・理学部・教授	
A02 公	26102542 (廃止) 動的秩序を示すバイオインスパイアードⅡ 電子系-イオン複合体の創製	前田 大光	立命館大学・薬学部・教授	
A01 公	26102544 X線小角散乱と液中高速AFMの相補利用による分子時計の離合集散計測	秋山 修志	分子科学研究所・協奏分子システム研究センター・教授	内橋貴之 (30326300) 向山厚 (80647446)
A02 公	26102502 動的秩序形成を利用した化学反応応答システムの開発	佐田 和己	北海道大学・大学院理学研究院・教授	
A02 公	26102504 コート小胞形成における動的秩序形成メカニズムの解明	二井 勇人	東北大学・大学院農学研究科・准教授	
A02 公	26102507 ATP駆動サイボーグ回転分子モーターの創生	飯野 亮太	自然科学研究機構・岡崎統合バイオサイエンスセンター・教授	
A02 公	26102508 ペプチドフォールディングと超分子錯体によるハイブリッド動的秩序形成	澤田 知久	東京大学・大学院工学系研究科・助教	
A02 公	26102513 人工分子針の細胞膜貫通制御	上野 隆史	東京工業大学・大学院生命理工学研究科・教授	
A02 公	26102517 高分子コロイド分散系における動的秩序の構築	鈴木 大介	信州大学・纖維学部・准教授	
A02 公	26102518 DNAを相互作用素子として細胞様運動する人工システムの構築	神谷 由紀子	名古屋大学・エコトピア科学研究所・講師	
A02 公	26102522 生体膜における曲率形成と膜の形態変化を誘導・制御するペプチドツール	二木 史朗	京都大学・化学研究所・教授	河野健一 (70732874)
A02 公	26102528 リポソーム内膜タンパク質合成系を用いた細胞膜動態の再構成	松浦 友亮	大阪大学・大学院工学研究科・准教授	
A02 公	26102540 メタロペプチドの分子認識化學を活用した動的秩序の多段階創生とタイムプログラミング	三宅 弘之	大阪市立大学・大学院理学研究科・准教授	
A02 公	26102546 感染性の超分子集合体：メカニズムの解明および時間発展の分子論的制御	杉安 和憲	国立研究開発法人物質・材料研究機構・分子機能化學グループ・主任研究員	
A03 公	26102505 アクチン骨格超分子集合体の動的秩序形成機構と細胞遊走、力覚応答における機能	水野 健作	東北大学・大学院生命科学研究科・教授	
A03 公	26102509 細胞内輸送小胞の形成を支える動的秩序の解明	佐藤 健	東京大学・大学院総合文化研究科・准教授	

A03 公	26102510 鞭毛の振動運動発現に至る動的秩序形成	真行寺 千佳子	東京大学・大学院理学系研究科・准教授	
A03 公	26102516 チャネル蛋白質の構造変化に連携した自己組織化動態：チャネル新規機能発現機構の解明	老木 成稔	福井大学・医学部・教授	
A03 公	26102520 タンパク質物性から振動の理論生物学へ	笛井 理生	名古屋大学・大学院工学研究科・教授	
A03 公	26102524 生体分子集合体が形成する動的平衡の中性子小角散乱による研究	杉山 正明	京都大学・原子炉実験所・教授	
A03 公	26102529 生命分子機能を、時空間を制御して解明する設計分子プローブ	菊地 和也	大阪大学・大学院工学研究科・教授	堀雄一郎 (00444563) 蓑島維文 (20600844)
A03 公	26102530 質量分析による蛋白質複合体形成動的メカニズムの解明	内山 進	大阪大学・大学院工学研究科・准教授	石井健太郎 (30748380) 野田勝紀 (大阪大学・大学院工学研究科・特任研究員)
A03 公	26102531 アミロイド伝播核生成相におけるタンパク質分子の集合・秩序化メカニズムの解明	茶谷 紘理	神戸大学・大学院理学研究科・准教授	山本直樹 (90580671)
A03 公	26102543 時計タンパク質の解離集合による時間自動補正メカニズム	寺内 一姫	立命館大学・生命科学部・准教授	
A03 公	26102545 ロタウイルスの感染と増殖における構造秩序形成の解析	村田 和義	生理学研究所・脳機能計測・支援センター・准教授	片山和彦 (60342903)
A03 公	26102547 細胞膜受容体の動的会合体形成と分子認識反応	佐甲 靖志	国立研究開発法人理化学研究所・佐甲細胞情報研究室・主任研究員	
A03 公	26102550 親水性／疎水性溶液界面でのアミロイドベータペプチド凝集機構の理論的研究	奥村 久士	自然科学研究機構・計算科学研究中心・准教授	伊藤暁 (90595381) 森義治 (90646928)

平成 28 年度～平成 29 年度

研究項目	課題番号 研究課題名	代表者氏名	所属機関 部局 職	研究分担者・研究連携者（研究者番号）
A01 公	16H00753 プレフォルディングー 2 型シャペロニンシステムのダイナミクスとフォールディング機構	養王田 正文	東京農工大学・大学院工学研究院・教授	篠原恭介 (20527387)
A01 公	16H00754 ロタキサン連結高分子系超分子における組織化制御	高田 十志和	東京工業大学・大学院理工学研究科・教授	青木大輔 (80736950)
A01 公	16H00756 脂質膜環境でのカルシトニンおよびグルカゴンのアミロイド線維形成分子機構の解明	内藤 晶	横浜国立大学・大学院工学研究院・名誉教授	川村 出 (20452047)

A01 公	16H00758 高速AFM計測によるKaiタンパク質のロバストな概日周期発生機構の解明	内橋 貴之	名古屋大学・理学研究科・教授	
A01 公	16H00773 蛍光標識脂質を用いた脂質膜の動的秩序解析	松森 信明	九州大学・大学院理学研究院・教授	木下祥尚 (40529517)
A01 公	16H00774 多価カチオン媒介型実効引力に注目した蛋白質溶液の相挙動と動的秩序構造制御機構	秋山 良	九州大学・大学院理学研究院・准教授	
A01 公	16H00778 光捕集複合体における自己組織化過程と機能発現の理論的研究	東 雅大	琉球大学・理学部・助教	斎藤真司 (70262847)
A01 公	16H00779 生細胞内の秩序と蛋白質構造安定性の解明に向けた基盤技術	池谷 鉄兵	首都大学東京・理工学研究科・助教	伊藤隆 (80261147) Peter Güntert (首都大学東京・理工学研究科・客員教授)
A01 公	16H00780 超分子・生体分子の自己集合機構の理解のための量子ミュレーション手法の高度化	立川 仁典	横浜市立大学・大学院生命ナノシステム科学研究科・教授	
A01 公	16H00782 人工脂質二重膜におけるドメイン構造の実験的探求	岩田 耕一	学習院大学・理学部・教授	高屋智久 (70466796)
A01 公	16H00783 細胞分裂の階層構造ダイナミクスの解明	松村 浩由	立命館大学・生命科学部・教授	内橋貴之 (30326300) 重田育照 (80376483)
A01 公	16H00785 X線溶液散乱と高速AFMを用いた生物時計因子の動的構造解析	秋山 修志	分子科学研究所・協奏分子システム研究センター・教授	内橋貴之 (30326300) 向山厚 (80647446)
A02 公	16H00755 T4ファージ由来蛋白質針による生体膜透過の動的秩序機構	上野 隆史	東京工業大学・大学院生命理工学研究科・教授	内橋貴之 (30326300) 古田忠臣 (10431834) 川野竜司 (90401702)
A02 公	16H00757 人工的なDNA類似体を基質とする核酸関連酵素反応の解析とその化学的深化	井上 将彦	富山大学・大学院医学薬学教育部(薬学系)・薬学部・教授	阿部肇 (10324055) 千葉順哉 (50436789)
A02 公	16H00760 高分子コロイド分散系における動的秩序の発展	鈴木 大介	信州大学・纖維学部・准教授	
A02 公	16H00761 人工蛋白質ナノブロック自己組織化超分子複合体の構造機能解析と動的秩序系設計構築	新井 亮一	信州大学・纖維学部・准教授	
A02 公	16H00762 人工核酸により発動する細胞様機能の構築	神谷 由紀子	名古屋大学・工学研究科・准教授	
A02 公	16H00763 生体膜における曲率形成と膜の形態変化を誘導・制御するペプチドツール	二木 史朗	京都大学・化学研究所・教授	河野健一 (70732874)
A02 公	16H00767 リポソーム内膜タンパク質発現系の発現ダイナミクス制御技術の確立と応用	松浦 友亮	大阪大学・大学院工学研究科・准教授	

A02 公	16H00775 複製開始蛋白質 Dna Aに 対する制御系の自律的運動シ ステムの創生と動態原理の解 析	片山 勉	九州大学・薬学研究院・教 授	加生和寿 (90726019) 川上広宣 (50403952)
A02 公	16H00777 配位結合を利用した脂質膜上 での動的秩序形成と機能発現	大谷 亮	熊本大学・自然科学研究 科・助教	速水真也 (30321912)
A02 公	16H00781 動的ペプチド金属錯体からなる 超分子集合体の秩序創生と 動的変換プログラミング	三宅 弘之	大阪市立大学・大学院理 学研究科・准教授	
A02 公	16H00787 分化する超分子集合体：エネ ルギーランドスケープに基づく 高次構造と機能の制御	杉安 和憲	国立研究開発法人物質・ 材料研究機構・分子機能 化学グループ・主任研究員	
A02 公	16H00789 糖質加水分解サイボーグリニア 分子モーターの創生	飯野 亮太	自然科学研究機構・岡崎 統合バイオサイエンスセ ンター・教授	
A03 公	16H00748 構造情報に基づいた機能変換 による膜孔形成毒素の会合機 構の解明	田中 良和	東北大学・生命科学研究 科・教授	姚閔 (40311518) 加藤公児 (30452428)
A03 公	16H00749 アクチン骨格超分子集合体の 動的秩序形成機構と細胞機能 発現機構	水野 健作	東北大学・大学院生命科 学研究科・教授	
A03 公	16H00752 鞭毛の振動運動発現に至るダ イニンの自律的複雑性の時空 間的展開	真行寺 千佳子	東京大学・大学院理学系 研究科・准教授	
A03 公	16H00759 チャネル蛋白質のゲーティン グに連動した離合集散動態の 分子機構の解明	老木 成稔	福井大学・医学部・教授	
A03 公	16H00764 P4-ATPaseによる生 体膜のリン脂質動的秩序の形 成機構	申 惠媛	京都大学・薬学研究科・准 教授	
A03 公	16H00765 微生物集団が示す特異な動的 秩序形成機構の解明とその制 御	山本 量一	京都大学・工学系研究科・ 教授	John Jairo Molina (京都大学・工学研究 科・助教)
A03 公	16H00766 巨大タンパク質会合体におけ る動的ネットワークとその機 能解明	杉山 正明	京都大学・原子炉実験所・ 教授	内山進 (90335381) 長田裕也 (60512762)
A03 公	16H00768 生命分子機能を、時空間を制 御して解明する設計分子プロ トタイプ	菊地 和也	大阪大学・大学院工学研 究科・教授	堀雄一郎 (00444563) 蓑島維文 (20600844)
A03 公	16H00770 超分子質量分析による蛋白質 複合体の離合集散過程の解明	内山 進	大阪大学・大学院工学研 究科・准教授	石井健太郎 (30748380) 野田勝紀 (大阪大学・大学院工 学研究科・特任研究 員)
A03 公	16H00772 アミロイド核の出現に先立つ タンパク質初期集合および秩 序化プロセスの解明	茶谷 紘理	神戸大学・大学院理学研 究科・准教授	山本直樹 (90580671)
A03 公	16H00784 生物時計再構成系の動的秩序 探求	寺内 一姫	立命館大学・生命科学部・ 教授	

A03 公	16H00786 無エンベロープウイルスの動的構造秩序形成の解析	村田 和義	生理学研究所・脳機能計測・支援センター・准教授	片山和彦 (60342903)
A03 公	16H00788 細胞膜受容体の動的会合体形成と分子認識反応	佐甲 靖志	国立研究開発法人理化学研究所・佐甲細胞情報研究室・主任研究員	
A03 公	16H00790 アミロイド線維の動的秩序形成過程の全貌の理論的解明	奥村 久士	自然科学研究機構・計算科学研究中心・准教授	伊藤暁 (90595381) 森義治 (90646928) 西澤宏晃 (日本学術振興会特別研究員)

【評価委員】

氏名	所属
増原 宏	台湾国立交通大学 および 奈良先端科学技術大学院大学
菅原 正	神奈川大学 および 東京大学
桑島 邦博	東京大学 および 韓国高等科学院
Christian Griesinger	Max Planck Institute for Biophysical Chemistry

交付決定額（配分額）

	直接経費	間接経費	合計
平成 25 年度	243,400,000	73,020,000	316,420,000
平成 26 年度	259,100,000	77,730,000	336,830,000
平成 27 年度	237,000,000	71,100,000	308,100,000
平成 28 年度	224,600,000	67,380,000	291,980,000
平成 29 年度	220,700,000	66,210,000	286,910,000
平成 30 年度	3,000,000	900,000	3,900,000
総計	1,187,800,000	356,340,000	1,544,140,000

研究発表

(1) 雑誌論文（査読あり。共同研究の場合、重複あり。）

A01：動的秩序の探査

【秋山修志】

1. *A. Mukaiyama, Y. Furuike, J. Abe, E. Yamashita, T. Kondo, *S. Akiyama, "Conformational Rearrangements of the C1 Ring in KaiC measure the Timing of Assembly with KaiB", *Sci. Rep.*, **8**, 8803, (2018), 10.1038/s41598-018-27131-8
2. Y. Furuike, J. Abe, A. Mukaiyama, *S. Akiyama, "Accelerating in Vitro Studies on Circadian Clock Systems Using an Automated Sampling Device", *Biophysics and Physicobiology*, **13**, 235-241, (2016), [10.2142/biophysico.13.0_235](https://doi.org/10.2142/biophysico.13.0_235)
3. *Y. Furukawa, Y. Suzuki, M. Fukuoka, K. Nagasawa, K. Nakagome, H. Shimizu, A. Mukaiyama, S. Akiyama, "A Molecular Mechanism Realizing Sequence-specific Recognition of Nucleic Acids by TDP-43", *Sci. Rep.*, **6**, 20576, (2016), 10.1038/srep20576
4. *Y. Furukawa, I. Anzai, S. Akiyama, M. Imai, Cruz FJC, T. Saio, K. Nagasawa, T. Nomura, K. Ishimori, "Conformational Disorder of the Most Immature Cu,Zn-Superoxide Dismutase Leading to Amyotrophic Lateral Sclerosis", *J. Biol. Chem.*, **291**, 4144-4155, (2016), 10.1074/jbc.M115.683763
5. *秋山 修志, 時間生物学と放射光科学の接点, 放射光, **29**, 56–63, (2016)
6. 阿部 淳, 向山 厚, *秋山 修志, 時計タンパク質 KaiC の「遅さ」が刻み込まれた原子構造 SPring-8/SACLA 利用者情報 **21**, 2-4 (2016)
7. J. Abe, A. Mukaiyama, *S. Akiyama, "Absolute Slowness Encoded in the Circadian Clock Protein KaiC", *SPring-8 Research Frontiers 2015*, 24-25, (2016)
8. J. Abe, T B Hiyama, A. Mukaiyama, S. Son, T. Mori, S. Saito, M. Osako, J. Wolanin, E. Yamashita, T. Kondo, *S. Akiyama, "Atomic-scale Origins of Slowness in the Cyanobacterial Circadian Clock", *Science*, **349**, 312-316 (2015), 10.1126/science.1261040
9. A. Mukaiyama, M. Osako, T. Hikima, T. Kondo, *S. Akiyama, "A Protocol for Preparing Nucleotide-free KaiC Monomer," *BIOPHYSICS* **11**, 79, (2015), [10.2142/biophysics.11.79](https://doi.org/10.2142/biophysics.11.79)
10. 向山 厚, 阿部 淳, 孫 世永, *秋山 修志, タンパク質の化学反応が細胞内の時を計る 実験医学 **33**, 3119-3122 (2015)

【秋山良】

1. A. Suematsu, T. Sawayama, *R. Akiyama, "Effective Potential between Negatively Charged Patches on Acidic Proteins Immersed in Various Electrolyte Solutions", *J. Chem. Phys.*, **149**, 074105, (2018), [10.1063/1.5038912](https://doi.org/10.1063/1.5038912)
2. *Y. Nakamura, A. Yoshimori, R. Akiyama, T. Yamaguchi "Stick Boundary Condition at Large Hard Sphere Arising from Effective Attraction in Binary Hard-sphere Mixtures", *J. Chem. Phys.*, **148**, 124502 (2018), [10.1063/1.5025202](https://doi.org/10.1063/1.5025202)
3. *K. Tokunaga, *R. Akiyama "Basic Cell Size Dependence of Displacement for a Solvation Motor in a Lennard-Jones Solvent", *J. Comp. Chem. Jpn.*, **17**, 80–84, (2018), [10.2477/jccj.2018-0004](https://doi.org/10.2477/jccj.2018-0004)
4. *H. Okumura, M. Higashi, Y. Yoshida, H. Sato, R. Akiyama, "Theoretical Approaches for Dynamical Ordering of Biomolecular Systems", *Biochim. Biophys. Acta, (BBA), Gen. Subj.*, **1862**, 212-228, (2018), [10.1016/j.bbagen.2017.10.001](https://doi.org/10.1016/j.bbagen.2017.10.001)
5. *A. Chiba, M. Inui, Y. Kajihara, K. Fuchizaki, R. Akiyama, "Isotactic Poly(4-methyl-1-pentene) Melt as a Porous Liquid: Reduction of Compressibility due to Penetration of Pressure Medium", *J. Chem. Phys.*, **146**, 194503-1-5, (2017), [10.1063/1.4983508](https://doi.org/10.1063/1.4983508)
6. *M. Hishida, Y. Nomura, R. Akiyama, Y. Yamamura, *K. Saito, "Electrostatic Double-layer Interaction between Stacked Charged Bilayers" *Phys. Rev. E*, **96**, 040601(R), (2017), [10.1103/PhysRevE.96.040601](https://doi.org/10.1103/PhysRevE.96.040601)
7. *A. Suematsu, A. Yoshimori, R. Akiyama, "Effects of Interactions between Depletants in Phase Diagrams of Binary Hard-sphere Systems", *EPL*, **116**, 38004-1-7, (2016), [10.1209/0295-5075/116/38004](https://doi.org/10.1209/0295-5075/116/38004)
8. *Y. Nakamura, A. Yoshimori, R. Akiyama, "Effects of Solvation Structure on Diffusion of Large Particle in Binary Mixture Studied by Perturbation Theory", *J. Mol. Liq.*, **200**, 85-88, (2014), [10.1016/j.molliq.2014.06.021](https://doi.org/10.1016/j.molliq.2014.06.021)

9. S. Fujihara, *R. Akiyama, "Attractive Interaction between Macroanions Mediated by Multivalent Cations in Biological Fluids", *J. Mol. Liq.*, **200**, 89-94, (2014), [10.1016/j.molliq.2014.06.022](https://doi.org/10.1016/j.molliq.2014.06.022)
10. *R. Akiyama, T. Yamashita, S. Fujihara, "Hidden Peak of Radial Distribution Function and Effective Attraction between Like-charged Proteins caused by Translational Motion of Solvent Molecules", *J. Mol. Liq.*, **200**, 72-76, (2014), [10.1016/j.molliq.2014.06.004](https://doi.org/10.1016/j.molliq.2014.06.004)
11. Y. Kawabata, *R. Akiyama, "Choice of the Center of Water Molecules in Calculations of Partial Molar Volume of Single Ions Immersed in Water: A Molecular Simulation Study", *J. Mol. Liq.*, **200**, 67-71, (2014), [10.1016/j.molliq.2014.05.022](https://doi.org/10.1016/j.molliq.2014.05.022)
12. Y. Nakamura, *A. Yoshimori, R. Akiyama, "Perturbation Theory of Large-Particle Diffusion in a Binary Solvent Mixture", *J. Phys. Soc. Jpn.*, **83**, 064601-1-9, (2014), <http://dx.doi.org/10.7566/JPSJ.83.064601>.

【安中雅彦】

1. M. Yanagisawa, Y. Yamashita, S. Mukai, M. Annaka, M. Tokita "Phase separation in binary polymer solution: Gelatin/Poly (ethylene glycol) system", *Journal of Molecular Liquid*, **200**, 2-6 (2014), [10.1016/j.molliq.2013.12.035](https://doi.org/10.1016/j.molliq.2013.12.035)
2. A. Shundo, K. Hori, D. P. Penaloza Jr. K. Yoshihiro, M. Annaka, K. Tanaka, "Nonsolvents-induced swelling of poly(methylmethacrylate) nanoparticles", *Physical Chemistry Chemical Physics*, **15**, 16574-16578, (2014), [10.1039/c3cp52673a](https://doi.org/10.1039/c3cp52673a)

【池谷鉄兵】

1. T. Ikeya,*Y. Ito, "Protein NMR Structure Refinement Based on Bayesian Inference for Dynamical Ordering of Biomacromolecules.", *J. Comput. Chem. Jpn.*, **7** (1), 65-75, (2018), [10.2477/jccj.2018-0009](https://doi.org/10.2477/jccj.2018-0009)
2. *T. Ikeya, D. Ban, D. Lee, Y. Ito, K. Kato, *C. Griesinger, "Solution NMR Views of Dynamical Ordering of Biomacromolecules", *Biochim. Biophys. Acta, (BBA), Gen. Subj.*, **1862**, 287-306, (2017), [10.1016/j.bbagen.2017.08.020](https://doi.org/10.1016/j.bbagen.2017.08.020)
3. T. Ikeya, T. Hanashima, S. Hosoya, M. Shimazaki, S. Ikeda, M. Mishima, P. Güntert,*Y. Ito, "In-cell Structure Determination of Proteins at Near-physiological Concentration", *Sci. Rep.*, **6**, 38312, (2016), [10.1038/srep38312](https://doi.org/10.1038/srep38312)
4. Y. Hikone, G. Hirai, M. Mishima, K. Inomata, T. Ikeya, S. Arai, M. Shirakawa, M. Sodeoka, *Y. Ito, "A New Carbamidemethyl-linked Lanthanide Chelating Tag for PCS NMR Spectroscopy of Proteins in Living HeLa cells" *J. Biomol. NMR*, **66**, 99-110, (2016), [10.1007/s10858-016-0059-4](https://doi.org/10.1007/s10858-016-0059-4)
5. Y. Nishida, T. Ikeya, T. Mikawa, J. Inoue, Y. Ito, Y. Shintani, R. Masui, S. Kuramitsu, *S. Takashima, "A Specific Single-stranded DNA Induces a Distinct Conformational Change in the Nucleoid-associated Protein HU", *BB Reports*, **8**, 318–324, (2016), [10.1016/j.bbrep.2016.09.014](https://doi.org/10.1016/j.bbrep.2016.09.014)
6. *T. Ikeya, S. Ikeda, T. Kigawa, Y. Ito, P. Güntert*, "Protein NMR Structure Refinement based on Bayesian Inference," *J. Phys. Conf. Ser.*, **699**, 012005, 1-14, (2016)
7. 葛西卓磨, 池谷鉄兵, 木川隆則*, 「生命分子の NMR 計測・解析への応用」, 電子情報通信学会, Vol.99, No.5(2016), pp.439-443
8. Y. Shigemitsu, T. Ikeya, A. Yamamoto, Y. Tsuchie, M. Mishima, B. O. Smith, P. Güntert and *Y. Ito... "Evaluation of the reliability of the maximum entropy method for reconstructing 3D and 4D NOESY-type NMR spectra of proteins", *Biochem. Biophys. Res. Commun.*, **457**(2), 200-205, (2015), [10.1016/j.bbrc.2014.12.088](https://doi.org/10.1016/j.bbrc.2014.12.088)
9. Y. J. Lin, T. Ikeya, D.K. Kirchner, P. Güntert, "Influence of Incomplete NOESY Peaks of the Interface Residues on Structure Determinations of Homodimeric Proteins", *J. Chin. Chem. Soc.* **61**, 1297-1306, (2014), [10.1002/jccs.201400095](https://doi.org/10.1002/jccs.201400095)
10. E. Schmidt, T. Ikeya, M. Takeda, F. Löhr, L. Buchner, Y. Ito, *M. Kainosh, *P. Güntert, "Automated Resonance Assignment of the 21 kDa Stereo-array Isotope Labeled Thiodisulfide Oxidoreductase DsbA", *J. Magn. Reson.*, **249**, 88-93, (2014), [10.1016/j.jmr.2014.10.005](https://doi.org/10.1016/j.jmr.2014.10.005)

【岩田耕一】

1. T. Takaya, T. Oda, Y. Shibasaki, Y. Hayashi, H. Shimomoto, *E. Ihara, Y. Ishibashi, *T. Asahi, *K. Iwata, "Excited-State Dynamics of Pyrene Incorporated into Poly(substituted methylene)s: Effects of Dense Packing of Pyrenes on Excimer Formation", *Macromolecules*, **51**, 5430-5439, (2018) [10.1021/acs.macromol.8b01060](https://doi.org/10.1021/acs.macromol.8b01060).
2. W. Xiong, L. Du, K. C. Lo, H. Shi, T. Takaya, K. Iwata, *W. K. Chan, *D. L. Phillips, "Control of Electron Flow Direction in Photoexcited Cycloplatinated Complex Containing Conjugated Polymer-

- Single Walled Carbon Nanotube Hybrids”, *J. Phys. Chem. Lett.*, **9**, 3819-3824, (2018), 10.1021/acs.jpclett.8b01713
3. *K. Iwata, *M. Terazima, *H. Masuhara, “Novel Physical Chemistry Approaches in Biophysical Researches with Advanced Application of Lasers: Detection and Manipulation”, *Biochim. Biophys. Acta, (BBA), Gen. Subj.*, **1862**, 335-357, (2018), [10.1016/j.bbagen.2017.11.003](https://doi.org/10.1016/j.bbagen.2017.11.003)
 4. *T. Takaya, M. Anan, *K. Iwata, “Vibrational Relaxation Dynamics of β -carotene and its Derivatives with Substituents on Terminal Rings in Electronically Excited States as Studied by Femtosecond Time-resolved Stimulated Raman Spectroscopy in near-IR”, *Phys. Chem. Chem. Phys.*, **20**, 3320-3327, (2018), [10.1039/C7CP06343A](https://doi.org/10.1039/C7CP06343A)
 5. F. Lu, N. Kitamura, T. Takaya, K. Iwata, *T. Nakanishi, Y. Kurashige, “Experimental and Theoretical Investigation of Fluorescence Solvatochromism of Dialkoxyphenyl-pyrene Molecules”, *Phys. Chem. Chem. Phys.*, **20**, 3258-3264, (2018), [10.1039/C7CP06811E](https://doi.org/10.1039/C7CP06811E)
 6. B. Narayan, K. Nagura, T. Takaya, K. Iwata, A. Shinohara, H. Shinmori, H. Wang, Q. Li, X. Sun, H. Li, S. Ishihara, *T. Nakanishi, “Regioisomeric Effect on Photo-physical Properties of Alkylated-naphthalene Liquids”, *Phys. Chem. Chem. Phys.*, **20**, 2970-2975, (2018), [10.1039/C7CP05584F](https://doi.org/10.1039/C7CP05584F)
 7. *R. Ohtani, T. Tokita, T. Takaya, *K. Iwata, M. Kinoshita, N. Matsumori, M. Nakamura, L. F. Lindoy, *S. Hayami, “Morphology Controls of Hybrid Liposomes Using Metal Complex Lipids and Viscosities for Photo-chemical Reaction in Hydrophobic Fields”, *Chem. Comm.*, **53**, 13249-13252, (2017), [10.1039/C7CC07944C](https://doi.org/10.1039/C7CC07944C)
 8. N. Dwivedi, A. Verma, T. Takaya, K. Iwata, *S. Saha, *S. S. Sunkari, “NIR Luminescent Heterodinuclear [ZnII LnIII] Complexes: Synthesis, Crystal Structures and Photophysical Properties”, *J. Luminesc.*, **192**, 156-165, (2017), [10.1016/j.jlumin.2017.06.045](https://doi.org/10.1016/j.jlumin.2017.06.045)
 9. F. Lu, *T. Takaya, K. Iwata, I. Kawamura, A. Saeki, M. Ishii, K. Nagura, *T. Nakanishi, “A Guide to Design Functional Molecular Liquids with Tailorable Properties using Pyrene-Fluorescence as a Probe”, *Sci. Rep.*, **7**, 3416, (2017), 10.1038/s41598-017-03584-1
 10. *T. Takaya, K. Iwata, “Development of a Femtosecond Time-resolved Near-IR Multiplex Stimulated Raman Spectrometer in Resonance with Transitions in the 900–1550 nm Region”, *Analyst*, **141**, 4283-4292, (2016), [10.1039/c6an01051b](https://doi.org/10.1039/c6an01051b)
 11. S. Okino, T. Takaya, *K. Iwata, “Femtosecond Time-resolved near-infrared Spectroscopy of Oligothiophenes and Polythiophene: Energy Location and Effective Conjugation Length of their Low-lying Excited States”, *Chem. Lett.*, **44**, 1059-1061, (2015), [10.1246/cl.150330](https://doi.org/10.1246/cl.150330)
 12. A. Z. Samuel, S. Yabumoto, K. Kawamura, *K. Iwata, “Rapid Microstructure Characterization of Polymer Thin Films with 2D-Array Multifocus Raman Microspectroscopy”, *Analyst*, **140**, 1847 – 1851, (2015), [10.1039/C4AN01983K](https://doi.org/10.1039/C4AN01983K)
 13. *T. Takaya, K. Iwata, “Relaxation Mechanism of β -Carotene from S_2 ($^1B_u^+$) State to S_1 ($^2A_g^-$) State: Femtosecond Time-Resolved Near-IR Absorption and Stimulated Resonance Raman Studies in 900–1550 nm Region”, *J. Phys. Chem. A*, **118**(23), 4071–4078, (2014), [10.1021/jp504272h](https://doi.org/10.1021/jp504272h)
 14. Y. Nojima, *K. Iwata, “Viscosity Heterogeneity Inside Lipid Bilayers of Single-Component Phosphatidylcholine Liposomes Observed with Picosecond Time-Resolved Fluorescence Spectroscopy”, *J. Phys. Chem. B*, **118**(29), 8631-8641, (2014), [10.1021/jp503921e](https://doi.org/10.1021/jp503921e)

【内橋貴之】

1. S. Matsui, Y. Nishizawa, *T. Uchihashi, *D. Suzuki, “Monitoring Thermo-responsive Morphological Changes in Individual Hydrogel Microspheres”, *ACS Omega*, in press, (2018)
2. K. Oohora*, N. Fujimaki, R. Kajihara, H. Watanabe, *T. Uchihashi, *T. Hayashi, “Supramolecular Hemoprotein Assembly with a Periodic Structure Showing Heme–Heme Exciton Coupling”, *J. Am. Chem. Soc.*, **140** (32), 10145–10148(2018), [10.1021/jacs.8b06690](https://doi.org/10.1021/jacs.8b06690)
3. T. Haruyama, Y. Sugano, N. Kodera, T. Uchihashi, T. Ando, Y. Tanaka, *H. Konno, *T. Tsukazaki, “Single-unit Imaging of Membrane Protein-embedded Nanodiscs from Two Oriented Sides by High-speed Atomic Force Microscopy”, *Structure*, (2018), in press
4. T. Mori, S. Sugiyama, M. Byme, C. H. Johnson, *T. Uchihashi, *T. Ando, “Revealing Circadian Mechanisms of Integration and Resilience by Visualizing clock Proteins Working in Real Time”, *Nat. Commun.*, **9**, 3245, (2018), [10.1038/s41467-018-05438-4](https://doi.org/10.1038/s41467-018-05438-4)
5. T. Uchihashi, *Y. Watanabe, Y. Nakazaki, T. Yamasaki, H. Watanabe, T. Maruno, K. Ishii, S. Uchiyama, C. Song, K. Murata, *R. Jino, *T. Ando, “Dynamic Structural States of ClpB Involved in its

- Disaggregation Function”, *Nat. Commun.*, **9**, 2147, (2018), 10.1038/s41467-018-04587-w
6. M. Shibata, K. Inoue, K. Ikeda, M. Konno, M. Singh, C. Kataoka, R. Abe-Yoshizumi, *H. Kandorix, *T. Uchihashi, “Oligomeric States of Microbial Rhodopsins Determined by High-speed Atomic Force Microscopy and Circular Dichroic Spectroscopy”, *Sci. Rep.*, **8**, 8262, (2018), 110.1038/s41598-018-26606-y
 7. T. Umakoshi, H. Udaka, T. Uchihashi, T. Ando, M. Suzuki, *T. Fukuda, “Quantum-dot Antibody Conjugation Visualized at the Single-molecule Scale with High-speed Atomic Force Microscopy”, *Colloids Surf., B*, **167**, 267-274, (2018), 10.1016/j.colsurfb.2018.04.015
 8. *内橋貴之, “高速原子間力顕微鏡によるタンパク質の動態可視化と画像解析”, *J. Comp. Chem.*, **17**, 20-30, (2018), 10.2477/jccj.2018-0001
 9. M. Hosoyamada, *N. Yanai, K. Okumura, T. Uchihashi, N. Kimizuka, “Translating MOF Chemistry into Supramolecular Chemistry: Soluble Coordination Nanofibers Showing Efficient Photon Upconversion”, *ChemComm.*, **54**, 6828-6831, (2018), 10.1039/C8CC01594E
 10. N. Terahara, Y. Inoue, N. Kodera, Y. V. Morimoto, T. Uchihashi, K. Imada, T. Ando, K. Namba, *T. Minamino, “Insight into Structural Remodeling of the FlhA Ring Responsible for Bacterial Flagellar type III Protein Export”, *Sci. Adv.*, **4**, 7054, (2018), 10.1126/sciadv.ao7054
 11. A. Oda, S. Nagao, M. Yamanaka, H. Watanabe, T. Uchihashi, I. Ueda, N. Shibata, Y. Higuchi, *S. Hirota, “Construction of a Triangle-Shaped Trimer and a Tetrahedral Structure Using an α -Helix-Inserted Circular Permutant of Cytochrome c_{555} ”, *Chem. Asian. J.*, **13**, 964-967, (2018), 10.1002/asia.201800252
 12. H. Tsukamoto, M. Higashi, H. Motoki, H. Watanabe, C. Ganser, K. Nakajo, Y. Kubo, T. Uchihashi, *Y. Furutani, “Structural Properties Determining Low K⁺ Affinity of the Selectivity Filter in the TWIK1 K⁺ Channel”, *J. Biol. Chem.*, **293**, 6969-6984, (2018), 10.1074/jbc.RA118.001817
 13. T. Maruno, H. Watanabe, T. Uchihashi, S. Adachi, K. Arai, T. Sawaguchi, *S. Uchiyama, “Sweeping of Adsorbed Therapeutic Proteins on Prefillable Syringe Enhances Subvisible Particles Generation”, *J. Pharm. Sci.*, **107**, 1521-1529 (2018), 10.1016/j.xphs.2018.01.021
 14. T. Haruyama, T. Uchihashi, Y. Yamada, N. Kodera, T. Ando, *H. Konno, “Negatively Charged Lipids are Essential for Functional and Structural Switch of Human 2-Cys Peroxiredoxin II”, *J. Mol. Biol.*, **430**, 602-610, (2018), 10.1016/j.jmb.2017.12.020
 15. T. Takeda, T. Kozai, H. Yang, D. Ishikuro, K. Seyama, Y. Kumagai, T. Abe, H. Yamada, T. Uchihashi, *T. Ando, *K. Takei, “Dynamic Clustering of Dynamin-amphiphysin Helices Regulates Membrane Constriction and Fission Coupled with GTP Hydrolysis”, *e-Life*, **7**, e3024, (2018), 10.7554/eLife.30246
 16. M. Yagi-Utsumi, A. Sikdar, T. Kozai, R. Inoue, M. Sugiyama, T. Uchihashi, T. Satoh, *K. Kato, “Conversion of Functionally Undefined Homopentameric Protein PbaA into a Proteasome Activator by Mutational Modification of its C-terminal Segment Conformation”, *Protein Eng. Des. Sel.*, **31**, 29-36, (2018), 10.1093/protein/gzx066
 17. A. Nakamura, T. Tasaki, Y. Okuni, C. Song, K. Murata, T. Kozai, M. Hara, H. Sugimoto, K. Suzuki, T. Watanabe, T. Uchihashi, H. Noji, *R. Iino, “Rate Constants, Processivity, and Productive Binding Ratio of Chitinase A Revealed by Single-molecule Analysis”, *Phys. Chem. Chem. Phys.*, **20**, 3010-3018, (2018), 10.1039/c7cp04606e
 18. T. Kozai, T. Sekiguchi, T. Satoh, H. Yagi, *K. Kato, *T. Uchihashi, “Two-step Process for Disassembly Mechanism of Proteasome α_7 Homo-tetradecamer by α_6 Revealed by High-speed Atomic Force Microscopy”, *Sci. Rep.* **7**, 15373, (2017), 10.1038/s41598-017-15708-8
 19. N. Terahara, N. Kodera, T. Uchihashi, T. Ando, *K. Namba, *T. Minamino, “Na⁺-induced Structural Transition of MotPS for Stator Assembly of Bacillus Flagellar Motor”, *Sci. Adv.*, **3**, eaao4119, (2017), 10.1126/sciadv.aao4119
 20. *M. Shibata, H. Watanabe, T. Uchihashi, T. Ando, R. Yasuda, “High-speed Atomic Force Microscopy Imaging of Live Mammalian Cells”, *BPPB*, **14**, 127-135, (2017), 10.2142/biophysico.14.0_127
 21. *M. Shibata, *H. Nishimasu, N. Kodera, S. Hirano, T. Ando, T. Uchihashi, *O. Nureki, “Real-space and Real-Time Dynamics of CRISPR-Cas9 Visualized by High-speed Atomic Force Microscopy”, *Nat. Commun.*, **8**, 1430, (2017), 10.1038/s41467-017-01466-8
 22. *T. Satoh, C. Song, T. Zhu, T. Toshimori, K. Murata, Y. Hayashi, H. Kamikubo, T. Uchihashi, *K. Kato, “Visualisation of a Flexible Modular Structure of the ER Folding-sensor Enzyme UGGT”, *Sci. Rep.*, **7**, 12142, (2017), 10.1038/s41598-017-12283-w

23. *T. Uchihashi, S. Scheuring, "Review: Applications of High-speed Atomic Force Microscopy to Real-time Visualization of Dynamic Biomolecular Processes", *Biochim. Biophys. Acta, (BBA), Gen. Subj.*, **1862**, 229-2410, (2017), [10.1016/j.bbagen.2017.07.010](https://doi.org/10.1016/j.bbagen.2017.07.010)
24. H. Harada, *A. Onoda, *T. Uchihashi, H. Watanabe, N. Sunagawa, M. Samejima, *K. Igarashi, T. Hayashi, "Interdomain Flip-flop Motion Visualized in Flavocytochrome Cellobiose Dehydrogenase Using High-speed Atomic Force Microscopy during Catalysis", *Chem. Sci.*, **8**, 6561-6565, (2017), [10.1039/c7sc01672g](https://doi.org/10.1039/c7sc01672g)
25. S. Matsui, T. Kureha, S. Hiroshige, M. Shibata, *T. Uchihashi, *D. Suzuki, "Fast Adsorption of Soft Hydrogel Microspheres on Solid Surfaces in Aqueous Solution", *Angew. Chem. Int. Ed. (Communication)* **56**, 12146-12149, (2017), [10.1002/anie.201705808](https://doi.org/10.1002/anie.201705808)
26. J. J. Keya, D. Inoue, Y. Suzuki, T. Kozai, D. Ishikuro, N. Kodera, T. Uchihashi, A. Md. R. Kabir, M. Endo, K. Sada, A. Kakugo, "High-Resolution Imaging of a Single Gliding Protofilament of Tubulins by HS-AFM", *Sci. Rep.*, **7**, 6166, (2017), [10.1038/s41598-017-06249-1](https://doi.org/10.1038/s41598-017-06249-1)
27. M. Mohamed, A. Kobayashi, A. Taoka, T. Watanabe-Nakayama, Y. Kikuchi, M. Hazawa, T. Minamoto, Y. Fukumori, N. Kodera, T. Uchihashi, T. Ando, *R. Wong, "High-Speed Atomic Force Microscopy Reveals Loss of Nuclear Pore Resilience as a Dying Code in Colorectal Cancer Cells", *ACS Nano*, **11**, 5567-5578, (2017), [10.1021/acsnano.7b00906](https://doi.org/10.1021/acsnano.7b00906)
28. A. Sumino, T. Uchihashi, *S. Oiki, "Oriented Reconstitution of the Full-Length KcsA Potassium Channel in a Lipid Bilayer for AFM Imaging", *J. Phys. Chem. Lett.*, **8**, 785-793, (2017), [10.1021/acs.jpclett.6b03058](https://doi.org/10.1021/acs.jpclett.6b03058)
29. K. Inoue, S. Ito, Y. Kato, Y. Nomura, M. Shibata, T. Uchihashi, S. P. Tsunoda, *H. Kandori, "A Natural Light-driven Inward Proton Pump", *Nat. Commun.* **8**, 13415, (2016), [10.1038/ncomms13415](https://doi.org/10.1038/ncomms13415)
30. T. Uchihashi, H. Watanabe, S. Fukuda, M. Shibata, *T. Ando, "Functional Extension of High-speed Atomic Force Microscopy", *Ultramicroscopy*, **160**, 182-196 (2016), [10.1016/j.ultramic.2015.10.017](https://doi.org/10.1016/j.ultramic.2015.10.017)
31. 内橋貴之、「光と生命の事典」：第5章 「光による生命現象の計測」 177節 高速原子間力顕微鏡, 真嶋哲郎, 七田芳則, 飯野盛利, 藤堂剛 (編), 朝倉書店, pp1-224, (2016), ISBN: 978-4-254-17161-7.
32. W. Sriwimol, A. Aroonkesorn, S. Sakdee, C. Kanchanawarin, T. Uchihashi, T. Ando, *C. Angsuthanasombat, "Potential Pre-pore Trimer Formation by the Bacillus Thuringiensis Mosquito-specific Toxin: Molecular Insights into a Critical Prerequisite of Membrane-bound Monomers", *J. Biol. Chem.*, **290** (34), 20793-20803, (2015), [10.1074/jbc.M114.627554](https://doi.org/10.1074/jbc.M114.627554)
33. S. Fukuda, T. Uchihashi, *T. Ando, "Method of Mechanical Holding of Cantilever Chip for Tip-scan High-speed Atomic Force Microscopy", *Rev. Sci. Instrum.*, **86**, 063703, (2015), [10.1063/1.4922381](https://doi.org/10.1063/1.4922381)
34. M. Imamura, T. Uchihashi, T. Ando, A. Leifert, U. Simon, *A. D. Maly, *J. G. Heddle, "Probing Structural Dynamics of an Artificial Protein Cage Using High-Speed Atomic Force Microscopy", *Nano Letters*, **15**(2), 1331-1335, (2015), [10.1021/nl5045617](https://doi.org/10.1021/nl5045617)
35. K. Takeda, T. Uchihashi, H. Watanabe, T. Ishida, *K. Igarashi, N. Nakamura, H. Ohno, "Real-time Dynamic Adsorption Processes of Cytochrome c on an Electrode Observed through Electrochemical High-speed Atomic Force Microscopy", *PLoS ONE*, **10**(2), e0116685, (2015), [10.1371/journal.pone.0116685](https://doi.org/10.1371/journal.pone.0116685)
36. *M. Shibata, T. Uchihashi, T. Ando, *R. Yasuda, Long-tip High-speed Atomic Force Microscopy for Nanometer-scale Imaging in Live Cells", *Sci. Rep.*, **5**, 8724, (2015), [10.1038/srep08724](https://doi.org/10.1038/srep08724)
37. 高速原子間力顕微鏡による生体試料のダイナミクス観察 Y. Shibafuji, A. Nakamura, T. Uchihashi, N. Sugimoto, S. Fukuda, H. Watanabe, M. Samejima, T. Ando, H. Noji, A. Koivula, K. Igarashi, *R. Iino, "Single-molecule Imaging Analysis of Elementary Reaction Steps of Trichoderma Reesei cellobiohydrolase I (Cel7A) Hydrolyzing Crystalline Cellulose", *J. Biol. Chem.*, **289**, 14056-14065, (2014), [10.1074/jbc.M113.546085](https://doi.org/10.1074/jbc.M113.546085)
38. ***K. Igarashi, ** T. Uchihashi, T. Uchiyama, H. Sugimoto, M. Wada, K. Suzuki, S. Sakuda, T. Ando, T. Watanabe, and M. Samejima, "Two-way Traffic of Glycoside Hydrolase Family 18 Processive Chitinases on Crystalline Chitin", *Nat. Commun.* **5**, 3975, (2014), [10.1038/ncomms4975](https://doi.org/10.1038/ncomms4975) ** Co-first authors
39. A. Nakamura, H. Watanabe, T. Ishida, T. Uchihashi, M. Wada, T. Ando, K. Igarashi, *M. Samejima, "Trade-off between Processivity and Hydrolytic Velocity of Cellobiohydrolases at the Surface of Crystalline Cellulose", *J. Am. Chem. Soc.* **136**, 4584-4592, (2014), [10.1021/ja4119994](https://doi.org/10.1021/ja4119994)
40. 杉本華幸, 五十嵐圭日子, 内橋貴之, 鈴木一史, *渡邊剛志、「キチナーゼによる結晶性キチンのプロセッサ（連続的）な分解機構の解明」、日本応用糖質化学会誌 **4** (2), 107-112 (2014)

41. 内橋貴之、膜誌 **39**(5), 322-328 (2014)
42. 古寺哲幸, 内橋貴之, 安藤敏夫、「高速原子間力顕微鏡による生体分子のナノ動体撮影」
日本物理学会誌 **69**(7), 459-464 (2014)
43. 内橋貴之, 飯野亮太, 安藤敏夫, 野地博行、「高速 AFM による F₁-ATPase 分子回転の直接可視化」、生化学 **86**(2), 127-136 (2014)

【上久保裕生】

1. 上久保裕生、「連続滴定 X 線溶液散乱測定を志向した μ 流路型自動サンプリングシステムの開発」、*Journal of Computer Chemistry Japan*, **17** 卷, 1 号, 57-64, (2018), org/10.2477/jccj.2018-0008
2. M. Yamaguchi, E. Ohta, T.Muto, T. Watanabe, T. Hohsaka, Y. Yamazaki, H. Kamikubo, *M. Kataoka, “Statistical Description of the Denatured Structure of a Single Protein, Staphylococcal Nuclease, by FRET Analysis” *Biophys Rev.*, **10**(2), 145-152, (2018), [10.1007/s12551-017-0334-y](https://doi.org/10.1007/s12551-017-0334-y)
3. R. Kawatani, Y. Nishiyama, H. Kamikubo, K.Kakiuchi, H. Ajiro, “Aggregation Control by Multi-stimuli-Responsive Poly (N-vinylamide) Derivatives in Aqueous System”, *Nanoscale Res. Lett.*, **12**, 461, (2017), [10.1186/s11671-017-2221-7](https://doi.org/10.1186/s11671-017-2221-7)
4. M. Nawata, H. Tsutsumi, Y Kobayashi, S. Unzai, S. Mine, T. Nakamura, K. Uegaki, H. Kamikubo, M. Kataoka, *D. Hamada, “Heat-induced Native Dimerization Prevents Amyloid Formation by Variable Domain from Immunoglobulin Light-chain REI”, *FEBS J.* **284**(18), 3114-3127, (2017), [10.1111/febs.14181](https://doi.org/10.1111/febs.14181)
5. K. Yonezawa, N. Shimizu, K. Kurihara, Y. Yamazaki, *H. Kamikubo, M. Kataoka, “Neutron Crystallography of Photoactive Yellow Protein Reveals Unusual Protonation State of Arg52 in the Crystal”, *Sci. Rep.*, **7**, 9361, (2017), [10.1038/s41598-017-09718-9](https://doi.org/10.1038/s41598-017-09718-9)
6. T. Satoh, C.Song, T. Zhu, T. Toshimori, K. Murata, Y. Hayashi, H. Kamikubo, T. Uchihashi, *K.Kato, “Visualisation of a Flexible Modular Structure of the ER Folding-sensor Enzyme UGGT”, *Sci. Reps.*, **7**, 12142, (2017), [10.1038/s41598-017-12283-w](https://doi.org/10.1038/s41598-017-12283-w)
7. P. Bernadó, N. Shimizu, G. Zaccai, *H. Kamikubo, *M. Sugiyama, “Solution Scattering Approaches to Dynamical Ordering in Biomolecular Systems”, *Biochim. Biophys. Acta, (BBA), Gen. Subj.*, **1862**, 253-274, (2017), [10.1016/j.bbagen.2017.10.015](https://doi.org/10.1016/j.bbagen.2017.10.015)
8. H. Kuramochi , S. Takeuchi ,K. Yonezawa , H. Kamikubo, M. Kataoka, *T.Tahara, “Probing the Early stages of Photoreception in Photoactive Yellow Protein with Ultrafast Time-domain Raman Spectroscopy”, *Nature Chemistry*, **9**, 660-666, (2017), [10.1038/nchem.2717](https://doi.org/10.1038/nchem.2717)
9. L. Zhang, H. Kondo, H. Kamikubo, M. Kataoka, *W. Sakamoto, “VIPP1 has a Disordered C-terminal Tail Necessary for Protecting Photosynthetic Membranes against Stress in Arabidopsis”, *Plant Physiology*, **171**, 1983-1995, (2016), [10.1104/pp.16.00532](https://doi.org/10.1104/pp.16.00532)
10. J. K. Endow, A. G. Rocha , A. J. Baldwin , R. L. Roston , T. Yamaguchi, H. Kamikubo, *K. Inoue, “Polyglycine Acts as a Rejection Signal for Protein Transport at the Chloroplast Envelope”, *PLOS ONE*, **11**(12), e0167802, (2016), [10.1371/journal.pone.0167802](https://doi.org/10.1371/journal.pone.0167802)
11. Y. Yoshimura, N. A. Oktaviani, K. Yonezawa, H. Kamikubo, *F. A. A. Mulder, “Unambiguous Determination of the Ionization State of a Photoactive Protein Active Site Arginine in Solution by NMR Spectroscopy”, *Angewandte Chemie*, **56**, 239-242, (2016), [10.1002/anie.201609605](https://doi.org/10.1002/anie.201609605)
12. Y. Kanematsu, H. Kamikubo, M. Kataoka, M. Tachikawa, “Vibrational Analysis on the Revised Potential Energy Curve of the Low-barrier Hydrogen Bond in Photoactive Yellow Protein”, *Comput. Struct. Biotechnol. J.*, **14**, 16-19, (2015), [10.1016/j.csbj.2015.10.003](https://doi.org/10.1016/j.csbj.2015.10.003)
13. S. Nagao, M. Ueda, H. Osuka, H. Komori, H. Kamikubo, M. Kataoka Y. Higuchi, *S. Hirota, “Domain-Swapped Dimer of Pseudomonas Aeruginosa Cytochrome c551: Structural Insights into Domain Wapping of Cytochrome c Family Proteins.”, *PLoS One.*, **10**(4), e0123653, (2015), [10.1371/journal.pone.0123653](https://doi.org/10.1371/journal.pone.0123653)
14. M. Deshpande, P. Parui, H. Kamikubo, M. Yamanaka, S. Nagao, H. Komori, M. Kataoka, Y. Higuchi, *S. Hirota, “Formation of Domain-Swapped Oligomer of Cytochrome c from Its Molten Globule State Oligomer” *Biochemistry*, **53** (28), 4696-4703, (2014), [10.1021/bi500497s](https://doi.org/10.1021/bi500497s)
15. Y. Kobayashi, H. Tsutsumi, T. Abe, K. Ikeda, Y. Tashiro, S. Unzai, H. Kamikubo, M. Kataoka, H. Hiroaki, D.*Hamada, “Decreased Amyloidogenicity by Mutational Modulation of Surface Properties of the Immunoglobulin Light Chain BRE Variable Domain,” *Biochemistry*. **53**(31), 5162-73, (2014), [10.1021/bi5007892](https://doi.org/10.1021/bi5007892)
16. H. Nakagawa, Y. Yonetani, K. Nakajima, S. Ohira-Kawamura, T.Kikuchi, Y. Inamura, M. Kataoka,

- *H. Kono, “Local Dynamics Coupled to Hydration Water Determines DNA-sequence Dependent Deformability”, *Physical Review E*, **90**, 22723, (2014), 10.1103/PhysRevE.90.022723
17. *D. Novitasari, H. Kamikubo, Y. Yamazaki, M. Yamaguchi, M. Kataoka, “Excited-State Proton Transfer in Fluorescent Photoactive Yellow Protein Containing 7-Hydroxycoumarin”, *Adv. Mater. Res.*, **896**, 85-88, (2014), 10.4028/www.scientific.net/AMR.896.85
 18. M. Hamaguchi, H. Kamikubo, K. N. Suzuki, Y. Hagihara, I. Yanagihara, I. Sakata, M. Kataoka, *D. Hamada, “Structural Basis of Alpha-Catenin Recognition by EspB from Enterohaemorrhagic *E. coli* Based on Hybrid Strategy Using Low-resolution Structural and Protein Dissection”, *PLoS One*, **8** (8), e71618, (2013), [10.1371/journal.pone.0071618](https://doi.org/10.1371/journal.pone.0071618)
 19. N. Inoue, D. Hamada, H. Kamikubo, K. Hirata, M. Kataoka, M. Yamamoto, M. Ikawa, M. Okabe, *Y. Hagihara, “Molecular Dissection of IZUMO1, a Sperm Protein Essential for Sperm-egg Fusion”, *Development*, **140**, 3221-3229, (2013), [10.1242/dev.094854](https://doi.org/10.1242/dev.094854)
 20. Y. Kita, H. Kamikubo, M. Kataoka, *M. Tachikawa, “Theoretical Analysis of the Geometrical Isotope Effect on the Hydrogen Bonds in Photoactive Yellow Protein with Multi-component Density Functional Theory”, *Chem. Phys.*, **419**, 50-53, (2013), [10.1016/j.chemphys.2012.11.022](https://doi.org/10.1016/j.chemphys.2012.11.022)
 21. J. Uewaki, H. Kamikubo, J. Kurita, N. Hiroguchi, H. Moriuchi, M. Yoshida, M. Kataoka, N. Utsunomiya-Tate, *S. Tate, “Preferential Domain Orientation of HMGB2 Determined by the Weak Intramolecular Interactions Mediated by the Interdomain Linker”, *Chem. Phys.*, **419**, 212-223, (2013), [10.1016/j.chemphys.2013.02.004](https://doi.org/10.1016/j.chemphys.2013.02.004)
 22. *J. Yuasa, T. Ohno, H. Tsumatori, R. Shiba, H. Kamikubo, M. Kataoka, Y. Hasegawa, *T. Kawai, “Fingerprint Signatures of Lanthanide Circularly Polarized Luminescence from Proteins Covalently Labeled with a Beta-diketonate Europium(III) Chelate”, *Chem. Commun.*, **49**(41), 4604-4606, (2013), [10.1039/c3cc40331a](https://doi.org/10.1039/c3cc40331a)
 23. P. Parui, M. Deshpande, S. Nagao, H. Kamikubo, Y. Higuchi, M. Kataoka, S. *Hirota “Formation of oligomeric cytochrome c during folding by intermolecular hydrophobic interaction between N- and C-terminal α -helices” *Biochemistry*, **52**(48), 8732-8744, (2013), [10.1021/bi400986g](https://doi.org/10.1021/bi400986g)

【佐藤啓文】

1. *H. Okumura, M. Higashi, Y. Yoshida, H. Sato, R. Akiyama, “Theoretical Approaches for Dynamical Ordering of Biomolecular Systems”, *Biochimica et Bio- physica Acta (BBA) - General Subjects*, **1862**(2), 212-228, (2018), [10.1016/j.bbagen.2017.10.001](https://doi.org/10.1016/j.bbagen.2017.10.001)
2. Y. Matsumura, S. Iuchi, *H. Sato, “A Model Electronic Hamiltonian for the Self-assembly of an Octahedron-shaped Coordination Capsule”, *Phys. Chem. Chem. Phys.*, **20**, 1164-1172, (2018), [10.1039/c7cp06094g](https://doi.org/10.1039/c7cp06094g)
3. Y. Matsumura, S. Iuchi, S. Hiraoka, *H. Sato, “Chiral Effects on the Final Step of an Octahedron-shaped Coordination Capsule Self-assembly”, *Phys. Chem. Chem. Phys.*, **20**, 7383-7386, (2018), [10.1039/C7CP08237A](https://doi.org/10.1039/C7CP08237A)
4. *T. Fujita, Y. Haketa, H. Maeda, T. Yamamoto, “Relating Stacking Structures and Charge Transport in Crystal Polymorphs of the Pyrrole-Based pi-Conjugate Molecule”, *Org. Electron.*, **49**, 53, (2017), [10.1016/j.orgel.2017.06.028](https://doi.org/10.1016/j.orgel.2017.06.028)
5. T. Fujita, *T. Yamamoto, “Assessing the Accuracy of Integral Equation Theories for Nano-sized Hydrophobic Solutes in Water”, *J. Chem. Phys.*, **147**, 014110, (2017), [10.1063/1.4990502](https://doi.org/10.1063/1.4990502)
6. H. Arefi, *T. Yamamoto, “Communication: Self-assembly of a Model Supramolecular Polymer Studied by Replica Exchange with Solute Tempering”, *J. Chem. Phys.*, **147**, 211102 (2017), [10.1063/1.5008275](https://doi.org/10.1063/1.5008275)
7. Y. Matsumura, S. Hiraoka, *H. Sato, “A Reaction Model on the Self-assembly Process of Octahedron-shaped Coordination Capsules”, *Phys. Chem. Chem. Phys.*, **19**, 20338-20342, (2017), [10.1039/c7cp03493h](https://doi.org/10.1039/c7cp03493h)
8. H. Arefi, *T. Yamamoto, “Self-assembly of a Model Supramolecular Polymer Studied by Replica Exchange with Solute Tempering”, *J. Chem. Phys. (Commun)*, **147**, 211102, (2017), [10.1063/1.5008275](https://doi.org/10.1063/1.5008275)
9. Y. Yoshida, *H. Sato, J.W.R. Morgan, D.J. Wales, “Potential Energy Landscapes of Tetragonal Pyramid Molecules”, *Chem.Phys.Lett.*, **664**, 5-9, (2016), [10.1016/j.cplett.2016.09.058](https://doi.org/10.1016/j.cplett.2016.09.058)
(Editor's choice, Front Cover に採用)
10. Y. Matsumura *H. Sato, “An Integral Equation Theory for Solvation Effects on the Molecular Structural Fluctuation”, *J. Chem. Phys.*, **143**, No. 1, 014104 , (2015), [10.1063/1.4923038](https://doi.org/10.1063/1.4923038)
11. K. Kikui, S. Hayaki, K. Kido, D. Yokogawa, K. Kasahara, Y. Matsumura, *H. Sato, S. Sakaki, “Solvent

- Structure of Ionic Liquid with Carbon Dioxide”, *J. Mol. Liq.*, **217**, 12-16, (2015), 10.1016/j.molliq.2015.06.061
12. K. Kido, K. Kasahara, *H. Sato, S. Sakaki, “A Molecular Level Study of Selective Cation Capture by a Host-guest Mechanism for 25,26,27,28-tetramethoxycalix[4]arene in MCLO”, *Mol. Simul.*, **41**, (10-12), 881- 891, (2014), 1080/08927022.2014.895002
 13. K. Kasahara, *H. Sato, “Development of Three-dimensional Site-site Smoluchowski- Vlasov Equation and Application to Electrolyte Solutions”, *J. Chem. Phys.*, **140**, 244110, (2014), 10.1063/1.4884386
 14. T. Inagaki, S. Aono, H. Nakano, *T. Yamamoto, “Like-Charge Attraction of Molecular Cations in Water: Subtle Balance between Interionic Interactions and Ionic Solvation Effect”, *J. Phys. Chem. B*, **118**(20), 5499-5508, (2014), 10.1021/jp501212y
 15. T. Inagaki, *T. Yamamoto, “Critical Role of Deep Hydrogen Tunneling to Accelerate the Antioxidant Reaction of Ubiquinol and Vitamin E”, *J. Phys. Chem. B*, **118**(4), 937-950, (2014), 10.1021/jp410263f
 16. H. Nakano, *T. Yamamoto, “Accurate and Efficient Treatment of Continuous Solute Charge Density in the Mean-Field QM/MM Free Energy Calculation”, *J. Chem. Theory Comp.*, **9**(1), 188-203, (2013), 10.1021/ct300831t
- 【高田十志和】
1. *T. Takata, D. Aoki, “Topology-Transformable Polymer: Linear-Branched Polymer Structure Change Utilizing Mechanical Linking of Polymer Chains”, *Polymer Journal*, **50**, 127–147, (2018), 10.1038/pj.2017.60
 2. H. Sato, D. Aoki, *T. Takata, “Which One is Bulkier: The 3,5-Dimethylphenyl or the 2,6-Dimethylphenyl Group? Development of Size-Complementary Molecular and Macromolecular [2]Rotaxanes”, *Chem. Asian J.*, **13**, 785-789, (2018), 10.1002/asia.201800170
 3. K. Iijima, D. Aoki, H. Otsuka, *T. Takata, “Synthesis of Rotaxane Cross-Linked Polymers With Supramolecular Cross-Linkers Based on γ -CD and PTHF Macromonomers: The Effect of The Macromonomer Structure on The Polymer Properties”, *Polymer*, **128**, 392–396, (2017), 10.1016/j.polymer.2017.01.024
 4. J. Sawada, D. Aoki, M. Kuzume, K. Nakazono, S. Uchida, H. Otsuka, *T. Takata, “Vinylic Rotaxane Cross-linker for Toughened Network Polymer via Radical Polymerization of Vinyl Monomers”, *Polym. Chem.*, **8**, 1878-1881, (2017), 10.1039/C7PY00193B
 5. D. Aoki, G. Aibara, S. Uchida, *T. Takata, “A Rational Entry to Cyclic Polymers via Selective Cyclization by Self-Assembly and Topology Transformation of Linear Polymers” *J. Am. Chem. Soc.*, **139**, 6791–6794, (2017), [10.1021/jacs.7b01151](#)
 6. J. Sawada, D. Aoki, *T. Takata, “Vinylic Rotaxane Cross-Linker Comprising Different Axle Length for the Characterization of Rotaxane Cross-linked Polymers” *Macromol. Symposia*, **372**, 115–119, (2017), [10.1002/masy.201600160](#)
 7. D. Aoki, *T. Takata, “Mechanically Linked Supramolecular Polymer Architectures Derived from Macromolecular [2]rotaxanes: Synthesis and Topology Transformation” *Polymer*, **128**, 276–296, (2017), [10.1016/j.polymer.2017.08.020](#)
 8. T. Kureha, D. Aoki, S. Hiroshige, K. Iijima, D. Aoki, *T. Takata, *D. Suzuki, “Decoupled Thermo- and pH-responsive Hydrogel Microspheres Cross-linked by Rotaxane Networks” *Angew. Chem., Int. Ed.*, **56**, 15393–15396, (2017), [10.1002/anie.201709633](#)
 9. S. Hiroshige, T. Kureha, D. Aoki, J. Sawada, D. Aoki, *T. Takata, *D. Suzuki, “Formation of Tough Films by Evaporation of Water from Dispersions of Elastomer Microspheres Crosslinked with Rotaxane Supramolecules” *Chem. Eur. J.*, **23**, 8405–8408, (2017), [10.1002/chem.201702077](#)
 10. H. Jang, K. Iijima, Y. Koyama, S. Uchida, S. Asai, *T. Takata, “Synthesis and Properties of Rotaxane-cross-linked polymers using a double-stranded γ -CD-based inclusion complex as a supramolecular cross-linker” *Polymer*, **128**, 379-385, (2017), [10.1016/j.polymer.2017.01.062](#)
 11. K. Iijima, D. Aoki, H. Otsuka, *T. Takata, “Synthesis of Rotaxane Cross-linked Polymers with Supramolecular Cross-linkers Based on γ -CD and PTHF Macromonomers: The effect of the Macromonomer Structure on the Polymer Properties”, *Polymer*, **128**, 392-396, (2017), [10.1016/j.polymer.2017.01.024](#)
 12. J. Sawada, D. Aoki, M. Kuzume, K. Nakazono, H. Otsuka, *T. Takata, “A Vinylic Rotaxane Cross-Linker for Toughened Network Polymers from the Radical Polymerization of Vinyl Monomers”, *Polym. Chem.*, **8**, 1878–1881, (2017), [10.1039/C7PY00193B](#)

13. K. Iijima, D. Aoki, H. Sogawa, S. Asai, *T. Takata, "Synthesis and Characterization of Supramolecular Cross-linkers Containing Cyclodextrin Dimer and Trimer," *Polym. Chem.*, **7**, 3492–3495, (2016), [10.1039/C6PY00367B](https://doi.org/10.1039/C6PY00367B)
14. H. Sato, D. Aoki, *T. Takata, "Synthesis and Star/Linear Topology Transformation of a Mechanically Linked ABC Terpolymer" *ACS Macro Lett.*, **5**, 699–703, (2016), [10.1021/acsmacrolett.6b00320](https://doi.org/10.1021/acsmacrolett.6b00320)
15. Z. Chen, D. Aoki, S. Uchida, H. Marubayashi, S. Nojima, *T. Takata, "Effect of Component Mobility on the Properties of Macromolecular [2]rotaxanes", *Angew. Chem. Int. Ed.*, **53**, 2778–2781, (2016), [10.1002/anie.201510953](https://doi.org/10.1002/anie.201510953)
16. M. Ogawa, H. Sogawa, Y. Koyama, *T. Takata, "Synthesis of Rotaxane Cross-linked Polymers Derived from Vinyl Monomers Using a Metal-containing Supramolecular Cross-linker", *Polym. J.*, **47**, 580–584, (2015), [10.1038/pj.2015.34](https://doi.org/10.1038/pj.2015.34)
17. M. Ogawa, M. Nagashima, H. Sogawa, S. Kuwata, *T. Takata, "Synthesis and Cavity Size Effect of Pd-containing Macrocycle Catalyst for Efficient Intramolecular Hydroamination of Allylurethane", *Org. Lett.*, **17**, 1664–1667, (2015), [10.1021/acs.orglett.5b00378](https://doi.org/10.1021/acs.orglett.5b00378)
18. J. Sawada, D. Aoki, S. Uchida, H. Otsuka, *T. Takata, "Synthesis of Vinylic Macromolecular Rotaxane Cross-linkers Endowing Network Polymers with Toughness" *Acs Macro Lett.*, **4**, 598–601, (2015), [10.1021/acsmacrolett.5b00242](https://doi.org/10.1021/acsmacrolett.5b00242)
19. D. Aoki, S. Uchida, *T. Takata, "Star/Linear Polymer Topology Transformation Facilitated by Mechanical Linking of Polymer Chains" *Angew. Chem. Int. Ed.*, **54**, 6770–6774, (2015), [10.1002/anie.201500578](https://doi.org/10.1002/anie.201500578)
20. T. Ogawa, K. Nakazono, D. Aoki, S. Uchida, *T. Takata, "Effective Approach to Cyclic Polymer from Linear Polymer: Synthesis and Transformation of Macromolecular [1]Rotaxane", *ACS Macro Lett.*, **4**, 343–347, (2015), [10.1021/acsmacrolett.5b00067](https://doi.org/10.1021/acsmacrolett.5b00067)
21. T. Ogawa, N. Usuki, K. Nakazono, Y. Koyama, *T. Takata, "Linear–cyclic Polymer Structural Transformation and its Reversible Control Using a Rational Rotaxane Strategy", *Chem. Commun.*, **51**, 5606—5609, (2015), [10.1039/C4CC08982K](https://doi.org/10.1039/C4CC08982K)
22. 打田聖, 澤田隼, 飯島圭祐, 青木大輔, 中薗和子, *高田十志和
‘新しい架橋剤：空間連結型架橋剤の合成とビニル重合系への展開’
高分子論文集, **72**, 93–103, (2015), [10.1295/koron.2014-0075](https://doi.org/10.1295/koron.2014-0075)
23. K. Nakazono, T. Ishino, T. Takashima, D. Saeki, D. Natsui, N. Kihara, *T. Takata, "Directed One-pot Syntheses of Crown Ether Wheel-containing Main Chain-type Polyrotaxanes with Controlled Rotaxanation Ratios", *Chem. Commun.*, **50**, 15341–15344, (2014), [10.1039/C4CC06943A](https://doi.org/10.1039/C4CC06943A)
24. Y. Akae, Y. Koyama, S. Kuwata, *T. Takata, "Cyclodextrin-Based Size-Complementary [3]Rotaxanes: Selective Synthesis and Specific Dissociation", *Chem. Eur. J.*, **20**, 17132 – 17136, (2014), [10.1002/chem.201405005](https://doi.org/10.1002/chem.201405005)
25. S. Suzuki, K. Matsuura, K. Nakazono, *T. Takata, "Effect of a Side Chain Rotaxane Structure on the Helix-Folding of Poly(*m*-phenylene diethynylene)", *Polym. J.*, **46**(6), 355–365, (2014), [10.1038/pj.2014.4](https://doi.org/10.1038/pj.2014.4) (Selected as a Cover Page Picture)
26. D. Aoki, S. Uchida, *T. Takata, "Mechanically Linked Block/Graft Copolymers: Effective Synthesis via Functional Macromolecular [2]Rotaxanes", *ACS Macro Lett.*, **3**(4), 324–328, (2014), [10.1021/mz5001306](https://doi.org/10.1021/mz5001306)
27. D. Aoki, S. Uchida, *T. Takata, "Synthesis and Characterization of A Mechanically Linked Transformable Polymer", *Polym. J.*, **46**(9), 546–552, (2014), [10.1038/pj.2014.22](https://doi.org/10.1038/pj.2014.22)
28. Y. Abe, H. Okamura, S. Uchida, *T. Takata, "Synthesis of Main Chain-type Liquid Crystalline Polyrotaxanes: Influence of The Wheel Components and Their Mobility on Liquid Crystalline Properties", *Polym. J.*, **46**(9), 553–558, (2014), [10.1038/pj.2014.23](https://doi.org/10.1038/pj.2014.23)

【立川仁典】

1. Y. Sugiura, K. Suzuki, *T. Takayanagi, Y. Kita, M. Tachikawa, "Reduction of OH Vibrational Frequencies in Amino Acids by Positron Attachment", *J. Comput. Chem.*, in press, (2018)
2. Y.-Y. Zhan, T. Kojima, T. Koide, M. Tachikawa, *S. Hiraoka, "A Balance Between van der Waals and Cation- π Interactions That Stabilizes Hydrophobic Assemblies", *Chem. Eur. J.*, **83**, 5132–5137, (2018), [10.1002/chem.201801376](https://doi.org/10.1002/chem.201801376)
3. Y.-Y. Zhan, N. Tanaka, Y. Ozawa, T. Kojima, T. Mashiko, U. Nagashima, M. Tachikawa, *S. Hiraoka, "Importance of Molecular Meshing for the Stabilization of Solvophobic Assemblies", *J. Org. Chem.*, **83**, 5132–5137, (2018), [10.1021/acs.joc.8b00495](https://doi.org/10.1021/acs.joc.8b00495)

4. K. Sugiura, M. Tachikawa, *T. Udagawa, "Nuclear Quantum Effect and H/D Isotope Effect on Cl · + (H₂O)_n → HCl + OH · (H₂O)_{n-1} (n = 1-3) Reactions", *RSC Advances*, **8**, 17191-17201, (2018), 10.1039/C8RA02679C
5. *R. Harada, T. Mashiko, M. Tachikawa, S. Hiraoka, *Y. Shigeta, "Programed Dynamical Ordering in the Self-organization Processes of a Nanocube: A Molecular Dynamics Study", *Phys. Chem. Chem. Phys.*, **20**, 9115-9122, (2018), 10.1039/C8CP00284C
6. N. Tanaka, Y.-Y. Zhan, Y. Ozawa, T. Kojima, T. Koide, T. Mashiko, U. Nagashima, M. Tachikawa, *S. Hiraoka, "Semi-quantitative Evaluation of Molecular Meshing by Surface Analysis with Varying Probe Radii", *Chem. Commun.*, **54**, 3335-3338, (2018), 10.1039/c8cc00695d
7. Y.-Y. Zhan, K. Ogata, T. Kojima, T. Koide, K. Ishii, T. Mashiko, M. Tachikawa, S. Uchiyama, *S. Hiraoka, "Hyperthermostable Cube-shaped Assembly in Water", *Communications Chemistry*, **1**, 14, (2018), 10.1038/s42004-018-0014-2
8. *H. Kato, S. Yoshimoto, A. Ueda, S. Yamamoto, Y. Kanematsu, M. Tachikawa, H. Mori, J. Yoshinobu, I. Matsuda, "Strong Hydrogen-Bonds at the interface between Proton-Donating and Accepting Self-Assembled Monolayers on Au(111)", *Langmuir*, **34**, 2189-2197, (2018), 10.1021/acs.langmuir.7b03451
9. *T. Udagawa, *M. Tachikawa, "Reaction Mechanism of Hydrogen-tritium Exchange Reactions between Several Organic and HTO Molecules: A role of the second HTO", *RSC Advances*, **8**, 3878-3888, (2018), 10.1039/C7RA13110K
10. T. Kawatsu, *M. Tachikawa, "The Quantum Fluctuations of the Fullerene Cage Modulate the Internal Magnetic Environment", *Phys. Chem. Chem. Phys.*, **20**, 1673-1684, (2018), 10.1039/C7CP06401B
11. K. Suzuki, *T. Takayanagi, Y. Kita, M. Tachikawa, T. Oyamada, "Quantum Dynamics Calculations for e+ + LiH → Li+ + [H; e+] Dissociative Positron Attachment Using a Pseudopotential Model", *Comput. Theo. Chem.*, **1123**, 135-141, (2018), 10.1016/j.comptc.2017.11.023
12. S. Kai, V. Martí-Centelles, Y. Sakuma, T. Mashiko, T. Kojima, U. Nagashima, M. Tachikawa, P. J. Lusby, and *S. Hiraoka, "Quantitative Analysis of Self-Assembly Process of a Pd2L4 Cage Consisting of Rigid Dtopic Ligands", *Chem. Eur. J.*, **24**, 663-671, (2018), 10.1002/chem.201704285
13. S. Kai, Y. Sakuma, T. Mashiko, T. Kojima, M. Tachikawa, *S. Hiraoka, "The Effect of Solvent and Coordination Environment of Metal Source on the Self-Assembly Pathway of a Pd(II)-mediated Coordination Capsule", *Inorg. Chem.*, **56**, 12652-12663, (2017), 10.1021/acs.inorgchem.7b02152
14. *N. Kungwan, C. Ngaojampa, Y. Ogata, T. Kawatsu, Y. Oba, Y. Kawashima, *M. Tachikawa, "Solvent Dependence of Double Proton Transfer in the Formic Acid Formamidine Complex: Path Integral Molecular Dynamics Investigation", *J. Phys. Chem. A*, **121**, 7324-7334, (2017), 10.1021/acs.jpca.7b07010
15. *T. Takayanagi, K. Suzuki, T. Yoshida, Y. Kita, M. Tachikawa, "Quantum Dynamics Study on the Binding of a Positron to vibrationally Excited States of Hydrogen Cyanide Molecule", *Chem. Phys. Lett.*, **675**, 118-123, (2017), 10.1016/j.cplett.2017.03.025
16. K. Yamamoto, Y. Kanematsu, U. Nagashima, A. Ueda, H. Mori, *M. Tachikawa, "Multicomponent DFT Study of Geometrical H/D Isotope Effect on Hydrogen-bonded Organic Conductor, κ-H₃(Cat EDT-ST)₂", *Chem. Phys. Lett.*, **674**, 168-172, (2017), 10.1016/j.cplett.2017.02.073
17. *C. Ngaojampa, T. Kawatsu, Y. Oba, N. Kungwan, *M. Tachikawa, "Asymmetric Hydrogen Bonding in Formic Acid-nitric Acid Dimer Observed by Quantum Molecular Dynamics Simulations", *Theor. Chem. Acc.*, **136**, 30, (2017), 10.1007/s00214-017-2057-3
18. *T. Udagawa, K. Sugiura, K. Suzuki, *M. Tachikawa, "Unusual H/D isotope effect in isomerization and keto-enol tautomerism reactions of pyruvic acid: Nuclear quantum effect restricts some rotational isomerization reactions", *RSC Advances*, **7**, 9328-9337, (2017), 10.1039/C6RA28271G
19. T. Mashiko, S. Hiraoka, U. Nagashima, *M. Tachikawa, "Theoretical Study on Substituent and Solvent Effects for Nanocube Formed with Gear-shaped Amphiphile Molecules", *Phys. Chem. Chem. Phys.*, **19**, 1627-1631, (2017), 10.1039/C6CP07754D
20. Y. Kita, *M. Tachikawa, "Effects of Vibrational Anharmonicity and Inter-mode Couplings on the Binding Energy of a Positron to Molecules", *AIP Conf. Proc.*, **791**, 012015, (2017), 10.1088/1742-6596/791/1/012015
21. *T. Udagawa, *M. Tachikawa, "Nuclear Quantum Effect and H/D Isotope Effect on F + (H₂O)_n → FH + (H₂O)_{n-1}OH (n = 1-3) Reactions", *J. Chem. Phys.*, **145**, 164310 (10 pages), (2016), 10.1063/1.4966162
22. K. Yamamoto, Y. Kanematsu, U. Nagashima, A. Ueda, H. Mori, *M. Tachikawa, "Theoretical Study of

- H/D Isotope Effect on Phase Transition of Hydrogen-bonded Organic Conductor κ -H₃(Cat-EDT-TTF)₂", *Phys. Chem. Chem. Phys.* (Communication), **18**, 29673-29680, (2016), 10.1039/C6CP05414E
23. Y. Hamada, Y. Kanematsu, *M. Tachikawa, "QM/MM Study on Sialyltransferase Reaction Mechanism", *Biochemistry*, **55**, 5764-5771, (2016), 10.1021/acs.biochem.6b00267
24. M. Narukawa-Nara, A. Nakamura, K. Kikuzato, Y. Kakei, A. Sato, Y. Mitani, Y. Yamasaki-Kokudo, T. Ishii, KI. Hayashi, T. Asami, T. Ogura, S. Yoshida, S. Fujioka, T. Kamakura, T. Kawatsu, M. Tachikawa, K. Soeno, *Y. Shimada, "Aminooxy-naphthylpropionic Acid and its Derivatives are Inhibitors of Auxin Biosynthesis Targeting Trp Aminotransferase: Structure-Activity Relationships", *The Plant Journal*, **87**, 245-257, (2016), 10.1111/tpj.13197
25. Y. Oba, T. Kawatsu, *M. Tachikawa, "A Path Integral Molecular Dynamics Study of the Hyperfine Coupling Constants of the Muoniated and Hydrogenated Acetone Radicals", *J. Chem. Phys.*, **145**, 064301 (15 pages), (2016), 10.1063/1.4960077
26. Y. Ogata, T. Kawatsu, *M. Tachikawa, "Can Low-barrier Hydrogen Bond Exist in Systems with Second Row Elements? An ab Initio Path Integral Molecular Dynamics Study for Deprotonated Hydrogen Sulfide Dimer", *Theor. Chem. Acc.*, **135**, 200 (11 pages), (2016), 10.1007/s00214-016-1958-x
27. M. Nummela, *H. Raebiger, D. Yoshida, M. Tachikawa, "Positron Binding Properties of Glycine and Its Aqueous Complexes", *J. Phys. Chem. A*, **120**, 4037-4042, (2016), 10.1021/acs.jpca.6b01780
28. *Y. Kanematsu, Y. Takano, M. Tachikawa, "Inverse Ubbelohde Effect in the Short Hydrogen Bond of Photosystem II: Relation between H/D Isotope Effect and Symmetry in Potential Energy Profile", *J. Comput. Chem.*, **37**, 2140-2145, (2016), 10.1002/jcc.24438 (Cover Image に採用)
29. Y. Takeda, *Y. Kita, *M. Tachikawa, "Theoretical Study of a Positron-attachment to Vibrational Excited States for Non-polar Carbon Disulfide Molecule", *Eur. Phys. J. D*, **70**, 132 (5 pages), (2016), 10.1140/epjd/e2016-70140-7
30. Y. Oba, T. Kawatsu, *M. Tachikawa, "Thermal Dependence on Structures of Muoniated and Hydrogenated Acetone Radicals", *AIP Conf. Proc.*, **1790**, 020022 (4pages), (2016), 10.1063/1.4968648
31. M. Hashimoto, T. Ishimoto, *M. Tachikawa, *T. Udagawa, "Analysis of Exponent Values of Gaussian-type Functions on Quantum Protons and Deuterons in Charged or Polarized Systems", *Int. J. Quant. Chem.*, **116**, 961-967, (2016), 10.1002/qua.25117
32. Y. Kanematsu, H. Kamikubo, M. Kataoka, *M. Tachikawa, "Vibrational Analysis on the Revised Potential Energy Curve of the Low-barrier Hydrogen Bond in Photoactive Yellow Protein", *Comput. Struct. Biotechnol. J.*, **14**, 16-19, (2016), 10.1016/j.csbj.2015.10.003
33. *M. Tachikawa, "Positron-attachment to Small Molecules: Vibrational Enhancement of Positron Affinities with Configuration Interaction Level of Multi-component Molecular Orbital Approach", *AIP Conf. Proc.*, **1702**, 090038 (4pages), (2015), 10.1063/1.4938847
34. Y. Kanematsu, H. Kamikubo, M. Kataoka, *M. Tachikawa, "Vibrational Analysis on the Revised Potential Energy Curve of the Low-barrier Hydrogen Bond in Photoactive Yellow Protein", *Comput. Struct. Biotechnol. J.* **14**, 16-19, (2015), 10.1016/j.csbj.2015.10.003
35. Y. Kanematsu, Y. Kamiya, K. Matsuo, K. Gekko, *K. Kato, *M. Tachikawa, "Isotope Effect on the Circular Dichroism Spectrum of Methyl α -D-glucopyranoside in Aqueous Solution", *Sci. Rep.*, **5**, 17900 (5pages) (2015), 10.1038/srep17900
36. Y. Ogata, Y. Kawashima, *K. Takahashi, *M. Tachikawa, "Theoretical Vibrational Spectra of OH⁻ (H₂O)₂: Effect of Quantum Distribution and Vibrational Coupling", *Phys. Chem. Chem. Phys.*, **17**, 25505-25515, (2015), 10.1039/C5CP03632A
37. *T. Udagawa, K. Suzuki, *M. Tachikawa, "Multicomponent Molecular Orbital-climbing Image-nudged Elastic Band Method to Analyze Chemical Reactions Including Nuclear Quantum Effect: Application to Hydrogen Transfer Reaction", *ChemPhysChem*, **16**, 3156-3160, (2015), 10.1002/cphc.201500498
38. S. Watanabe, Y. Ogata, T. Kawatsu, Y. Kawashima, *M. Tachikawa, "Effects of Monohydration on an Adenine-thymine Base Pair", *Theor. Chem. Acc.*, **134**, 84 (12 pages), (2015),
39. *T. Udagawa, *M. Tachikawa, "H/D Isotope Effect on Charge-inverted Hydrogen-bonded Systems: Systematic Classification of Three Different Types in H₃XH...YH₃ (X = C, Si, or Ge, and Y = B, Al, or Ga) with Multicomponent Calculation", *J. Comput. Chem.*, **36**, 1647-1654, (2015), 10.1002/jcc.23978
40. Y. Kanematsu, *M. Tachikawa, "Performance Test of Multicomponent Quantum Mechanical Calculation with Polarizable Continuum Model for Proton Chemical Shift", *J. Phys. Chem. A*, **119**, 4933-4938, (2015), 10.1021/jp512877a

41. *T. Udagawa, *M. Tachikawa, "Why does Deuterium Substitution Lead to the Contraction of X...Pi distance? Origin of the Reverse Ubbelohde Effect in XH...Pi Interaction", *Theor. Chem. Acc.*, **134**, 24 (5 pages), (2015), 10.1007/s00214-015-1633-7
42. *K. Egashira, Y. Yamada, Y. Kita, M. Tachikawa, "Ferromagnetic Spin Coupling in the Chromium Dimer Cation: Measurements by Photodissociation Spectroscopy Combined with Coupled-cluster Calculations", *J. Chem. Phys.*, **142**, 054309 (4pages), (2015), 10.1063/1.4907197
43. T. Mashiko, K. Yamada, S. Hiraoka, U. Nagashima, *M. Tachikawa, "Molecular Dynamics Simulations of Self-assembled Nanocubes in Methanol", *Mol. Sim.*, **41**, 845-849, (2014), [10.1080/08927022.2014.940523](https://doi.org/10.1080/08927022.2014.940523)
44. K. Yamada, Y. Kawashima, *M. Tachikawa, "Quantum Simulation for Muoniated and Deuterated Methyl Radicals in Implicit Water Solvent: Combined Ab Initio Path Integral Molecular Dynamics and the Polarizable Continuum Model Simulation Study", *Mol. Sim.*, **41**, 832-839, (2014), [10.1080/08927022.2014.938070](https://doi.org/10.1080/08927022.2014.938070)
45. Y. Ogata, Y. Kawashima, K. Takahashi, *M. Tachikawa, "Is the Structure of Hydroxide Dihydrate OH⁻(H₂O)₂? : An ab Initio Path Integral Molecular Dynamics Study", *Theor. Chem. Acc.*, **134**, 1582-1587, (2014), 10.1007/s00214-014-1587-1
46. Y. Kanematsu, *M. Tachikawa, "Theoretical Analysis of Geometry and NMR Isotope Shift in Hydrogen-bonding Center of Photoactive Yellow Protein by Combination of Multicomponent Quantum Mechanics and ONIOM Scheme", *J. Chem. Phys.*, **141**, 185101 (8pages), (2014), <http://dx.doi.org/10.1063/1.4900987>
47. *N. Kungwana, Y. Ogata, S. Hannongbua, *M. Tachikawa, "Nuclear Quantum Effect and Temperature Dependency on the Hydrogen-bonded Structure of 7-azaindole Dimer", *Theor. Chem. Acc.*, **133**, 1553-1562, (2014), [10.1007/s00214-014-1553-y](https://doi.org/10.1007/s00214-014-1553-y)
48. *T. Udagawa, T. Ishimoto, *M. Tachikawa, "H/D Isotope Effect on Structures, Binding Energies, and Basis Set Superposition Errors in F-(H₂O)_n (n = 1-3) Clusters", *Chem. Phys.*, **441**, 101-108, (2014), [10.1016/j.chemphys.2014.07.014](https://doi.org/10.1016/j.chemphys.2014.07.014)
49. *M. Tachikawa, "Positron-attachment to Acetonitrile, Acetaldehyde, and Acetone Molecules: Vibrational Enhancement of Positron Affinities with Configuration Interaction Level of Multi-component Molecular Orbital Approach", *J.Phys.Conf.Ser.*, **488**, 012053 (7pages), (2014), [10.1088/1742-6596/488/1/012053](https://doi.org/10.1088/1742-6596/488/1/012053)
50. Y. Yamada, *Y. Kita, M. Tachikawa, "Theoretical Prediction of the Binding of a Positron to a Formaldehyde Molecule Using a First-principles Calculation", *Phys. Rev. A*, **89**, 062711 (5pages), (2014), [10.1103/PhysRevA.89.062711](https://doi.org/10.1103/PhysRevA.89.062711).
51. *T. Oyamad, *M. Tachikawa, "Multi-component Molecular Orbital Sudy on Positron Attachment to Alkali-metal Hydride Molecules: Nature of Chemical Bonding and Dissociation Limits in [LiH; e⁺]", *Eur. Phys. J. D*, **68**, 231-239 (2014), [10.1140/epjd/e2014-40708-4](https://doi.org/10.1140/epjd/e2014-40708-4).
52. T. Udagawa, T. Tsuneda, *M. Tachikawa, "Electron-nucleus Correlation Functional for Multicomponent Density-functional Theory", *Phys. Rev. A*, **89**, 052519 (5pages), (2014), [10.1103/PhysRevA.89.052519](https://doi.org/10.1103/PhysRevA.89.052519)
53. Y. Kanematsu, *M. Tachikawa, "Development of Multicomponent Hybrid Density Functional Theory with Polarizable Continuum Model for the Analysis of Nuclear Quantum Effect and Solvent Effect on NMR Chemical Shift", *J. Chem. Phys.*, **140**, 164111 (7pages), (2014), [10.1063/1.4872006](https://doi.org/10.1063/1.4872006)
54. K. Yamada, Y. Kawashima, *M. Tachikawa, "Accurate Prediction of Hyperfine Coupling Constants in Muoniated and Hydrogenated Ethyl Radicals: Ab Initio Path Integral Simulation Study with Density Functional Theory Method", *J. Chem. Theor. Comput.*, **10**(5), 2005-2015, (2014), [10.1021/ct500027z](https://doi.org/10.1021/ct500027z)
55. *Y. Kita, M. Tachikawa, "Theoretical Investigation of the Binding of a Positron to Vibrational Excited States of Hydrogen Cyanide Molecule", *Eur. Phys. J. D*, **68**, 116-122, (2014), [10.1140/epjd/e2014-40799-9](https://doi.org/10.1140/epjd/e2014-40799-9)
56. Y. Yamada, *Y. Kita, M. Tachikawa, M. Towler, R. J. Needs, "Quantum Monte Carlo and High-level Ab Initio Molecular Orbital Investigation of Dissociation Channels of the Positronic Alkali-metal Hydrides, [XH;e⁺] (X = Li, Na, and K)", *Eur.Phys.J.D*, **68**, 63-68, (2014), [10.1140/epjd/e2014-40734-2](https://doi.org/10.1140/epjd/e2014-40734-2)
57. Y. Oba, *M. Tachikawa, "Theoretical Investigation of a Positron Binding to an Aspartame Molecule Using the ab Initio Multicomponent Molecular Orbital Approach", *Int. J. Quant. Chem.*, **114**(17), 1146-1149, (2014), [10.1002/qua.24641](https://doi.org/10.1002/qua.24641)
58. *Y. Kawashima, *M. Tachikawa, "An Ab Initio Path Integral Molecular Dynamics Study of the Nuclear Quantum Effect on Out-of-plane Ring Deformation of Hydrogen Maleate Anion", *J. Chem. Theor.*

Comput., **10**(1), 153-163, (2014), [10.1021/ct4007986](https://doi.org/10.1021/ct4007986)

59. *T. Udagawa, *M. Tachikawa, “Why is N...Be Distance of NH₃H⁺...DBeH Shorter than that of NH₃D⁺...HBeH? Paradoxical Geometrical Isotope Effects for Partially Isotope-substituted Dihydrogen-bonded Isotopomers”, *J. Comput. Chem. (Communication)*, **35**(4), 271-274, (2014), [10.1002/jcc.23505](https://doi.org/10.1002/jcc.23505)
60. K. Yamada, Y. Kawashima, *M. Tachikawa, “Muon-Electron Hyperfine Coupling Constants of Muoniated Ethyl Radical: a Path Integral Simulation Study with Semiempirical Molecular Orbital Method”, *Chin. J. Phys.*, **52**, 126-137, (2014), [10.6122/CJP.52.126](https://doi.org/10.6122/CJP.52.126)

【田中良和】

1. 加藤早苗, 松井崇, *田中良和, “3.8MDa の超巨大酸素運搬蛋白質へモシアニン会合体の結晶構造”, 生化学 Vol.90, 238-243, (2018) , [10.14952/SEIKAGAKU.2018.900238](https://doi.org/10.14952/SEIKAGAKU.2018.900238)
2. Z. Peng, M. Takeshita, N. Shibata, H. Tada, Y. Tanaka, *J. Kaneko, “Rim Domain Loops of Staphylococcal β -pore Forming Bi-component Toxin S-components 3 Recognize Target Human Erythrocytes in a Coordinated Manner,” *J. Biochem.*, Accepted (2018), [10.1093/jb/mvy030](https://doi.org/10.1093/jb/mvy030).
3. *T. Uchida, T. Funamizu, M. Chen, Y. Tanaka, K. Ishimori, “Heme Binding to Porphobilinogen Deaminase from Vibrio cholerae Decelerates the Formation of 1-Hydroxymethylbilane,” *ACS Chem. Biol.*, **13**, 750-760, (2018), [10.1021/acscchembio.7b00934](https://doi.org/10.1021/acscchembio.7b00934)
4. *S. Kato, T. Matsui, C. Gatsogiannis, *Y. Tanaka, “Molluscan Hemocyanin: Structure, Evolution, and Physiology”, *Biophys. Rev.*, **10**, 191-202 (2018), [10.1007/s12551-017-0349-4](https://doi.org/10.1007/s12551-017-0349-4)
5. M. Chen, M. Kubo, K. Kato, Y. Tanaka, Y. Liu, F. Long, W. Whitman, P. Lill, C. Gatsogiannis, S. Raunser, N. Shimizu, A. Shinoda, A. Nakamura, I. Tanaka, *M. Yao, “Structural Basis for tRNA-Dependent Cysteine Biosynthesis”, *Nature Commun.*, **8**, 1512, (2017), [10.1038/s41467-017-01543-y](https://doi.org/10.1038/s41467-017-01543-y).
6. T. Kunthic, H. Watanabe, R. Kawano, Y. Tanaka, B. Promdonkoy, M. Yao, *P. Boonserm, “pH Regulates Pore Formation of a Protease Activated Vip3Aa from *Bacillus thuringiensis*”, *Biochim. Biophys. Acta, (BBA), Biomembranes*, **1859**, 2234-2241, (2017), [10.1016/j.bbamem.2017.08.018](https://doi.org/10.1016/j.bbamem.2017.08.018)
7. M. Chen, S. Asai, S. Narai, S. Nambu, N. Omura, Y. Sakaguchi, T. Suzuki, M. Ikeda-Saito, K. Watanabe, M. Yao, *N. Shigi, Y. Tanaka, “Biochemical and Structural Characterization of Oxygen-sensitive 2-thiouridine Synthesis Catalyzed by the Iron-sulfur Protein TtuA”, *Proc. Natl. Acad. Sci. USA*, **114**, 4954-4959, (2017), [10.1073/pnas.1615585114](https://doi.org/10.1073/pnas.1615585114)
8. Y. Miyabe, T. Furuta, T. Takeda, G. Kanno, T. Shimizu, Y. Tanaka, Z. Gai, H. Yasui, *H. Kishimura, “Structural Properties of Phycoerythrin from Dulse *Palmaria Palmate*”, *J. Food Biochem.*, **41**, e12301, (2017), [10.1111/jfbc.12301](https://doi.org/10.1111/jfbc.12301)
9. Y. Sekine, T. Tanzawa, Y. Tanaka, K. Ishimori, *T. Uchida, “Cytoplasmic Heme-binding Protein (HutX) from *Vibrio Cholerae* is an Intracellular Heme Transport Protein for the Heme-degrading Enzyme, HutZ”, *Biochemistry*, **55**, 884-893, (2016), [10.1021/acs.biochem.5b01273](https://doi.org/10.1021/acs.biochem.5b01273)
10. M. Chen, S. Narai, N. Omura, N. Shigi, S. Chimnaronk, *Y. Tanaka, M. Yao, “Crystallographic Study of Two-Thiouridine Synthetic Complex TtuA-TtuB from *Thermus Thermophiles*”, *Acta Crystallogr. F.*, **72**, 777-781, (2016), [10.1107/S2053230X16014242](https://doi.org/10.1107/S2053230X16014242)
11. K. Oshima, Y. Kakiuchi, Y. Tanaka, T. Ueda, T. Nakashima, *M. Kimura, M. Yao, “Structural Basis for Recognition of a Kink-turn Motif by an Archaeal Homologue of Human RNase P Protein Rpp38”. *Biochem. Biophys. Res. Commun.*, **474**, 541-546, (2016), [10.1016/j.bbrc.2016.04.118](https://doi.org/10.1016/j.bbrc.2016.04.118)
12. Z.Gai, A. Matsuno, K. Kato, S. Kato, R.I.Khan, T. Shimizu, T. Yoshioka, Y. Kato, H. Kishimura, G. Kanno, Y. Miyabe, T. Terada, *Y. Tanaka, Y. Min, “Crystal Structure of the 3.8 MDa Respiratory Supermolecule Hemocyanin at 3.0 Angstrom Resolution”, *Structure*, **23**, 2204-2212, (2015), [10.1016/j.str.2015.09.008](https://doi.org/10.1016/j.str.2015.09.008)
13. T. Sugawara, D. Yamashita, K. Kato, Z. Peng, J. Ueda, J. Kaneko, Y. Kamio, Y. Tanaka, Min Yao, “Structural basis for pore-forming mechanism 1 of staphylococcal α -hemolysin”, *Toxicon*, **108**, 226-231, (2015), [10.1016/j.toxicon.2015.09.033](https://doi.org/10.1016/j.toxicon.2015.09.033)
14. T. Uchida, M. Sasaki, Y. Tanaka, K. Ishimori, “A dye-decolorizing peroxidase from *Vibrio cholera*”, *Biochemistry*, **54**, 6610-6621, (2015), [10.1021/acs.biochem.5b00952](https://doi.org/10.1021/acs.biochem.5b00952)
15. 田中良和, 黄色ブドウ球菌が产生する膜孔形成毒素の作用機構 *Isotope News* 736, 7-11 (2015)
16. 田中良和, 黄色ブドウ球菌の2成分性膜孔形成毒素 γ -ヘモリジンの膜孔形成メカニズム 失敗から明らかになった毒素の戦略 *化学と生物* **53** (3), 136-137 (2015)

17. R.Sasaki, S.Kitazawa, R.Kitahara, H.Nakazawa, Y.Tanaka, I.Kumagai, M.Umetsu, *K.Makabe, “Zinc ion-binding activity of an anti-ZnO VH antibody, 4F2”, *Chem. Letters*, **44**, 1309-1311, (2015), 10.1246/cl.150537
18. A.Matsuno, Z.Gai, M. Taaka, K.Kato, S. Kato, T. Katoh, T. Shimizu, T.Yoshioka, H. Kishimura, Y.Tanaka, M. Yao, “Crystallization and Preliminary X-ray Crystallographic Study of a 3.8-MDa Respiratory Supermolecule Hemocyanin”, *J. Struct. Biol.*, **190**, 379-382, (2015), 10.1016/j.jsb.2015.04.015
19. S. Ito, S.Horikawa, T.Suzuki, H.Kawauchi, Y.Tanaka, T.Suzuki, T.Suzuki, “Human NAT10 is an ATP-dependent RNA Acetyltransferase Responsible for N4-acetylcytidine Formation in 18S rRNA.” *J. Biol. Chem.*, **289**, 35724-35730, (2014), 10.1074/jbc.C114.602698
20. D. Yamashita, T. Sugawara, M. Takeshita, J. Kaneko, Y. Kamio, I. Tanaka, *Y.Tanaka, M. Yao, “Molecular Basis of Transmembrane Beta-barrel Formation of Staphylococcal Pore-forming Toxins”, *Nature Commun.*, **5**, 4897, (2014), [10.1038/ncomms5897](https://doi.org/10.1038/ncomms5897)
21. T. Hayashi, *Y.Tanaka, N. Sakai, U. Okada, M. Yao, N. Watanabe, T. Tamura, I. Tanaka, “Structural and Genomic DNA Analysis of the Putative TetR Transcriptional Repressor SCO7518 from Streptomyces Coelicolor A3(2)”, *FEBS Letters*, **588**(23), 4311-4318, (2014), <http://dx.doi.org/10.1016/j.febslet.2014.09.037>
22. Y. Ushijima, R. Ohniwa, A. Maruyama, S. Saito, Y.Tanaka, *K. Morikawa, “Nucleoid Compaction by MrgAAsp56Ala/Glu60Ala does not Contribute to Staphylococcal Cell Survival against Oxidative Stress and Phagocytic Killing by Macrophage”, *FEMS Microbiol. Letters*, **360**(2), 144-151, (2014), [10.1111/1574-6968.12598](https://doi.org/10.1111/1574-6968.12598)
23. A. Shinoda, Y.Tanaka, M. Yao, I. Tanaka, “Anchoring Protein Crystals to Mounting Loops with Hydrogel Using Inkjet Technology”, *Acta Cryst., D*, **70**, 2794-2799, (2014), [10.1107/S139900471401476X](https://doi.org/10.1107/S139900471401476X)
24. T. Suzuki, K. Yamashita, Y.Tanaka, I. Tanaka, M. Yao, “Crystallization and Preliminary X-ray Crystallographic Analysis of a Bacterial Asn-transamidosome”, *Acta Cryst. F*, **70**, 790-793 (2014), [10.1107/S2053230X14007274](https://doi.org/10.1107/S2053230X14007274)

【寺嶋正秀】

1. K. Takeda, *M.Terazima, “Photoinduced Orientation Change of the Dimer Structure of the Pr-I state of Cph1Δ2”, *Biochemistry*, in press, (2018), 10.1021/acs.biochem.8b00605
2. Y. Nakasone, M. Ohshima, K. Okajima, S. Tokutomi, *M.Terazima, “Photoreaction Dynamics of LOV1 and LOV2 of Phototropin from Chlamydomonas Reinhardtii”, *J.Phys.Chem.B*, **122**, 1801-1815, (2018), 10.1021/acs.jpcb.7b10266
3. A. Takakado, Y.Nakasone, *M.Terazima, “Sequential DNA Binding and Dimerization Processes of the Photosensory Protein EL222”, *Biochemistry*, **57**, 1603-1610, (2018), 10.1021/acs.biochem.7b01206
4. K. Shibata, Y. Nakasone, *M.Terazima, “Photoreaction of BlrP1: a Role of Nonlinear Photo-intensity Sensor”, *Phys.Chem.Chem.Phys.*, **20**, 8133-8142, (2018), 10.1039/c7cp08436f
5. *K. Iwata; *M.Terazima; *H. Masuhara , “Novel Physical Chemistry Approaches in Biophysical Researches with Advanced Application of Lasers: Detection and Manipulation”, *Biochim.Biophys.Acta,(BBA), Gen. Subj.*, **1862**, 335-357, (2018), 10.1016/j.bbagen.2017.11.003
6. A. Takakado, Y. Nakasone, K. Okajima, S. Tokutomi, *M.Terazima, “Light-Induced Conformational Changes of the LOV2-Kinase and the Linker Region in Arabidopsis Phototropin2”, *J.Phys.Chem.B*, **121**, 4414-4421, (2017), [10.1021/acs.jpcb.7b01552](https://doi.org/10.1021/acs.jpcb.7b01552)
7. S. Nozue, M. Katayama, M.Terazima, *S. Kumazaki, “Comparative Study of Thylakoid Membranes in Terminal Heterocysts and Vegetative Cells from Two Cyanobacteria, Rivularia M-261 and Anabaena Variabilis, by Fluorescence and Absorption Spectral Microscopy”, *Biochim. Biophys. Acta, (BBA)*, **1858**, 742-749, (2017), [10.1016/j.bbabi.2017.05.007](https://doi.org/10.1016/j.bbabi.2017.05.007)
8. A. Takakado, Y. Nakasone, *M.Terazima, “Photoinduced Dimerization of a Photosensory DNA-binding Protein EL222 and its LOV Domain”, *Phys.Chem.Chem.Phys.*, **19**, 24855-24865, (2017), [10.1039/c7cp03686h](https://doi.org/10.1039/c7cp03686h)
9. M. Kondoh, *M.Terazima, “Conformational and Intermolecular Interaction Dynamics of Photolyase/Cryptochrome Proteins Monitored by the Time-resolved Diffusion Technique”, *Photochem.Photobiol.*, **93**, 15–25, (2017), [10.1111/php.12681](https://doi.org/10.1111/php.12681)
10. T. Yoshitake, T. Toyooka, Y. Nakasone, K. Zikihara, S. Tokutomi, * M.Terazima, “Macromolecular Crowding Effect for Photoreactions of LOV2 Domains of Arabidopsis Thaliana Phototropin 1”, *J.Mol.Liq.*,

- 217**, 43-50, (2016), 10.1016/j.molliq.2015.08.030
11. S. Nozue, A. Mukuno, Y. Tsuda, T. Shiina, M. Terazima, *S. Kumazaki, "Characterization of Thylakoid Membrane in Filamentous Cyanobacteria and Green Alga with Dual-detector Fluorescence Lifetime Imaging Microscopy with a Systematic Change of Incident Laser Power", *Biochim. Biophys. Acta.*, **1857**, 46-59, (2016), 10.1016/j.bbabi.2015.10.003
 12. K. Kuroi, F. Sato, Y. Nakasone, K. Zikihara, S. Tokutomi, * M. Terazima, "Time-Resolved Fluctuation during the Photochemical Reaction of a Photoreceptor Protein: Phototropin1LOV2-Linker", *Phys. Chem. Chem. Phys.*, **18**, 6228-38, (2016), 10.1039/c5cp07472j
 13. T. Yoshitake, T. Toyooka, Y. Nakasone, K. Zikihara, S. Tokutomi, *M. Terazima, "Macromolecular Crowding Effect for Photoreactions of LOV2 Domains of Arabidopsis Thaliana Phototropin 1", *J. Mol. Liq.*, **217**, 43-50, (2016), 10.1016/j.molliq.2015.08.030
 14. S. Choi, Y. Nakasone, K. J Hellingwerf, *M. Terazima, "Photochemical Reactions of the LOV and the LOV-linker Domains of the Blue Light Sensor Protein YtvA", *Biochemistry*, **55**, 3107-3115, (2016), 10.1021/acs.biochem.6b00263
 15. T. Nakajima, K. Kuroi, Y. Nakasone, K. Okajima, M. Ikeuchi, S. Tokutomi, *M. Terazima, "Anomalous Pressure Effects on the Photoreaction of a Light-sensor Protein from Synechocystis, PixD (Slr1694), and the Compressibility Change of its Intermediates" *Phys. Chem. Chem. Phys.*, **18**, 25915-25925, (2016), 10.1039/C6CP05091C
 16. Y. Nakasone, H. Ooi, Y. Kamiya, H. Asanuma, *M. Terazima, "Dynamics of Inter-DNA Chain Interaction of Photoresponsive DNA", *J. Am. Chem. Soc. (Communication)*, **138**, 9001-9004, (2016), 10.1021/jacs.6b02525
 17. Y. Akiyama, Y. Nakasone, Y. Nakatani, O. Hisatomi, *M. Terazima, "Time-resolved Detection of Light-induced Dimerization of Monomeric Aureochrome-1 and Change in Affinity for DNA", *J. Phys. Chem. B*, **120**, 7360-7370, (2016), 10.1021/acs.jpcb.6b05760
 18. K. Kuroi, K. Okajima, M. Ikeuchi, S. Tokutomi, T. Kamiyama, *M. Terazima, "Pressure-sensitive Reaction Yield of the TePixD Blue-light Sensor Protein", *J. Phys. Chem. B*, **119**, 2897-2907(2015), 10.1021/jp511946u
 19. T. Miyamori, Y. Nakasone, K. Hitomi, J.M. Christie, E. D. Getzoff, M. Terazima, "Reaction dynamics of the UV-B photosensor UVR8", *Photochem. Photobiol. Sci.*, **14**, 995 – 1004, (2015), 10.1039/c5pp00012b
 20. M. Terazima, "Photo-induced Inter-protein Interaction Changes in the time Domain; a Blue Light Sensor protein PixD", *Rapid Comm. Photosci.*, **4**, 1-8 (2015), ISSN 2288-4564
 21. K. Kuroi, K. Okajima, M. Ikeuchi, S. Tokutomi, M. Terazima, "Transient Conformational Fluctuation of TePixD During a Reaction", *Proc. Natl. Acad. Sci. USA*, **111**, 14764-14769, (2014), 10.1073/pnas.1413222111
 22. Y. Nakasone, Y. Kawaguchi, S-G. Kong, M. Wada, M. Terazima, "Photo-Induced Oligomerization of Arabidopsis Thaliana Phototropin 2 LOV1", *J. Phys. Chem. B*, **118**, 14314–14325, (2014), 10.1021/jp509448b
 23. *M. Nishiyama, Y. Shimoda, Y. Kimura, M. Terazima, M. Homma, S. Kojima, "Pressure-Speed Relationship of the Sodium-Driven Flagellar Motor of *Vibrio Alginolyticus*", *Biophys. J.*, **106**, 578A-578A, (2014), 10.1016/j.bpj.2013.11.3206
 24. K. Kuroi, K. Tanaka, K. Okajima, M. Ikeuchi, S. Tokutomi, *M. Terazima, "Anomalous Diffusion of TePixD and Identification of the Photoreaction Product", *Photochem. Photobiol. Sci.*, **12**, 1180-1186, (2013), 10.1039/C3PP25434H
 25. Y. Nakasone, K. Zikihara, S. Tokutomi, *M. Terazima, "Photochemistry of Arabidopsis Phototropin 1 LOV1: Transient Tetramerization", *Photochem. Photobiol. Sci.*, **12**(7), 1171-1179, (2013), 10.1039/c3pp50047k
 26. *M. Nishiyama, Y. Sowa, Y. Kimura, M. Homma, A. Ishijima, M. Terazima, "High Hydrostatic Pressure Induces Counterclockwise to Clockwise Reversals of the *Escherichia coli* Flagellar Motor", *J. Bacteriol.*, **195**(8), 1809-1814, (2013), 10.1128/JB.02139-12
 27. K. Suda, M. Terazima, *Y. Kimura, "Anomalous Ground-state Proton Transfer of 4'-N,N-diethylamino-3-hydroxyflavone in Ionic Liquids of Imidazolium-based Cations with Tetrafluoroborate", *Chem. Commun.*, **49**(38), 3976-3978, (2013), 10.1039/c3cc40943k
 28. K. Suda, M. Terazima, H. Sato, *Y. Kimura, "Excitation Wavelength Dependence of Excited State Intramolecular Proton Transfer Reaction of 4'-N,N-Diethylamino-3-hydroxyflavone in Room

- Temperature Ionic Liquids Studied by Optical Kerr Gate Fluorescence Measurement”, *J. Phys. Chem. B*, **117**(41), 12567-12582, (2013), 10.1021/jp405537c
29. K. Takeda, Y. Nakasone, K. Zikihara, S. Tokutomi, *M. Terazima, “Dynamics of the Amino-Terminal and Carboxyl-Terminal Helices of Arabidopsis Phototropin 1 LOV2 Studied by the Transient Grating”, *J. Phys. Chem. B*, **117**(49), 15606-15613, (2013), 10.1021/jp406109j
- 【内藤晶】
1. *T. Asakura, Y. Tasei, H. Matsuda, A. Naito, “Dynamics of Alanine Methyl Groups in Alanine Oligopeptides and Spider Dragline Silks with Different Packing Structures as Studied by ^{13}C Solid-state NMR Relaxation”, *Macromolecules*, in press, (2018), 10.1021/acs.macromol.8b01402
 2. A. Naito, Y. Tasei, A. Nishiura, T. Asakura, “Unusual Dynamics of Alanine Residues in Polyalanine Region with Staggered Packing Structure of Samia Cynthia Ricini Silk Fiber in Dry and Hydrated States Studied by ^{13}C Solid-state NMR and Molecular Dynamics Simulation”, *J. Phys. Chem. B*, **122**, 6511-6520, (2018), 10.1021/acs.jpcb.8b03509,
 3. Y. Makino, I. Kawamura, T. Okitsu, A. Wada, N. Kamo, Y. Sudo, K. Ueda, A. Naito, “Retinal Configuration of ppR Intermediates Revealed by Photo-irradiation Solid-state NMR and DFT”, *Biophys. J.*, **115**, 72–83, (2018), 10.1016/j.bpj.2018.05.030
 4. A. Naito, K. Okushita, K. Nishimura, G. B. Boutis, A. Aoki, *T. Asakura, “Quantitative Analysis of Solid-state Homonuclear Correlation Spectra of Antiparallel β -sheet Alanine Tetramers”, *J. Phys. Chem. Chem. B*, **122**, 2715-2724, (2018), 10.1021/acs.jpcb/b11126
 5. B. Mijiddorj, S. Kaneda, H. Sato, Y. Kitahashi, N. Javkhlanugs, A. Naito, K. Ueda, *I. Kawamura “The Role of D-allo-isoleucine in the Deposition of the Anti-Leishmania Peptide Bombinin H4 as Revealed by ^{31}P Solid-state NMR, VCD Spectroscopy, and MD Simulation”, *BBA-Proteins and Proteomics*, **1866**, 789-798 (2018), 10.1016/j.bbapap.2018.01.005
 6. 上田和義, *内藤晶, 川村出, “生体膜内でのペプチドの挙動の分子動力学シミュレーション”, 酵素工学ニュース, **79**, 27-30, (2018)
 7. *内藤晶, “固体高分解能 NMR スペクトルに現れる異方性相互作用 –Narrow is beautiful-”, 分光研究, **67**, 13-15, (2018)
 8. *A. Naito, N. Matsumori, A. Ramamoorthy, “Dynamic Membrane Interactions of Antibacterial and Antifungal Biomolecules, and Amyloid Peptides, Revealed by Solid-state NMR Spectroscopy”, *Biochim. Biophys. Acta (BBA)*, **1862**, 307-323, (2018), [10.1016/j.bbagen.2017.06.004](https://doi.org/10.1016/j.bbagen.2017.06.004)
 9. *T. Asakura, A. Nishimura, S. Kametani, S. Kawanishi, A. Aoki, F. Suzuki, H. Kaji, A. Naito, “Refined Crystal Structure of Samia Cynthia Silk Fibroin Revealed by Solid-state NMR Investigations”, *Biomacromolecules*, **18**, 1965-1974, (2017), [10.1021/acs.biomac.7b00441](https://doi.org/10.1021/acs.biomac.7b00441)
 10. A. Naito, Y. Tasei, A. Nishimura, *T. Asakura. “Packing Arrangements and Intersheet Interaction of Alanine Oligopeptides as Revealed by Relaxation Parameters Obtained from High-resolution ^{13}C solid-state NMR”, *J. Phys. Chem. B*, **121**(38), 8946-8955, (2017), [10.1021/acs.jpcb.7b07068](https://doi.org/10.1021/acs.jpcb.7b07068)
 11. K. Kamgar-Parsi, L. Hong, A. Naito, C.L. Brooks III, *A. Ramamoorthy, “Growth-incompetent Monomers of Human Calcitonin Lead to a Noncanonical Direct Relationship between Peptide Concentration and Aggregation Lag Time”. *J. Biol. Chem.*, **292**, 14963-14976, (2017), [10.1074/jbc.M117.791236](https://doi.org/10.1074/jbc.M117.791236)
 12. K. Norisada, N. Javkhlanugs, D. Mishima, I. Kawamura, H. saito, K. Ueda, *A. Naito, “Dynamic Structure and Orientation of Melittin Bound to Acidic Lipid Bilayers, as Revealed by Solid-state NMR and Molecular Dynamic Simulation”, *J. Phys. Chem. B*, **121**, 1802-1811, (2017), [10.1021/acs.jpcb.6b11207](https://doi.org/10.1021/acs.jpcb.6b11207)
 13. *T. Asakura, K. Miyazawa, Y. Tasei, S. Kametani, Y. Nakazawa, A. Aoki, A. Naito, “Packing Arrangement of ^{13}C selectively Labeled Sequence Model Peptides of Samia Cynthia Ricini Silk Fibroin Fiber Studied by Solid-state NMR”, *Phys. Chem. Chem. Phys.*, **19**, 13379-13386, (2017), 10.1039/C7CP01199G
 14. T. Asakura, K. Horiguchi, A. Aoki, Y. Tasei, A. Naito, “Parallel β -sheet Structure of Alanine Tetrapeptide in the Solid State as Studied by Solid-state NMR Spectroscopy”, *J. Phys. Chem. B*, **120**, 8912-8941, (2016), [10.1021/acs.jpcb.6b06292](https://doi.org/10.1021/acs.jpcb.6b06292)
 15. K. Kamgar-Parsi, J. Tolchard, B. Habenstein, A. Loquest, A. Naito, *A. Ramamoorthy, “Structural Biology of Calcitonin: from Aqueous Therapeutic Properties to Amyloid Aggregation,” *Isr. J. Chem.*, (2016), **57**, 634 – 65, [10.1002/ijch.201600096](https://doi.org/10.1002/ijch.201600096)
 16. *H. Yoshitake, T. Kodate, T.Takagi, I. Kawamura, A. Naito, “Polysilsesquioxanes with Mixed Self-

- assembled Organic Tethers: Alkyl Chains and Alkanoate-aminopropyl Pairs.” *React. Funct. Polym.*, **99**, 9-16, (2016), 10.1016/j.reactfunctpolym.2015.12.002
17. K. Oshima, A. Shigeta, Y. Makino, *I. Kawamura, T. Okitsu, A. Wada, S. Tuzi, T. Iwasa, *A. Naito, “Characterization of Photo-intermediates in the Photo-reaction Pathways of a Bacteriorhodopsin Y185F Mutant Using in Situ Photo-irradiation Solid-state NMR Spectroscopy”, *Photoche. Photobiol. Sci.*, **14**, 1694-1702, (2015), 10.1039/c5pp00154d
 18. T. Nagao, D. Mishima, N. Javkhlanlugs, J. Wang, D. Ishioka, K. Yokota, K. Norisada, I. Kawamura, K. Ueda, *A. Naito, “Structure and Orientation of Antibiotic Peptide Alamethicin in Phospholipid Bilayers as Revealed by Chemical Shift Oscillation Analysis of Solid State Nuclear Magnetic Resonance and Molecular Dynamics Simulation”, *Biochim. Biophys. Acta*, **1848**, 2789-2798, (2015) ,10.1016/j.bbamem.2015.07.019
 19. Y. Tasei, F. Tanigawa, *I. Kawamura, T. Fujito, M. Sato, A. Naito, “The Microwave Heating Mechanism of N-(4-methoxybenzyliden)-4-butylaniline in Liquid Crystalline and Isotropic Phases as Determined Using in Situ Microwave Irradiation NMR Spectroscopy”, *Phys. Chem. Chem. Phys.*, **17**, 9082-9089, (2015), 10.1039/c5cp00476d
 20. Y. Tasei, T. Yamakami, I. Kawamura, T. Fujito, K. Ushida, M. Sato, *A. Naito, “Mechanism for Microwave Heating of 1-(4'-cyanophenyl)-4-propylcyclohexane Characterized by in Situ Microwave Irradiation NMR Spectroscopy.” *J. Magn. Reson.*, **254**, 27-34, (2015), 10.1016/j.jmr.2015.02.002
 21. H. Yomoda, Y. Makino, Y. Tomonaga, T. Hidaka, *I. Kawamura, T. Okitsu, A. Wada, *Y. Sudo, *A. Naito, “Color-Discriminating Retinal Configurations of Sensory Rhodopsin I by Photo-Irradiation Solid-State NMR Spectroscopy” *Angew. Chem. Int. Ed.* **53**, 6960-6964, (2014), [10.1002/anie.201309258](https://doi.org/10.1002/anie.201309258)
 22. A. Kira, N. Javkhlanlugs, T. Miyamori, Y. Sasaki, M. Eguchi, I. Kawamura, K. Ueda, *A. Naito, “Interaction of Extracellular Loop II of κ-Opioid Receptor (196-228) with Opioid Peptide Dynorphin in Membrane Environments as Revealed by Solid State Nuclear Magnetic Resonance, Quartz Crystal Microbalance and Molecular Dynamics Simulation”, *J. Phys. Chem. B*, **118**, 9604-9612, (2014), [10.1021/jp505412j](https://doi.org/10.1021/jp505412j)

【東雅大】

1. *N. Yoshida, *M. Higashi, H. Motoki, S. Hirota, “Theoretical Analysis of the Domain-swapped Dimerization of Cytochrome *c*: An MD and 3D-RISM Approach,” *J. Chem. Phys.*, **148**, 025102,(7 pages), (2018), 10.1063/1.5009785
2. 根木秀佳, 吉田紀生, 廣田俊, 東雅大, “シトクロム *c* の多量体形成に関する理論的研究”, *J. Comput. Chem. Jpn.*, **17**, 8-13, (2018), 10.2477/jccj.2018-0006
3. H. Tsukamoto, M. Higashi, H. Motoki, H. Watanabe, C. Ganser, K. Nakajo, Y. Kubo, T. Uchihashi, *Y. Furutani, “Structural Properties Determining Low K⁺ affinity of the Selectivity Filter in the TWIK1 K⁺ Channel,” *J. Biol. Chem.*, **293**, 6969-6984, (2018), 10.1074/jbc.RA118.001817
4. *H. Okumura, M. Higashi, Y. Yoshida, H. Sato, R. Akiyama, ‘Theoretical Approaches for Dynamical Ordering of Biomolecular Systems”, *Biochim. Biophys. Acta, (BBA)*, **1862**, 212-228, (2018), [10.1016/j.bbagen.2017.10.001](https://doi.org/10.1016/j.bbagen.2017.10.001)
5. *S. Arimitsu, T. Yonamine, *M. Higashi, ‘Cinchona-Based Primary Amine Catalyzed a Proximal Functionalization of Dienamines: Asymmetric α-Fluorination of α-Branched Enals”, *ACS Catal.*, **7**, 4736, (2017), [10.1021/acscatal.7b01178](https://doi.org/10.1021/acscatal.7b01178)
6. P. Ahmadi, M. Higashi, N. J. de Voogd, *J. Tanaka, “Two Furanosesterterpenoids from the Sponge *Luffariella variabilis*,” *Mar. Drugs.*, **15**, 249, (2017), [10.3390/md15080249](https://doi.org/10.3390/md15080249)
7. I. Hermawan, A. Furuta, M. Higashi, Y. Fujita, N. Akimitsu, A. Yamashita, K. Moriishi, S. Tsuneda, H. Tani, M. Nakakoshi, M. Tsubuki, Y. Sekiguchi, *N. Noda, *J. Tanaka, “Four Aromatic Sulfates with an Inhibitory Effect against HCV NS3 Helicase from the Crinoid *Alloeocomatella polycladlia*”, *Mar. Drugs*, **15**, 117, (2017), [10.3390/md15040117](https://doi.org/10.3390/md15040117)
8. S. Tanimoto, *M. Higashi, *N. Yoshida, H. Nakano, “The Ion-dependence of Carbohydrate Binding of CBM36: MD and 3D-RISM Study”, *J. Phys., Condens. Matter*, **28**, 344005, (2016), [10.1088/0953-8984/28/34/344005](https://doi.org/10.1088/0953-8984/28/34/344005)
9. K. Watanabe, *N. Nakatani, A. Nakayama, M. Higashi, *J. Hasegawa, “Spin-blocking Effect in CO and H₂ Binding Reactions to Molybdenocene and Tungstenocene: A theoretical Study on the Reaction Mechanism via Minimum Energy Intersystem-crossing Point”, *Inorg. Chem.*, **55**, 8082-8090, (2016), [10.1021/acs.inorgchem.6b01187](https://doi.org/10.1021/acs.inorgchem.6b01187)

10. *M. Higashi, *S. Saito, "Quantitative Evaluation of Site Energies and their Fluctuations of Pigments in the Fenna–Matthews–Olson Complex with an Efficient Method for Generating a Potential Energy Surface", *J. Chem. Theory Comput.*, **12**, 4128-4137, (2016), [10.1021/acs.jctc.6b00516](https://doi.org/10.1021/acs.jctc.6b00516)
11. Y. Toma, M. Kunigami, K. Watanabe, *M. Higashi, *S. Arimitsu, "One-pot Synthesis and Theoretical Calculation for Trifluoromethylated Pyrrolizidines by 1,3-dipolar Cycloaddition with Azomethine Ylides and β -trifluoromethyl Acrylamides", *J. Fluor. Chem.*, **22**, 189, (2016), [10.1016/j.jfluchem.2016.07.013](https://doi.org/10.1016/j.jfluchem.2016.07.013)
12. A. Agena, S. Iuchi, M. Higashi, "Theoretical Study on Photoexcitation Dynamics of a Bis-diimine Cu(I) Complex in Solutions", *Chem. Phys. Lett.*, **679**, 60-65, (2016), [10.1016/j.cplett.2017.04.082](https://doi.org/10.1016/j.cplett.2017.04.082)

【松村浩由】

1. K. Nii, *M. Maruyama, S. Okada, H. Adachi, K. Takano, S. Murakami, H. Y. Yoshikawa, H. Matsumura, T. Inoue, M. Imanishi, K. Tsukamoto, M. Yoshimura, Y. Mori, "Improvement of Metastable Crystal of Acetaminophen via Control of Crystal Growth Rate," *Appl. Phys. Express.*, **11**, 035501, (2018), [10.7567/APEX.11.035501](https://doi.org/10.7567/APEX.11.035501)
2. D. Murata, H. Okano, C. Angkawidjaja, M. Akutsu, S-i. Tanaka, K. Kitahara, T. Yoshizawa, H. Matsumura, Y. Kado, E. Mizohata, T. Inoue, S. Sano, Y. Koga, S. Kanaya, K. Takano, "Structural Basis for the *Serratia Marcescens* Lipase Secretion System: Crystal Structures of the Membrane Fusion Protein and Nucleotide-binding Domain", *Biochemistry*, **56**, 6281-6291, (2017), [10.1021/acs.biochem.7b00985](https://doi.org/10.1021/acs.biochem.7b00985)
3. J. Fujita, Y. Maeda, E. Mizohata, T. Inoue, M. Kaul, A. K. Parhi, E. J. LaVoie, *D. S. Pilch, H. Matsumura*, "Structural Flexibility of an Inhibitor Overcomes Drug Resistance Mutations in *Staphylococcus aureus* FtsZ", *ACS Chem. Biol.*, **12**(7), 1947-1955, (2017), [10.1021/acschembio.7b00323](https://doi.org/10.1021/acschembio.7b00323)
4. H. Kajiura, N. Suzuki, Y. Tokumoto, T. Yoshizawa, S. Takeno, K. Fujiyama, Y. Kaneko, H. Matsumura, Y. Nakazawa, "Two *Eucommia* Farnesyl Diphosphate Synthases Exhibit Distinct Enzymatic Properties Leading to End Product Preferences", *Biochimie*, **139**, 95-106, (2017), [10.1016/j.biochi.2017.05.001](https://doi.org/10.1016/j.biochi.2017.05.001)
5. J. Fujita, *R. Harada, Y. Maeda, Y. Saito, E. Mizohata, T. Inoue, Y. Shigeta, *H. Matsumura, "Identification of the Key Interactions in Structural Transition Pathway of FtsZ from *Staphylococcus aureus*", *J. Struct. Biol.*, **198**, 65-73, (2017), [10.1016/j.jsb.2017.04.008](https://doi.org/10.1016/j.jsb.2017.04.008)
6. T. Kono, S. Mehrotra, C. Endo, N. Kizu, M. Matsuda, H. Kimura, E. Mizohata, T. Inoue, T. Hasunuma, A. Yokota, *H. Matsumura, *H. Ashida, "A RuBisCO-mediated Carbon Metabolic Pathway in Methanogenic Archaea", *Nature Commun.*, **8**(14007), 1-12 (2017), [10.1038/ncomms14007](https://doi.org/10.1038/ncomms14007)
7. *T. Sato, T. Kawasaki, S. Mine, H. Matsumura, "Functional role of the C-terminal Amphipathic Helix 8 of Olfactory Receptors and Other G Protein-coupled Receptors", *Int. J. Mol. Sci.*, **17**(11), 1930, (2016), [10.3390/ijms17111930](https://doi.org/10.3390/ijms17111930)
8. Y. Tominaga, M. Maruyama, M. Yoshimura, H. Koizumi, M. Tachibana, S. Sugiyama, H. Adachi, K. Tsukamoto, H. Matsumura, K. Takano, S. Murakami, T. Inoue, *H. Y. Yoshikawa, Y. Mori, "Promotion of Protein Crystal Growth by Actively Switching Crystal Growth Mode via Femtosecond Laser Ablation", *Nature Photonics*, **10**, 723-726, (2016), [10.1038/nphoton.2016.202](https://doi.org/10.1038/nphoton.2016.202)
9. *M. Maruyama, Y. Hayashi, H. Y. Yoshikawa, S. Okada, H. Koizumi, M. Tachibana, S. Sugiyama, H. Adachi, H. Matsumura, T. Inoue, K. Takano, S. Murakami, M. Yoshimura, Y. Mori, "A Crystallization Technique for Obtaining Large Protein Crystals with Increased Mechanical Stability Using Agarose Gel Combined with a Stirring Technique", *J. Crystal Growth*, **452**, 172-178, (2016), [10.1016/j.jcrysgro.2015.11.008](https://doi.org/10.1016/j.jcrysgro.2015.11.008)
10. S. Sugiyama, S. Ishikawa, H. Tomitori, M. Niyyama, M. Hirose, Y. Miyazaki, K. Higashi, M. Murata, H. Adachi, K. Takano, S. Murakami, T. Inoue, Y. Mori, K. Kashiwagi, K. Igarashi, *H. Matsumura, "Molecular Mechanism Underlying Promiscuous Polyamine Recognition by Spermidine Acetyltransferase", *Int. J. Biochem. Cell Biol.*, **76**, 87-97, (2016), [10.1016/j.biocel.2016.05.003](https://doi.org/10.1016/j.biocel.2016.05.003)
11. *T. Sato, R. Kobayakawa, K. Kobayakawa, M. Emura, S. Itohara, T. Kawasaki, A. Tsuboi, H. Matsumura, "Supersensitive Odor Discrimination is Controlled in Part by Initial Transient Interactions between the Most Sensitive Dorsal Olfactory Receptors and G-proteins", *Receptors Clin. Invest.*, **3**, e1117, (2016), [10.14800/rci.1117](https://doi.org/10.14800/rci.1117)
12. M. Matsuoka, K. Kakinouchi, H. Adachi, M. Maruyama, S. Sugiyama, I. Nakabayashi, H. Tsuchikura, A. Kuwahara, S. Sano, H. Y. Yoshikawa, Y. Takahashi, M. Yoshimura, H. Matsumura, S. Murakami, T. Inoue, Y. Mori, *K. Takano "Growth of High-strength Protein Crystals with Nanofibers", *Appl. Phys. Express*, **9**, 035503, (2016), [10.7567/APEX.9.035503](https://doi.org/10.7567/APEX.9.035503)

13. Y. Kado, E. Mizohata, S. Nagatoishi, M. Iijima, K. Shinoda, T. Miyafusa, T. Nakayama, T. Yoshizumi, A. Sugiyama, T. Kawamura, Y-H Lee, H. Matsumura, H. Doi, H. Fujitani, T. Kodama, Y. Shibasaki, K. Tsumoto, *T. Inoue, "Epiregulin Recognition Mechanisms by Anti-epiregulin Antibody 9E5: Structural, Functional and Molecular Dynamics Simulation Analyses", *J. Biol. Chem.*, **291**, 2319-2330, (2016), 10.1074/jbc.M115.656009
14. Y. Fukuda , K. M. Tse , M. Suzuki, K. Diederichs, K. Hirata, T. Nakane, M. Sugahara, E. Nango, K. Tono, Y. Joti, T. Kameshima, C. Song, T. Hatsui, M. Yabashi, O. Nureki, H. Matsumura, T. Inoue, S. Iwata, *E. Mizohata, "Redox-coupled Structural Changes in Nitrite Reductase Revealed by Serial Femtosecond and Microfocus Crystallography", *J. Biochem.*, **59**(5):527-38, (2016), 10.1093/jb/mvv133
15. Y. Fukuda, K. M. Tse, Y. Kado, E. Mizohata, H. Matsumura, *T. Inoue "Insights into Unknown Foreign Ligand in Copper Nitrite Reductase", *Biochem. Biophys. Res. Commun.*, **464**, 622, (2015), 10.1016/j.bbrc.2015.07.025
16. A. Fujii, Y. Sekiguchi, *H. Matsumura, T. Inoue, *W-S. Chung, S. Hirota, *T. Matsuo " Excimer Emission Properties on Pyrene-labeled Protein Surface: Correlation between Emission Spectra, Ring Stacking Modes, and Flexibilities of Pyrene Probes", *Bioconjugate Chem.*, **26**, 537, (2015), 10.1021/acs.bioconjchem.5b00026
17. T. Kawasaki, T. Saka, S. Mine, E. Mizohata, T. Inoue, *H. Matsumura, T. Sato "The N-terminal Acidic Residue of the Cytosolic Helix 8 of an Odorant Receptor is Responsible for Different Response Dynamics via G-protein", *FEBS Lett.*, **589**, 1136, (2015), 10.1016/j.febslet.2015.03.025
18. R. N. Abd Rahman, M.S. Ali, S. Sugiyama, A.T. Leow, T. Inoue, M. Basri, A.B. Salleh, *H. Matsumura, "A Comparative Analysis of Microgravity and Earth Grown Thermostable T1 Lipase Crystals using HDPCG Apparatus", *Protein Pept. Lett.*, **22**, 173, (2015), 10.2174/0929866521666141019193604
19. *N. Maruyama, T. Goshi, S. Sugiyama, M. Niyyama, H. Adachi, K. Takano, S. Murakami, T. Inoue, Y. Mori, H. Matsumura, B. Mikami " Preliminary X-ray Analysis of the Binding Domain of the Soybean Vacuolar Sorting Receptor Complexed with a Sorting Determinant of a Seed Storage Protein", *Acta Cryst.*, F71, **132**, (2015), 10.1107/S2053230X14027484
20. J. Fujita, Y. Maeda, C. Nagao, Y. Tsuchiya, Y. Miyazaki, M. Hirose, E. Mizohata, Y. Matsumoto, T. Inoue, K. Mizuguchi, *H. Matsumura, "Crystal Structure of FtsA from *Staphylococcus Aureus*", *FEBS Lett.*, **588**(10), 1879-1885, (2014), 10.1016/j.febslet.2014.04.008
21. K. Ishibashi, Y. Kezuka, C. Kobayashi, M. Kato, T. Inoue, T. Nonaka, M. Ishikawa, *H. Matsumura, *Etsuko Katoh , " Structural Basis for the Recognition-evasion Arms Race between Tomato Mosaic Virus and the Resistance Gene Tm-1", *Proc. Natl. Acad. Sci. USA*, **111**, E3486, (2014), 10.1073/pnas.1407888111

【松森信明】

1. 西村慎一、掛谷秀昭、松森信明、“生体膜を標的にする天然有機化合物”，*化学と生物*, (2018), 10月号, in press
2. K. Cornelio, RA. Espiritu, S. Hanashima, Y. Todokoro, R. Malabed, M. Kinoshita, *N. Matsumori, *M. Murata, S. Nishimura, H. Kakeya, M. Yoshida, S. Matsunaga, "Theonellamide A, a Marine-sponge-derived Bicyclic Peptide, Binds to Cholesterol in Aqueous DMSO: Solution NMR-based Analysis of Peptide-sterol Interactions Using Hydroxylated Sterol." *Biochim. Biophys. Acta (BBA)-Biomembranes*, (2018), in press, 10.1016/j.bbamem.2018.07.010
3. *M. Kinoshita, KGN. Suzuki, M. Murata, *N. Matsumori, "Evidence of Lipid Rafts based on the Partition and Dynamic Behavior of Sphingomyelins." *Chem. Phys. Lipids.*, **215**, 84-95, (2018), 10.1016/j.chemphyslip.2018.07.002
4. Y. Umegawa, N. Matsumori, *M. Murata, "Recent Solid-State NMR Studies of Hydrated Lipid Membranes." *Annu. Rep. NMR Spectrosc.*, **94**, 41-72, (2018), 10.1016/bs.arnmr.2017.12.003
5. Y. Wakamiya, M. Ebine, M. Murayama, H. Omizu, N. Matsumori, M. Murata, *T. Oishi, "Synthesis and Stereochemical Revision of the C31-C67 Section of Amphidinol 3" *Angew. Chem. Int. Ed. Engl.*, **130**, 6168-6172 , (2018), 10.1002/anie.201712167
6. T. Watanabe, H. Shibata, M. Ebine, H. Tsuchikawa, N. Matsumori, M. Murata, M. Yoshida, M. Morisawa, S. Lin, K. Yamauchi, K. Sakai, *T. Oishi, "Synthesis and Complete Structure Determination of a Sperm-Activating and -Attracting Factor Isolated from the Ascidian *Ascidia Sydneiensis*." *J. Nat. Prod.*, **81**, 985–997, (2018), 10.1021/acs.jnatprod.7b01052

7. T. Matsufuji, M. Kinoshita, A. Möuts, J. P. Slotte, *N. Matsumori, "Preparation and Membrane Properties of Oxidized Ceramide Derivatives", *Langmuir*, **34**, 465-471, (2018), 10.1021/acs.langmuir.7b02654
8. *A. Naito, N. Matsumori, A. Ramamoorthy, "Dynamic Membrane Interactions of Antibacterial and Antifungal Biomolecules, and Amyloid Peptides, Revealed by Solid-state NMR Spectroscopy." *Biochim. Biophys. Acta.*, **1862**, 307-323, (2018), 10.1016/j.bbagen.2017.06.004
9. Y. Umegawa, N. Matsumori, *M Murata, "Recent Solid-State NMR Studies of Hydrated Lipid Membranes", *Annual Reports on NMR Spectroscopy*, **94**, 41-72, (2018), 10.1016/bs.arnmr.2017.12.003
10. M. Kinoshita, H. Ano, M. Murata, K. Shigetomi, J. Ikenouchi, *N. Matsumori, "Emphatic Visualization of Sphingomyelin-rich Domains by Inter-lipid FRET Imaging Using Fluorescent Sphingomyelins." *Sci. Rep.*, **7**, 16801, (2017), 10.1038/s41598-017-16361-x
11. *R. Ohtani, T. Tokita, T. Takaya, *K. Iwata, M. Kinoshita, N. Matsumori, M. Nakamura, L. F. Lindoy, *S. Hayami, "The Impact of Metal Complex Lipids on Viscosity and Curvature of Hybrid Liposomes." *Chem. Commun.*, **53**, 13249-13252, (2017), 10.1039/c7cc07944c
12. M. Iwamoto, A. Sumino, E. Shimada, M. Kinoshita, N. Matsumori, *S. Oiki, "Channel Formation and Membrane Deformation via Sterol-Aided Polymorphism of Amphidinol 3" *Sci. Rep.*, **7**, 10782, (2017), 10.1038/s41598-017-11135-x.
13. *M. Satake, K. Cornelio, S. Hanashima, R. Malabed, M. Murata, N. Matsumori, H. Zhang, F. Hayashi, S. Mori, J.S. Kim, C.H. Kim, J.S. Lee, "Structures of the Largest Amphidinol Homologues from the Dinoflagellate Amphidinium carterae and Structure-Activity Relationships." *J. Nat. Prod.*, **80**, 2883-2888, (2017), 10.1021/acs.jnatprod.7b00345
14. 松森信明, 「脂質ラフトのNMR解析」, 『日本核磁気共鳴学会機関紙』, **8**, 9-16 (2017)
15. M. Kinoshita, K.G.N. Suzuki, *N. Matsumori, M. Takada, H. Ano, K. Morigaki, M. Abe, A. Makino, T. Kobayashi, K.M. Hirosawa, T. K Fujiwara, *A. Kusumi, M. Murata, "Raft-based Sphingomyelin Interactions Revealed by New Fluorescent Sphingomyelin Analogs", *J Cell Biol.*, **216**, 1183, (2017), 10.1083/jcb.201607086
16. T. Nakane, S. Hanashima, M. Suzuki, H. Saiki, T. Hayashi, K. Kakinouchi, S. Sugiyama, S. Kawatake, S. Matsuoka, N. Matsumori, E. Nango, J. Kobayashi, T. Shimamura, K. Kimura, C. Mori, N. Kunishima, M. Sugahara, Y. Takakyu, S. Inoue, T. Masuda, T. Hosaka, K. Tono, Y. Joti, T. Kameshima, T. Hatsui, M. Yabashi, T. Inoue, O. Nureki, S. Iwata, *M. Murata, *E. Mizohata, "Membrane Protein Structure Determination by SAD, SIR, or SIRAS Phasing in Serial Femtosecond Crystallography Using an Iododetergent", *Proc. Natl. Acad. Sci. USA*, **113**, 13039, (2016), 10.1073/pnas.1602531113
17. K. Cornelio, R.A. Espiritu, Y. Todokoro, S. Hanashima, M. Kinoshita, N. Matsumori, *M. Murata, S. Nishimura, H. Kakeya, M. Yoshida, S. Matsunaga, "Sterol-dependent Membrane Association of the Marine Sponge-derived Bicyclic Peptide Theonellamide A as Examined by ¹H NMR", *Bioorg. Med. Chem.*, **24**, 5235, (2016), 10.1016/j.bmc.2016.08.043
18. R.A. Espiritu, K. Cornelio, M. Kinoshita, *N. Matsumori, M. Murata, S. Nishimura, H. Kakeya, M. Yoshida, S. Matsunaga, "Marine Sponge Cyclic Peptide Theonellamide A Disrupts Lipid Bilayer Integrity without Forming Distinct Membrane Pores", *Biochim Biophys Acta Biomembranes*, **1858**, 1373, (2016), 10.1016/j.bbamem.2016.03.019
19. Y. Nakagawa, Y. Umegawa, N. Matsushita, T. Yamamoto, H. Tsuchikawa, S. Hanashima, T. Oishi, *N. Matsumori, *M. Murata, "The Structure of the Bimolecular Complex between Amphotericin B and Ergosterol in Membranes is Stabilized by Face-to-face Van Der Waals Interaction with their Rigid Cyclic Cores", *Biochemistry*, **55**, 3392, (2016), 10.1021/acs.biochem.6b00193
20. O. Engberg, T. Yasuda, V. Hautala, N. Matsumori, T.K.M. Nyholm, *M. Murata, *J. P. Slotte, "Lipid interactions and organization in complex bilayer Membranes", *Biophys J*, **110**, 1563, (2016), 10.1016/j.bpj.2015.12.043
21. Y. Umegawa, Y. Tanaka, N. Matsumori, *M. Murata, "¹³C-TmDOTA as Versatile Thermometer Compound for Solid-state NMR of Hydrated Lipid Bilayer Membranes", *Magn Reson Chem.*, **54**, 227, (2016), 10.1002/mrc.4371
22. J. Cui, S. Kawatake, Y. Umegawa, S. Lethu, M. Yamagami, S. Matsuoka, F. Sato, N. Matsumori, *M. Murata, "Stereoselective Synthesis of the Head Group of Archaeal Phospholipid PGP-Me to Investigate Bacteriorhodopsin-lipid Interactions," *Org. Biomol. Chem.*, **13**, 10279 (2015), 10.1039/c5ob01252j

23. T. Yasuda, N. Matsumori, H. Tsuchikawa, M. Lonnfors, T. K. M. Nyholm, J. P. Slotte, *M. Murata, "Formation of Gel-like Nanodomains in Cholesterol-Containing Sphingomyelin or Phosphatidylcholine Binary Membrane As Examined by Fluorescence Lifetimes and H-2 NMR Spectra", *Langmuir*, **31**, 13783, (2015), 10.1021/acs.langmuir.5b03566
24. H. Shibata, H. Tsuchikawa, T. Hayashi, N. Matsumori, *M. Murata, T. Usui, "Modification of Bafilomycin Structure to Efficiently Synthesize Solid-state NMR Probes that Selectively Bind to Vacuolar-type ATPase", *Chem. Asian J.* **10**, 915-924, (2015), 10.1002/asia.201403299
25. J. Cui, S. Matsuoka, M. Kinoshita, N. Matsumori, F. Sato, *M. Murata, J. Ando, H. Yamakoshi, K. Dodo, M. Sodeoka, "Novel Raman-tagged Sphingomyelin that Closely Mimics Original raft-forming Behavior", *Bioorg. Med. Chem.* **23**, 2989-2994, (2015), 10.1016/j.bmc.2015.05.014
26. *N. Matsumori, T. Yamaguchi, Y. Maeta, M. Murata, "Orientation and Order of the Amide Group of Sphingomyelin in Bilayers Determined by Solid-State NMR", *Biophys. J.* **108**, 2816-2824, (2015), 10.1016/j.bpj.2015.05.011
27. *M. Murata, S. Sugiyama, S. Matsuoka, N. Matsumori, "Bioactive Structure of Membrane Lipids and Natural Products Elucidated by a Chemistry-Based Approach." *Chem. Rec.* **15**, 675-690, (2015), 10.1002/tcr.201402097
28. T. Yamamoto, Y. Umegawa, H. Tsuchikawa, N. Matsumori, S. Hanashima, *M. Murata, R. Haser, B.J. Rawlings, P. Caffrey P, "Role of Polyol Moiety of Amphotericin B in Ion Channel Formation and Sterol Selectivity in Bilayer Membrane", *Bioorg. Med. Chem.* **23**, 5782-5788, (2015), 10.1016/j.bmc.2015.07.009
29. T. Yasuda, H. Tsuchikawa, M. Murata, *N. Matsumori, "Deuterium NMR of Raft Model Membranes Reveals Domain-specific Order Profiles and Compositional Distribution", *Biophysical J.*, **108**, 2502-2506, (2015), 10.1016/j.bpj.2015.04.008
30. J. Cui, S. Lethu, T. Yasuda, S. Matsuoka, N. Matsumori, F. Sato, *M. Murata, "Phosphatidylcholine Bearing 6, 6-dideuterated Oleic Acid: a Useful Solid-state (2)HNMR Probe for Investigating Membrane Properties", *Bioorg. Med. Chem. Lett.*, **25**, 203, (2015), 10.1016/j.bmcl.2014.11.072
31. Y. Nakagawa, Y. Umegawa, K. Nonomura, N. Matsushita, T. Takano, H. Tsuchikawa, S. Hanashima, T. Oishi, *N. Matsumori, *M. Murata, "Axial Hydrogen at C7 Position and Bumpy Tetracyclic Core Markedly Reduce Sterol's Affinity to Amphotericin B in Membrane", *Biochemistry*, **54**, 303, (2015), 10.1021/bi5012942
32. R. A. Espiritu, *N. Matsumori, M. Tsuda, M. Murata, "Direct and Sereospecific Interaction of Amphidinol 3 with Sterol in Lipid Bilayers", *Biochemistry*, **53**(20), 3287-3293, (2014), 10.1021/bi5002932
33. Y. Nakagawa, Y. Umegawa, T. Takano, H. Tsuchikawa, *N. Matsumori, M. Murata, "Effect of Sterol Side chain on Ion Channel Formation by Amphotericin B in Lipid Bilayers", *Biochemistry*, **53**(19), 3088-3094, (2014), 10.1021/bi500122c
34. M. Kinoshita, N. Matsumori, *M. Murata, "Coexistence of Two Liquid Crystalline Phases in Dihydrosphingomyelin and Dioleoylphosphatidylcholine Binary Mixtures", *Biochim. Biophys. Acta Biomembranes*, **1838**, 1372-1381, (2014), 10.1016/j.bbamem.2014.01.017
35. R. Sugiyama, S. Nishimura, N. Matsumori, Y. Tsunematsu, A. Hattori, *H. Kakeya, "Structure and Biological Activity of 8-deoxyheronamide C from a Marine-derived Streptomyces sp.: Heronamides Target Saturated Hydrocarbon Chains in Lipid Membranes", *J. Am. Chem. Soc.*, **136**(14), 5209-5212, (2014), 10.1021/ja500128u
36. H. Shibata, H. Tsuchikawa, N. Matsumori, *M. Murata, T. Usui, "Design and Synthesis of 24-fluorinated Bafilomycin Analogue as an NMR Probe with Potent Inhibitory Activity to Vacuolar-type ATPase", *Chem. Lett.*, **43**(4), 474-476, (2014), 10.1246/cl.131099
37. *松森信明, "脂質膜における構造および相互作用解析:バイセルを用いた検討" *有機合成化学協会誌* **72**(5), 529-537, (2014)

【養田正文】

1. R. Nakamura, T. Obata, R. Nojima, Y. Hashimoto, K. Noguchi, T. Ogawa, M. Yohda "Functional Expression and Characterization of Tetrachloroethene Dehalogenase From Geobacter sp.", *Front. Microbiol.*, **9**, 1774 (2018), 10.3389/fmicb.2018.01774
2. K. Morita, Y. Y. Yamamoto, A. Hori, T. Obata, Y. Uno, K. Shinohara, K. Noguchi, K. Noi, T. Ogura, K. Ishii, K. Kato, M. Kikumoto, R. Arranz, J. M. Valpuesta, *M. Yohda "Expression, Functional Characterization, and Preliminary Crystallization of the Cochaperone Prefoldin from the Thermophilic

- Fungus *Chaetomium thermophilum*”, *Int. J. Mol. Sci.*, **19**, E2452, (2018), 10.3390/ijms19082452
3. K. Ikegami, Y. Aita, A. Shiroma, M. Shimoji, H. Tamotsu, N. Ashimine, M. Shinzato, S. Ohki, K. Nakano, K. Teruya, K. Satou, T. Hirano, *M. Yohda “Complete Genome Sequence of Petrimonas sp. Strain IBARAKI, Assembled from the Metagenome Data of a Culture Containing Dehalococcoides spp.”, *Genome Announc.*, **6**, e00384-18, (2018), 10.1128/genomeA.00384-18.
 4. N. Ogawa, Y. Y. Yamamoto, K. Abe, H. Sekiguchi, Y. C. Sasaki, A. Ishikawa, J. Frydman, *M. Yohda “Time-Resolved Measurement of the ATP-Dependent Motion of the Group II Chaperonin by Diffracted Electron Tracking”, *Int. J. Mol. Sci.*, **19**, E950, (2018), 10.3390/ijms19040950
 5. M. Sahlan, T. Zako, *M. Yohda, “Prefoldin, a Jellyfish-like Molecular Chaperone: Functional Cooperation with a Group II Chaperonin and Beyond”, *Biophys. Rev.*, **10**, 339, (2018), 10.1007/s12551-018-0400-0
 6. M. Asakawa, Y. Fukutani, A. Savangsuksa, K. Noguchi, H. Matsunami, *M. Yohda, “Modification of the Response of Olfactory Receptors to Acetophenone by CYP1a2”, *Sci. Rep.*, **7**, 10167, (2017), [10.1038/s41598-017-10862-5](https://doi.org/10.1038/s41598-017-10862-5)
 7. *M. Yohda, K. Ikegami, Y. Aita, M. Kitajima, A. Takechi, M. Iwamoto, T. Fukuda, N. Tamura, J. Shibasaki, S. Koike, D. Komatsu, S. Miyagi, M. Nishimura, Y. Uchino, A. Shiroma, M. Shimoji, H. Tamotsu, N. Ashimine, M. Shinzato, S. Ohki, K. Nakano, K. Teruya, K. Satou, T. Hirano, O. Yagi, “Isolation and Genomic Characterization of a Dehalococcoides Strain Suggests Genomic Rearrangement during Culture”, *Sci. Rep.*, **7**, 2230, (2017), [10.1038/s41598-017-02381-0](https://doi.org/10.1038/s41598-017-02381-0)
 8. Y. Y. Yamamoto, Y. Uno, E. Sha, K. Ikegami, N. Ishii, N. Dohmae, H. Sekiguchi, Y. C. Sasaki, *M. Yohda, “Asymmetry in the Function and Dynamics of the Cytosolic Group II Chaperonin CCT/TRiC”, *PLoS One*, **12**, e0176054, (2017), [10.1371/journal.pone.0176054](https://doi.org/10.1371/journal.pone.0176054)
 9. S. Sonotaki, T. Takami, K. Noguchi, M. Odaka, M. Yohda, *Y. Murakami, “Successful PEGylation of Hollow Encapsulin Nanoparticles from *Rhodococcus erythropolis* N771 without Affecting their Disassembly and Reassembly Properties”, *Biomater. Sci.*, **5**, 1082-1089, (2017), [10.1039/c7bm00207](https://doi.org/10.1039/c7bm00207)
 10. K. Hasegawa, R. Negishi, M. Matsumoto, M. Yohda, *K. Hosokawa, M. Maeda, “Specificity of MicroRNA Detection on a Power-free Microfluidic Chip with Laminar Flow-assisted Dendritic Amplification”, *Anal. Sci.* **33**, 171-177, (2017), [10.2116/analsci.33.171](https://doi.org/10.2116/analsci.33.171)
 11. M. M. Islam, M. Yohda, S. I. Kidokoro, *Y. Kuroda, “Crystal Structures of Highly Simplified BPTIs Provide Insights into Hydration-driven Increase of Unfolding Enthalpy”, *Sci. Rep.*, **7**, 41205, (2017), [10.1038/srep41205](https://doi.org/10.1038/srep41205)
 12. R. Sharma, Y. Ishimaru, I. Davison, K. Ikegami, M. S. Chien, H. You, Q. Chi, M. Kubota, M. Yohda, M. Ehlers, *H. Matsunami, “Olfactory Receptor Accessory Proteins Play Crucial Roles in Receptor Function and Gene Choice”, *Elife*, (2017) , **6**, e21895, [10.7554/elife.21895](https://doi.org/10.7554/elife.21895)
 13. Y. Fukutani, J. Ishii, A. Kondo, T. Ozawa, H. Matsunami, *M. Yohda, “Split Luciferase Complementation Assay for the Analysis of G Protein-coupled Receptor Ligand Response in *Saccharomyces cerevisiae*”, *Biotechnol. Bioeng.*, **114**(6),1354-1361, (2017), [10.1002/bit.26255](https://doi.org/10.1002/bit.26255)
 14. C. Sugimoto, K. Takeda, Y. Kariya, H. Matsumura, M. Yohda, H. Ohno, *N. Nakamura, “A method of Expression for an Oxygen-tolerant Group III Alcohol Dehydrogenase from *Pyrococcus horikoshii* OT3”, *J. Biol. Inorg. Chem.*, **22**, 527-534, (2017), [10.1007/s00775-017-1439-2](https://doi.org/10.1007/s00775-017-1439-2)
 15. K. Hakamada, H. Watanabe, R. Kawano, K. Noguchi, *M. Yohda, “Expression and Characterization of the Plasmid Translocon of the Exported Proteins Component EXP2”, *Biochem. Biophys. Res. Commun.*, **482**, 700-705, (2017), [10.1016/j.bbrc.2016.11.097](https://doi.org/10.1016/j.bbrc.2016.11.097)
 16. Y. Y. Yamamoto, K. Tsuchida, K. Noguchi, N. Ogawa, H. Sekiguchi, Y. C. Sasaki, *M. Yohda, “Characterization of Group II Chaperonins from an Acidothermophilic Archaeon *Picrophilus torridus*”, *FEBS Open Bio.*, **6**, 751-764, (2016), [10.1002/2211-5463.12090](https://doi.org/10.1002/2211-5463.12090)
 17. F. Watanabe, F. Yu, A. Ohtaki, Y. Yamanaka, K. Noguchi, M. Odaka, *M. Yohda, “Improvement of Enantioselectivity of the B-type Halohydrin Hydrogen-halide-lyase from *Corynebacterium* sp. N-1074”, *J. Biosci. Bioeng.*, **122**, 270-275, (2016), [10.1016/j.jbiosc.2016.02.003](https://doi.org/10.1016/j.jbiosc.2016.02.003)
 18. T. Aihara, T. Ito, Y. Yamanaka, K. Noguchi, M. Odaka, M. Sekine, H. Homma, *M. Yohda, “Structural and Functional Characterization of Aspartate Racemase from the Acidothermophilic Archaeon *Picrophilus torridus*”, *Extremophiles* **20**, 385-393, (2016), [10.1007/s00792-016-0829-7](https://doi.org/10.1007/s00792-016-0829-7)
 19. T. Zako, M. Sahlan, S. Fujii, Y. Y. Yamamoto, P. T. Tai, K. Sakai, M. Maeda M, *M. Yohda,

- “Contribution of the C-Terminal Region of a Group II Chaperonin to its Interaction with Prefoldin and Substrate Transfer”, *J. Mol. Biol.*, **428**, 2405-2417, (2016), [10.1016/j.jmb.2016.04.006](https://doi.org/10.1016/j.jmb.2016.04.006)
20. M. Kayanuma, M. Shoji, M. Yohda, M. Odaka, *Y. Shigeta, “Catalytic Mechanism of Nitrile Hydratase Subsequent to Cyclic Intermediate Formation: A QM/MM Study”, *J. Phys. Chem. B.*, **120**, 3259-3266, (2016), [10.1021/acs.jpcb.5b11363](https://doi.org/10.1021/acs.jpcb.5b11363)
 21. Y.Y. Yamamoto, M. Yohda, ”Thermosome: A Group II Chaperonin of Archaea”, *eLS*. **1**, (2016), 10.1002/9780470015902.a0026332
 22. M. M. Islam, S. Nakamura, K. Noguchi, M. Yohda, S. Kidokoro, *Y. Kuroda, “Analysis and Control of Protein Crystallization Using Short Peptide Tags That Change Solubility without Affecting Structure, Thermal Stability, and Function”, *Cryst Growth Des.*, **15**, 2703, (2015), [10.1021/acs.cgd.5b00010](https://doi.org/10.1021/acs.cgd.5b00010)
 23. Y. Yamanaka, Y. Kato, K. Hashimoto, K. Iida, K. Nagasawa, H. Nakayama, M. Yohda, *M. Odaka, “Time-Resolved Crystallography of the Reaction Intermediate of Nitrile Hydratase: Revealing a Role for the Cysteinesulfenic Acid Ligand as a Catalytic Nucleophile”, *Angew. Chem.*, **54**, 10763, (2015), 10.1002/anie.201502731
 24. F. Watanabe, F. Yu, A. Ohtaki, Y. Yamanaka, K. Noguchi, *M. Yohda, M. Odaka, “Crystal Structures of Halohydrin Hydrogen-halide-lyases from *Corynebacterium* sp. N-1074”, *Proteins*. **83**, 2230-2239, (2015), [10.1002/prot.24938](https://doi.org/10.1002/prot.24938)
 25. T. Arai, S. Kimata, D. Mochizuki, K. Hara, T. Zako, M. Odaka, M. Yohda, F. Arisaka, S. Kanamaru, T. Matsumoto, S. Yajima, J. Sato, S. Kawasaki, *Y. Niimura, “NADH Oxidase and Alkyl Hydroperoxide Reductase Subunit C (peroxiredoxin) from *Amphibacillus Xylanus* Form an Oligomeric Assembly”, *FEBS Open Bio*. **5**, 124, (2015), [10.1016/j.fob.2015.01.005](https://doi.org/10.1016/j.fob.2015.01.005)
 26. Y. Fukutani, A. Hori, S. Tsukada, R. Sato, J. Ishii, A. Kondo, H. Matsunami, *M. Yohda, “Improving the Odorant Sensitivity of Olfactory Receptor-expressing Yeast with Accessory Proteins”, *Anal. Biochem.* **471**, 1, (2015), [10.1016/j.ab.2014.10.012](https://doi.org/10.1016/j.ab.2014.10.012).
 27. Y. Yokoyama, A. Ohtaki, I. Jantan, M. Yohda, *H. Nakamoto , “Goniothalamin Enhances”the ATPase Activity of the Molecular Chaperone Hsp90 but Inhibits its Chaperone Activity”, *J. Biochem.* **157**, 161, (2015), [10.1093/jb/mvu061](https://doi.org/10.1093/jb/mvu061)
 28. Y. Yamamoto, Y. Abe, K. Moriya, M. Arita, K. Noguchi, N. Ishii, H. Sekiguchi, Y. C. Sasaki, *M. Yohda, “Inter-ring Communication is Dispensable in the Reaction Cycle of Group II Chaperonins”, *J. Mol. Biol.* **426**(14), 2667-2678, (2014), [10.1016/j.jmb.2014.05.013](https://doi.org/10.1016/j.jmb.2014.05.013)
 29. A. Tamura, Y. Fukutani, T. Takami, M. Fujii, Y. Nakaguchi, Y. Murakami, K. Noguchi, M. Yohda, *M. Odaka, “Packaging Guest Proteins into the Encapsulin Nanocompartment from *Rhodococcus erythropolis* N771”, *Biotechnol. Bioeng.*, **112**(1), 13-20, (2014), [10.1002/bit.25322](https://doi.org/10.1002/bit.25322)

A02 : 動的秩序の創生

【新井亮一】

1. *N. Kawakami, H. Kondo, Y. Matsuzawa, K. Hayasaka E. Nasu, K. Sasahara, R. Arai, *K. Miyamoto, “Design of Hollow Protein Nanoparticles with Modifiable Interior and Exterior Surfaces”, *Angew. Chem. Int. Ed.*, in press, (2018), [10.1002/anie.201805565](https://doi.org/10.1002/anie.201805565)
2. N. Kobayashi, K. Inano, K. Sasahara, T. Sato, K. Miyazawa, T. Fukuma, M. H. Hecht, C. Song, K. Murata, *R. Arai, ‘Self-Assembling Supramolecular Nanostructures Constructed from *de Novo* Extender Protein Nanobuilding Blocks’ *ACS Synth. Biol.*, **7**, 1381–1394, (2018), [10.1021/acssynbio.8b00007](https://doi.org/10.1021/acssynbio.8b00007)
3. *R. Arai, “Hierarchical Design of Artificial Proteins and Complexes toward Synthetic Structural Biology”, *Biophys. Rev.*, **10**, 391-410 (2018), [10.1007/s12551-017-0376-1](https://doi.org/10.1007/s12551-017-0376-1)
4. N. Kobayashi, *R. Arai, “Design and Construction of Self-assembling Supramolecular Protein Complexes Using Artificial and Fusion Proteins as Nanoscale Building Blocks.” *Curr. Opin. Biotech.*, **46**, 57-65, (2017), [10.1016/j.copbio.2017.01.001](https://doi.org/10.1016/j.copbio.2017.01.001)

【飯野亮太】

1. F. Kawai, A. Nakamura, A. Visootsat, *R. Iino, “Plasmid-based One-pot Saturation Mutagenesis and Robot-based Automated Screening for Protein Engineering”, *ACS Omega*, **3**, 7715–7726, (2018), [10.1021/acsomega.8b00663](https://doi.org/10.1021/acsomega.8b00663)
2. #T. Uchihashi, #*TH. Watanabe, Y. Nakazaki, T. Yamasaki, T. Watanabe, T. Maruno, K. Ishii, S. Uchiyama, C. Song, K. Murata, *R. Iino, *T. Ando (#Equal contribution), “Dynamic Structural States of ClpB Involved in its Disaggregation Function”, *Nature Communications*, **9**, Article number: 2147,

(2018), 10.1038/s41467-018-04587-w

3. A. Nakamura , T. Tasaki, Y. Okuni, C. Song, K. Murata, T. Kozai, M. Hara, H. Sugimoto, K. Suzuki, T. Watanabe, T. Uchihashi, H. Noji, *R. Iino, “Rate Constants, Processivity, and Productive Binding Ratio of Chitinase A Revealed by Single-molecule Analysis”, *Phys. Chem. Chem. Phys.*, **20**, 3010-3018, (2018), [10.1039/C7CP04606E](https://doi.org/10.1039/C7CP04606E)
4. *R. Iino, T. Iida, A. Nakamura, E. Saita, *H. You H, *Y. Sako, “Single-molecule Imaging and Manipulation of Biomolecular Machines and Systems”, *Biochim. Biophys. Acta, (BBA), Gen. Subj.*, **1862**, 241-252, (2017), [10.1016/j.bbagen.2017.08.008](https://doi.org/10.1016/j.bbagen.2017.08.008)
5. #M. Baba, #K. Iwamoto, R. Iino, H. Ueno, M. Hara, A. Nakanishia, J. Kishikawa, *H. Noji, *K. Yokoyama (#Equal contribution), “Rotation of Artificial Rotor Axles in Rotary Molecular Motors”, *Proc Natl Acad Sci USA*, **113**, 11214-11219, (2016), [10.1073/pnas.1605640113](https://doi.org/10.1073/pnas.1605640113)
6. A. Nakamura, T. Tasaki, D. Ishiwata, M. Yamamoto, Y. Okuni, A. Visootsat, M. Maximilien, H. Noji, T. Uchiyama, M. Samejima, K. Igarashi, *R. Iino, “Single-molecule Imaging Analysis of Binding, Processive Movement, and Dissociation of Cellobiohydrolase *Trichoderma reesei* Cel6A and its Domains on Crystalline Cellulose”, *J. Biol. Chem.*, **291**, 22404-22413, (2016), [10.1074/jbc.M116.752048](https://doi.org/10.1074/jbc.M116.752048)
7. H. Isojima, R. Iino, Y. Niitani, H. Noji, *M. Tomishige (#Equal contribution) “Direct Observation of Intermediate States during the Stepping Motion of Kinesin-1”, *Nat. Chem. Biol.*, **12**, 290-297, (2016), [10.1038/nchembio.2028](https://doi.org/10.1038/nchembio.2028)
8. *Y. Matsumoto, S. Sakakihara, A. Grushnikov, K. Kikuchi, H. Noji, A. Yamaguchi, R. Iino, Y. Yagi, K. Nishino, “A Microfluidic Channel Method for Rapid Drug-susceptibility Testing of Pseudomonas Aeruginosa”, *PLOS ONE*, **11**, e0148797, (2016), [10.1371/journal.pone.0148797](https://doi.org/10.1371/journal.pone.0148797)
9. Y. Obayashi, R. Iino, *H. Noji, “A single-molecule Digital Enzyme Assay Using Alkaline Phosphatase with a Cumarin-based Fluorogenic Substrate”, *Analyst*, **140**, 5065-5073, (2015), [10.1039/c5an00714c](https://doi.org/10.1039/c5an00714c).
10. A. Yukawa, R. Iino, R. Watanabe, S. Hayashi, *H. Noji, “Key Chemical Factors of Arginine Finger Catalysis of F1-ATPase Clarified by an Unnatural Amino Acid Mutation”, *Biochemistry*, **54**, 472-480, (2015), [10.1021/bi501138b](https://doi.org/10.1021/bi501138b)
11. *R. Iino, H. Ueno, Y. Minagawa, K. Suzuki, *T. Murata, “Rotational Mechanism of Enterococcus Hirae V1-ATPase by Crystal-structure and Single-molecule Analyses”, *Curr. Opin. Struct. Biol.*, **31**, 49-56, (2015), [10.1016/j.sbi.2015.02.013](https://doi.org/10.1016/j.sbi.2015.02.013)
12. S. Enoki, R. Iino, Y. Niitani, Y. Minagawa, M. Tomishige, *H. Noji, “High-speed Angle-resolved Imaging of Single Gold Nanorod with Microsecond Temporal Resolution and One-degree Angle Precision”, *Anal. Chem.* **87**, 2079-2086, (2015), [10.1021/ac502408c](https://doi.org/10.1021/ac502408c)
13. Y. Shibafuji, A. Nakamura, T. Uchihashi, N. Sugimoto, S. Fukuda, H. Watanabe, M. Samejima, T. Ando, H. Noji, A. Koivula, K. Igarashi, *R. Iino, “Single-molecule Imaging Analysis of Elementary Reaction Steps of *Trichoderma reesei* Cellobiohydrolase I (Cel7A) Hydrolyzing Crystalline Cellulose Iα and III”, *J. Biol. Chem.*, **289**, 14056-14065, (2014), [10.1074/jbc.M113.546085](https://doi.org/10.1074/jbc.M113.546085)
14. T. Ikeda, R. Iino, *H. Noji, “Real-time Fluorescence Visualization of Slow Tautomerization of Single Free-base Phthalocyanines under Ambient Conditions”, *Chem. Commun.*, **50**, 9443-9446, (2014), [10.1039/C4CC02574A](https://doi.org/10.1039/C4CC02574A)
15. T. Ikeda, T. Tsukahara, R. Iino, M. Takeuchi, *H. Noji, “Motion Capture and Manipulation of Single Synthetic Molecular Rotors by Optical Microscopy”, *Angew. Chem. Int. Ed.*, **53**, 10082–10085, (2014), [10.1002/anie.201403091](https://doi.org/10.1002/anie.201403091)
16. *R. Iino, Y. Minagawa, H. Ueno, M. Hara, K. Murata, “Molecular Structure and Rotary Dynamics of Enterococcus hirae V1-ATPase” *IUBMB Life*, **66**(9), 624-630, (2014), [10.1002/iub.1311](https://doi.org/10.1002/iub.1311)
17. H. Ueno, Y. Minagawa, M. Hara, S. Rahman, I. Yamato, E. Muneyuki, H. Noji, *T. Murata, *R. Iino, “Torque Generation of Enterococcus hirae V-ATPase” *J. Biol. Chem.*, **289**, 31212-31223, (2014), [10.1074/jbc.M114.598177](https://doi.org/10.1074/jbc.M114.598177)

【井上将彦】

1. K. Hayashi, Y. Miyaoka, Y. Ohishi, T. Uchida, M. Iwamura, K. Nozaki, *M. Inouye, “The Observation of Circularly Polarized Luminescence of the Excimer from Two Perylene Cores in the form of [4]Rotaxane”, *Chem. Eur. J.*, in press, (2018), [10.1002/chem.201803215](https://doi.org/10.1002/chem.201803215)
2. F. Kurosaki, *J. Chiba, *M. Inouye, “Design and Synthesis of A DNA-like Structure Composed of Alkynyl C-nucleotide with 2-aminopyrimidin-4-one as a Nucleobase”, *Heterocycles*, in press, (2018), [10.3987/COM-18-S\(T\)53](https://doi.org/10.3987/COM-18-S(T)53)

3. Y. Kunishita, K. Nogami, R. Jomura, S. Akanuma, H. Abe, M. Inouye, *Y. Kubo, K. Hosoya
“Investigation of Receptor-mediated Cyanovobalamin (vitamin B12) Transport Across the Inner Blood-Retinal Barrier Using Fluorescence-labeled Cyanocobalamin”, *Mol. Pharm.*, **15**, 3538-3594, (2018)
10.1021/acs.molpharmaceut.8b00617
4. T. Hayashi, Y. Ohishi, S.H. Soo, S. Matsumoto, H. Abe, *M. Inouye, “Spontaneous Helic Formation of “meta”-Ethynylphenol Oligomers by Sequential Intramolecular Hydrogen-Bonding inside the Cavities”, *J. Org. Chem.*, **83**, 8742-8730, (2018), 10.1021/acs.joc.8b00996
5. Y. Ohishi, N. Yamamoto, H. Abe, *M. Inouye, “Nonplanar Macrocycle Consisting of Four Pyridine and Phenol Units Connected with Acetylene Bonds Displaying Preferential Binding to Maltoside over Monosaccharides”, *J. Org. Chem.*, **83**, 5766-5770, (2018), 10.1021/acs.joc.8b00062
6. *H. Abe, D. Hashikawa, T. Minami, K. Ohtani, K. Masuda, S. Matsumoto, M. Inouye, “Hexaphenolic Rigid Cages Prepared by Self-organization of C3v Tridentates”, *J. Org. Chem.*, **83**, 3132-3141, (2018), 10.1021/acs.joc.7b03111
7. Y. Oda, *J. Chiba, *M. Inouye, ‘Synthesis of Alkynyl C-Nucleotide Triphosphates Toward Enzymatic Elongation of Artificial DNA,” *Heterocycles*, in press, (2018), [10.3987/COM-18-S\(T\)48](#)
8. *H. Abe, C. Sato, Y. Ohishi, *M. Inouye, ‘Metathesis-Based Stapling of a Pyridine-Acetylene-Phenol Oligomers Having Alkenyl Side Chains after Intermolecular Templatation by Native Saccharides ,” *Eur. J. Org. Chem.*, **2018**(24), 3131-3138, (2018), 10.1002/ejoc.201800531
9. I. Sakaguchi, T. Fukasawa, K. Fujimoto, *M. Inouye, ‘Immobilization of Crosslinked Peptides that Possess High helical Contents and their Binding to Target DNAs on Au Surfaces,” *Chem. Lett.*, **83**, 3132, (2018), 10.1246/cl.171153
10. A.Yoshizawa, *M. Inouye, ‘A Bis(phenylethynyl)pyrene-Based [3]Rotaxane as an Extremely Photostable Fluorescence Probe Suitable for Hard-edged Irradiation Experiments,” , *ChemPhotoChem.*, **2**, 353, (2018), 10.1002/cptc.201700223
11. Y. Ohishi, H. Abe, *M. Inouye, ‘Saccharide Recognition and Helix Formation in Water with an Amphiphilic Pyridine-Phenol Alternating Oligomer” *Eur. J. Org. Chem.*, **6975**, (2017), 10.1002/ejoc.201701522
12. *H. Abe, S. Takashima, *M. Inouye, ‘Kinetic Switching of Achirality/Chirality memorization of *meta*-Ethynylpyridine polymer by Coordination of Cu(II) Outside the Polymer” *Heterocycles*, **95**, 730, (2017), 10.1002/ejoc.201700807
13. S. Hirokane, S. Takashima, *H. Abe, *M. Inouye, ‘Helix-Rotaxane hybrid Systems: Rotaxane-Stabilized, Saccharide-induced Chiral Ethynylpyridine Helices by Thermodynamic Process”, *Eur. J. Org. Chem.*, **762** (2017), 10.1002/ejoc.201601323
14. K. Hayashi, *M. Inouye, ‘Reliable and Reproducible Separation of 3,9- and 3,10-Dibromoperylenes and the Photophysical Properties of their Alkynyl Derivatives”, *Eur. J. Org. Chem.*, **2017**, 4334-4337, (2017), [10.1002/ejoc.201700807](#)
15. D. Suzuki, *H. Abe, T. Minami, S. Matsumoto, *M. Inouye, ‘Preparation and Higher-Order Structures of 2,6-Pyridylene and 2,6-Pyrazylene Alternating Macrocycle with the Inner Nitrogen Atoms in All the Aromatic Rings”, *Chem. Lett.*, **46**, 1740-1742, (2017), [10.1246/cl.170815](#)
16. K. Nogami, H. Tokumaru, G. Isokawa, T. Oyoshi, K. Fujimoto, *M. Inouye, ‘Bcl-XL-Binding Helical Peptides Possessing D-Ala Residues at their C-Termini with the Advantage of Long-Lasting Intracellular Stabilities”, *Chem. Commun.*, **53**, 12104 (2017), [10.1039/c7cc06904a](#)
17. Y. Ohishi, *H. Abe, *M. Inouye, ‘Saccharide Recognition and Helix Formation in Water with an Amphiphilic Pyridine-Phenol Alternating Oligomer”, *Eur. J. Org. Chem.*, in press, (2017), [10.1002/ejoc.201701522](#)
18. *H. Abe, S. Takashima, *M. Inouye, ‘Kinetic Switching of Achirality/Chirality Memorization of *meta*-Ethynylpyridine Polymer by Coordination of Cu(II) Outside the Polymer”, *Heterocycles*, **95**, 730-737, (2017), [10.3987/COM-16-S\(S\)64](#)
19. S. Hirokane, S. Takashima, *H. Abe, *M. Inouye, ‘Helix-Rotaxane Hybrid Systems: Rotaxane-Stabilized, Saccharide-Induced Chiral Ethynylpyridine Helices by a Thermodynamic Process”, *Eur. J. Org. Chem.*, **2017**, 726-733, (2017), [10.1002/ejoc.201601323](#)
20. *H. Abe, R. Yumoto, *M. Inouye, ‘A D_{3h} -Symmetric Macrocycle Alternatingly Composed of Pyridine and

- Benzyl Alcohol Units Linked with Acetylene Bonds”, *Heterocycles*, **93**, 580–592, (2016),
21. *H. Abe, T. Yoneda, Y. Ohishi, *M. Inouye, “ D_{3h} -Symmetrical Shape-Persistent Macrocycles Consisting of Pyridine-Acetylene-Phenol Conjugates as a Highly Efficient Host Architecture for Saccharide Recognition”, *Chem.-Eur. J.*, **22**, 18944–18952, (2016), [10.1002/chem.201603987](https://doi.org/10.1002/chem.201603987)
22. *M. Inouye, A. Yoshizawa, M. Shibata, Y. Yonenaga, K. Fujimoto, T. Sakata, S. Matsumoto, M. Shiro, “Cyclodextrin-Isolated Alkynylpyrenes as UV-Stable and Blue-Light-Emitting Molecules Even in Condensed States”, *Org. Lett.*, **18**, 1960–1963 (2016), [10.1021/acs.orglett.6b00420](https://doi.org/10.1021/acs.orglett.6b00420)
- 【上野隆史】
1. H. Negishi, *S. Abe, K. Yamashita, K. Hirata, K. Niwase, M. Boudes, F. Coulibaly, H. Mori, *T. Ueno, “Supramolecular Protein Cages Constructed from a Crystalline Protein Matrix”, *Chem. Commun.*, **54**, 1988–1991, (2018), [10.1039/C7CC08689J](https://doi.org/10.1039/C7CC08689J)
 2. H. Tabe, H. Takahashi, T. Shimoi, S. Abe, *T. Ueno, *Y. Yamada, “Photocatalytic Hydrogen Evolution Systems Constructed in Cross-linked Porous Protein Crystals”, *Appl. Catal., B*, in press, (2018), [10.1016/j.apcatb.2018.01.046](https://doi.org/10.1016/j.apcatb.2018.01.046)
 3. S. Ryu, Y. Matsumoto, T. Matsumoto, T. Ueno, Y. R. Silberberg, *C. Nakamura, “Improved Efficiency of Nanoneedle Insertion by Modification with a Cell-puncturing Protein”, *Jpn. J. Appl. Phys.*, **57**, 03EB02 (2018), [10.7567/JJAP.57.03EB02](https://doi.org/10.7567/JJAP.57.03EB02)
 4. T. K. Nguyen and *T. Ueno, “Engineering of Protein Assemblies within Cells”, *Curr. Opin. Struct. Biol.*, **51**, 1–8, (2018), [10.1016/j.sbi.2017.12.005](https://doi.org/10.1016/j.sbi.2017.12.005)
 5. 安部聰、上野隆史, 「超分子タンパク質の分子設計によるバイオハイブリッド材料の開発」, 有機合成化学協会誌, 75 卷 (2017) 12 号 p. 1264-1273, [10.5059/yukigoseikyokaishi.75.1264](https://doi.org/10.5059/yukigoseikyokaishi.75.1264)
 6. S. Abe, B. Maity, T. Ueno, “Functionalization of Protein Crystals with Metal Ions, Complexes and Nanoparticles”, *Curr. Opin. Chem. Biol.*, **43**, 68–76, (2017), [10.1016/j.cbpa.2017.11.015](https://doi.org/10.1016/j.cbpa.2017.11.015)
 7. H. Inaba, T. Ueno, “Artificial bio-nanomachines Based on Protein Needles Derived from Bacteriophage T4”, *Biophys. Rev.*, **10**, 641–658, (2017), [10.1007/s12551-017-0336-9](https://doi.org/10.1007/s12551-017-0336-9)
 8. S. Abe, K. Atsumi, K. Yamashita, K. Hirata, H. Mori, T. Ueno, “Structure of in Cell Protein Crystals Containing Organometallic Complexes”, *Phys. Chem. Chem. Phys.*, **20**, 2986–2989, (2017), [10.1039/C7CP06651A](https://doi.org/10.1039/C7CP06651A)
 9. B. Maity, S. Abe, and T. Ueno, “Observation of Gold Sub-nanocluster Nucleation Within a Crystalline Protein Cage”, *Nat. Commun.*, **8**, 14820, (2017), [10.1038/ncomms14820](https://doi.org/10.1038/ncomms14820)
 10. S. Abe, H. Tabe, H. Ijiri, K. Yamashita, K. Hirata, K. Atsumi, T. Shimoi, M. Akai, H. Mori, S. Kitagawa, *T. Ueno, “Crystal Engineering of Self-Assembled Porous Protein Materials in Living Cells”, *ACS Nano*, **11**, 2410–2419, (2017), [10.1021/acsnano.6b06099](https://doi.org/10.1021/acsnano.6b06099)
 11. B. Maity, T. Ueno, “Design of Bioinorganic Materials At the Interface of Coordination and Biosupramolecular Chemistry”, *Chem. Rec.*, **17**, 383–398, (2017), [10.1002/tcr.201600122](https://doi.org/10.1002/tcr.201600122)
 12. B. Maity, K. Fukumori, S. Abe, T. Ueno, “Immobilization of Two Organometallic Complexes into a Single Cage to Construct Protein-based Microcompartments”, *Chem. Commun.*, **52**, 5463–5466, (2016), [10.1039/C6CC00679E](https://doi.org/10.1039/C6CC00679E)
 13. H. Tabe, T. Shimoi, M. Boudes, S. Abe, F. Coulibaly, S. Kitagawa, H. Mori, *T. Ueno, “Photoactivatable CO Release from Engineered Protein Crystals to Modulate NF-κB Activation”, *Chem. Commun.*, **52**, 4545–4548, (2016), [10.1039/C5CC10440H](https://doi.org/10.1039/C5CC10440H)
 14. K. Fujita, Y. Tanaka, S. Abe, *T. Ueno, “A Photoactive CO Releasing Protein Cage for Dose-Regulated Delivery in Living Cells”, *Angew. Chem. Int. Ed.*, **55**, 1056–1060, (2016), [10.1002/anie.201506738](https://doi.org/10.1002/anie.201506738)
 15. B. Maity, K. Fujita, *T. Ueno, “Use of the Confined Spaces of Apo-Ferritin and Virus Capsids as Nanoreactors for Catalytic Reactions”, *Curr. Opin. Chem. Biol.*, **25**, 88–97, (2015), [10.1016/j.cbpa.2014.12.026](https://doi.org/10.1016/j.cbpa.2014.12.026)
 16. S. Abe, *T. Ueno, “Design of Protein Crystals in the Development of Solid Biomaterials”, *RSC Advances*, **5**, 21366–21375, (2015), [10.1039/C4RA16748A](https://doi.org/10.1039/C4RA16748A)
 17. S. Abe, H. Ijiri, H. Negishi, H. Yamanaka, K. Sasaki, K. Hirata, H. Mori, *T. Ueno, “Design of Enzyme-Encapsulated Protein Containers by in Vivo Crystal Engineering”, *Adv. Mater.*, **27**, 7951–7956, (2015), [10.1002/adma.201503827](https://doi.org/10.1002/adma.201503827).
 18. *H. Nakajima, M. Kondo, T. Nakane, S. Abe, T. Nakao, Y. Watanabe, *T. Ueno, “Construction of an Enterobactin Analogue with Symmetrically Arranged Monomer Subunits of Ferritin”, *Chem. Commun.*, **51**, 16609–16612, (2015), [10.1039/C5CC06904A](https://doi.org/10.1039/C5CC06904A).

19. H. Inaba, N.J.M. Sanghamitra, K. Fujita, T. Sho, T. Kuchimaru, S. Kitagawa, S. Kizaka-Kondohc, *T. Ueno, "A Metal Carbonyl-protein Needle Composite Designed for Intracellular CO Delivery to Modulate NF-κB activity", *Mol. BioSyst.*, **11**, 3111-3118, (2015), [10.1039/C5MB00327J](https://doi.org/10.1039/C5MB00327J)
20. H. Inaba, K. Fujita, *T. Ueno, "Design of Biomaterials for intracellular delivery of carbon monoxide", *Biomaterials Science*, **3**, 1423-1438, (2015), [10.1039/C5BM00210A](https://doi.org/10.1039/C5BM00210A)
21. H. Inaba, S. Kitagawa, *T. Ueno, "Protein Needles as Molecular Templates for Artificial Metalloenzymes" *Isr. J. Chem.*, **55**, 40-50, (2015), [10.1002/ijch.201400097](https://doi.org/10.1002/ijch.201400097)
22. H. Tabe, T. Shimo, K. Fujita, S. Abe, H. Ijiri, M. Tsujimoto, T. Kuchimaru, S. Kizaka-Kondo, H. Mori, S. Kitagawa, *T. Ueno, "Design of a CO-releasing Extracellular Scaffold using in-vivo Protein Crystals", *Chem. Lett.*, **44**, 342-344, (2015), [10.1246/cl.141035](https://doi.org/10.1246/cl.141035)
23. H. Tabe, K. Fujita, S. Abe, M. Tsujimoto, T. Kuchimaru, S. Kizaka-Kondoh, M. Takano, S. Kitagawa, *T. Ueno, "Preparation of a Cross-linked Porous Protein Crystal containing Ru carbonyl complexes as a CO-releasing Extracellular Scaffold", *Inorg. Chem.*, **54**, 215-220 (2015), [10.1021/ic502159x](https://doi.org/10.1021/ic502159x)
24. S. Abe, Y. Tokura, R. Pal, N. Komura, A. Imamura, K. Matsumoto, H. Ijiri, N. J. M. Sanghamitra, H. Tabe, H. Ando, M. Kiso, H. Mori, S. Kitagawa, *T. Ueno, "Surface Functionalization of Protein Crystals with Carbohydrate Using Site-selective Bioconjugation", *Chem. Lett.*, **44**, 29-31 (2015), [10.1246/cl.140865](https://doi.org/10.1246/cl.140865)
25. H. Tabe, S. Abe, T. Hikage, S. Kitagawa, *T. Ueno, "Porous Protein Crystals as Catalytic Vessels for Organometallic Complexes" *Chem. Asian J.*, **9**, 1373-1378, (2014), (Selected as a Cover Picture) [10.1002/asia.201301347](https://doi.org/10.1002/asia.201301347)
26. H. Inaba, N. J. M. Sanghamitra, T. Fukai, T. Matsumoto, K. Nishijo, S. Kanamaru, F. Arisaka, S. Kitagawa, *T. Ueno, "Intracellular Protein Delivery System with Protein Needle-GFP Construct" *Chem. Lett.*, **43**, 1505-1507 (2014), [10.1246/cl.140481](https://doi.org/10.1246/cl.140481)
27. N. J. M. Sanghamitra, H. Inaba, F. Arisaka, D. -O. Wang, S. Kanamaru, S. Kitagawa, *T. Ueno, "Plasma Membrane Translocation of a Protein Needle Based on a Triple-stranded β-helix Motif" *Mol. BioSyst.*, **10**, 2677-2683 (2014), [10.1039/C4MB00293H](https://doi.org/10.1039/C4MB00293H)
28. K. Fujita, Y. Tanaka, T. Sho, S. Ozeki, S. Abe, T. Hikage, T. Kuchimaru, S. Kizaka-Kondoh, *T. Ueno, "Intracellular CO Release from Composite of Ferritin and RutheniumCarbonyl Complexes", *J. Am. Chem. Soc.*, **136**(48), 16902-16908, (2014), [10.1021/ja508938f](https://doi.org/10.1021/ja508938f)

【大谷亮】

1. *R. Ohtani, T. Tokita, T. Takaya, *K. Iwata, M. Kinoshita, N. Matsumori, M. Nakamura, L. F. Lindoy, *S. Hayami, "The Impact of Metal Complex Lipids on Viscosity and Curvature of Hybrid Liposomes", *Chem. Commun.*, **53**, 13249-13252, (2017), [10.1039/C7CC07944C](https://doi.org/10.1039/C7CC07944C)
2. Y. Sekimoto, R. Ohtani, M. Nakamura, L. F. Lindoy, *S. Hayami, "Tunable Pressure Effects in Graphene Oxide Layers", *Sci. Rep.*, **7**, 12159, (2017), [10.1038/s41598-017-12444-x](https://doi.org/10.1038/s41598-017-12444-x)
3. F. Kobayashi, R. Ohtani, S. Teraoka, W. Kosaka, H. Miyasaka, Y. Zhang, L. F. Lindoy, *S. Hayami, *M. Nakamura, "Syntheses, Structures and Magnetic Properties of Tetranuclear Cubane-type and Heptanuclear Wheel-type Nickel(II) Complexes with 3-Methoxysalicylic Acid Derivatives", *Dalton Trans.*, **46**, 8555-8561, (2017), [10.1039/C7DT01757J](https://doi.org/10.1039/C7DT01757J)
4. *R. Ohtani, A. Grosjean, R. Ishikawa, R. Yamamoto, M. Nakamura, J. K. Clegg, *S. Hayami, "Zero in-plane Thermal Expansion in Guest-tunable 2D Coordination Polymers", *Inorg. Chem.*, **56**, 6225-6233, (2017), [10.1021/acs.inorgchem.7b00282](https://doi.org/10.1021/acs.inorgchem.7b00282)
5. R. Akiyoshi, K. Kuroiwa, S. A. Amolegbé, M. Nakaya, R. Ohtani, M. Nakamura, L. F. Lindoy, *S. Hayami, "Supramolecular Architectures Self-assembled Using Long Chain Alkylated Spin Crossover Cobalt(II) Compounds", *Chem. Commun.*, **53**, 4685-4687, (2017), [10.1039/C7CC01501A](https://doi.org/10.1039/C7CC01501A)
6. M. Nakaya, R. Ohtani, K. Sugimoto, M. Nakamura, L. F. Lindoy, *S. Hayami, "Molecular Assemblies of Metal Complexes via Base Pairing of Nucleic Acids in the Crystalline State", *Chem. Eur. J.*, **23**, 7232-7237, (2017), [10.1002/chem.201700593](https://doi.org/10.1002/chem.201700593)
7. *R. Ohtani, Y. Kitamura, Y. Hijikata, M. Nakamura, L. F. Lindoy, *S. Hayami, "Modulation of Redox Potentials Utilizing the Flexible Coordination Sphere of a Penta-coordinate Complex in the Solid States", *Dalton Trans.*, **46**, 3749-3754, (2017), [10.1039/c7dt00233e](https://doi.org/10.1039/c7dt00233e)
8. *R. Ohtani, *S. Hayami, "Guest-dependent Spin-transition Behavior of Porous Coordination Polymers", *Chem. Eur. J.*, **23**, 2236-2248, (2017), [10.1002/chem.201601880](https://doi.org/10.1002/chem.201601880)
9. Y. Sekimoto, M. R. Karim, N. Saigo, R. Ohtani, M. Nakamura, *S. Hayami, "Crystal Structures and

Spin-Crossover Behavior of Iron(II) Complexes with Chiral and Racemic Ligands”, *Eur. J. Inorg. Chem.*, **2017**, 1049-1053, (2017), [10.1002/ejic.201601232](https://doi.org/10.1002/ejic.201601232)

10. K. Wakata, M. R. Karim, M. S. Islam, R. Ohtani, M. Nakamura, M. Koinuma, *S. Hayami, “Superionic Conductivity in Hybrid of 3-hydroxypropanesulfonic Acid and Graphene Oxide”, *Chem. Asian J.*, **12**, 194-197, (2017), [10.1002/asia.201601488](https://doi.org/10.1002/asia.201601488)
11. H. Ohmagari, R. Ohtani, M. Nakaya, M. Ohba, M. Nakamura, L. F. Lindoy, *S. Hayami, “Water-Dependent Charge-transfer-induced Spin Transition of a Prussian Blue Analog”, *Dalton Trans.*, **45**, 16784-16788, (2016), [10.1039/C6DT03474H](https://doi.org/10.1039/C6DT03474H)

【片山勉】

1. Y. Sakiyama, M. Nishimura, C. Hayashi, Y. Akama, S. Ozaki, *T. Katayama, “The DnaA AAA+ domain His136 Residue Directs DnaB Replicative Helicase to the Unwound Region of the Replication Origin, oriC”, *Front. Microbiol.*, in press, (2018), [10.3389/fmicb.2018.02017](https://doi.org/10.3389/fmicb.2018.02017)
2. Y. Sakiyama, K. Kasho, Y. Noguchi, H. Kawakami, *T. Katayama, “Regulatory Dynamics in the Ternary DnaA Complex for Initiation of Chromosomal Replication in *Escherichia coli*”, *Nucleic Acids Res.*, **45**, 12354-12373, (2017), [10.1093/nar/gkx914](https://doi.org/10.1093/nar/gkx914)
3. *T. Katayama, K. Kasho, H. Kawakami, “The DnaA cycle in *Escherichia coli*: Activation, Function and Inactivation of the Initiator Protein”, *Front. Microbiol.*, **8**, 2496, (2017), [10.3389/fmicb.2017.02496](https://doi.org/10.3389/fmicb.2017.02496)
4. K. Kasho, H. Tanaka, R. Sakai, *T. Katayama, “Cooperative DnaA Binding to the Negatively Supercoiled datA Locus Stimulates DnaA-ATP Hydrolysis”, *J. Biol. Chem.*, **292**, 1251-1266, (2017), [10.1074/jbc.M116.762815](https://doi.org/10.1074/jbc.M116.762815)
5. M. Shimizu, Y. Noguchi, Y. Sakiyama, H. Kawakami, *T. Katayama, *S. Takada, “Near-atomic Structural model for Bacterial DNA Replication Initiation Complex and its Functional Insights”, *Proc. Natl. Acad. Sci. USA*, **113**, E8021-E8030, (2016), [10.1073/pnas.1609649113](https://doi.org/10.1073/pnas.1609649113)
6. Y. Inoue, H. Tanaka, K. Kasho, K. Fujimitsu, T. Oshima, *T. Katayama, “Chromosomal Location of the DnaA-reactivating Sequence DARS2 is Important to Regulate Timely Initiation of DNA Replication in *Escherichia Coli*”, *Genes Cells.*, **21**, 1015-1023, (2016), [10.1111/gtc.12395](https://doi.org/10.1111/gtc.12395)

【神谷由紀子】

1. *H. Kashida, Y. Hattori, K. Tazoe, T. Inoue, K. Nishikawa, K. Ishii, S. Uchiyama, H. Yamashita, M. Abe, Y. Kamiya, *H. Asanuma, “Bifacial Nucleobases for Hexaplex Formation in Aqueous Solution,” *J. Am. Chem. Soc.*, **140**, 27, 8456-8462, (2018), [10.1021/jacs.8b02807](https://doi.org/10.1021/jacs.8b02807)
2. *Y. Kamiya, Y. Arimura, H. Ooi, K. Kato, X.G. Liang, *H. Asanuma, “Development of visible light-responsive RNA scissors based on the 10-23 DNzyme”, *ChemBioChem*, **19**, 1305-1311, (2018), [10.1002/cbic.201800020](https://doi.org/10.1002/cbic.201800020)
3. *Y. Kamiya, Y. Yamada, T. Muro, K. Matsuura, *H. Asanuma, “DNA Microcapsule for Photo-triggered Drug Release System”, *ChemMedChem*, **12**, 2016-2021, (2017), [10.1002/cmdc.201700512](https://doi.org/10.1002/cmdc.201700512)
4. *Y. Kamiya, Y. Donoshita, H. Kamimoto, K. Murayama, J. Ariyoshi, *H. Asanuma, “Introduction of 2,6-diaminopurines into Serinol Nucleic Acid (SNA) Improves Anti-miRNA Performance”, *ChemBioChem*, **18**, 1917-1922, (2017), [10.1002/cbic.201700272](https://doi.org/10.1002/cbic.201700272)
5. Y. Nakasone, H. Ooi, Y. Kamiya, H. Asanuma, *M. Terazima, “Dynamics of Inter-DNA Chain Interaction of Photoresponsive DNA”, *J. Am. Chem. Soc.*, **138**, 9001, (2016), [10.1021/jacs.6b02525](https://doi.org/10.1021/jacs.6b02525)
6. *H. Asanuma, R. Niwa, M. Akahane, K. Murayama, H. Kashida, Y. Kamiya, “Strand-invading Linear Probe Combined with Unmodified PNA”, *Bioorg. Med. Chem.*, **24**, 4129, (2016), [10.1016/j.bmc.2016.06.055](https://doi.org/10.1016/j.bmc.2016.06.055)
7. *H. Kashida, T. Osawa, K. Morimoto, Y. Kamiya, *H. Asanuma, “Molecular Design of Cy3 Derivative for Highly Sensitive in-stem Molecular Beacon and its Application to the Wash-free FISH”, *Bioorg. Med. Chem.*, **23**, 1758-1762, (2015), [10.1016/j.bmc.2015.02.030](https://doi.org/10.1016/j.bmc.2015.02.030)
8. K. Murayama, Y. Kamiya, *H. Kashida, *H. Asanuma, “Ultra-Sensitive Molecular Beacon Designed with Totally Serinol Nucleic Acid (SNA) for Monitoring mRNA in Cell”, *ChemBioChem*, **16**, 1298-1301, (2015), [10.1002/cbic.201500167](https://doi.org/10.1002/cbic.201500167)
9. Y. Kamiya, K. Iishiba, T. Doi, K. Tsuda, H. Kashida, *H. Asanuma, “Terminus-free siRNA Prepared by Photo-crosslinking Activated via Slicing by Ago2”, *Biomater. Sci.*, **3**, 1534-1538, (2015), [10.1039/C5BM00231A](https://doi.org/10.1039/C5BM00231A)
10. Y. Kanematsu, Y. Kamiya, K. Matsuo, K. Gekko, *K. Kato, *M. Tachikawa, “Isotope Effect on the Circular Dichroism Spectrum of Methyl α-D-glucopyranoside in Aqueous

Solution”, *Sci. Rep.*, **5**, 17900, (2015), 10.1038/srep17900

11. S. Ninagawa, T. Okada, Y. Sumitomo, S. Horimoto, T. Sugimoto, T. Ishikawa, S. Takeda, T Yamamoto, T. Suzuki, Y. Kamiya, *K. Kato, *K. Mori, “Forcible Destruction of Severely Misfolded Mammalian Glycoproteins by the Non-glycoprotein ERAD Pathway”, *J. Cell Biol.*, **211**, 775-784, (2015), 10.1083/jcb.201504109
12. K. Inagaki, T. Satoh, M. Yagi-Utsumi, A.C. Gulluche, T. Anzai, Y. Uekusa, Y. Kamiya, *K. Kato, “Redox-coupled Structural Changes of the Catalytic a' domain of Protein Disulfide Isomerase”, *FEBS Lett.*, **589**, 2690-2694, (2015), [10.1016/j.febslet.2015.07.041](https://doi.org/10.1016/j.febslet.2015.07.041)
13. Y. Kamiya, T. Takagi, H. Ooi, H. Ito, X.G. Liang, *H. Asanuma, “Synthetic Gene Involving Azobenzene-tethered T7 Promoter for the Photocontrol of Gene Expression by Visible Light”, *ACS Synth. Biol.*, **4** (4), 365–370, (2015), [10.1021/sb5001092](https://doi.org/10.1021/sb5001092)
14. *H. Asanuma, M. Akahane, R. Niwa, H. Kashida, Y. Kamiya, “Highly Sensitive and Robust Linear Probe for Detection of mRNA in Cells” *Angew. Chem. Int. Ed.*, **15**, 54, 4315-4319, (2015), 10.1002/anie.201411000
15. Y. Kamiya, J. Takai, H. Ito, K. Murayama, H. Kashida, *H. Asanuma, “Enhancement of Stability and Activity of siRNA by Terminal Substitution with Serinol Nucleic Acid (SNA)”, *ChemBioChem*, **15**(17), 2549-2555, (2014), [10.1002/cbic.201402369](https://doi.org/10.1002/cbic.201402369)
16. S. Ninagawa, T. Okada, Y. Sumitomo, Y. Kamiya, K. Kato, S. Horimoto, T. Ishikawa, S. Takeda, T. Sakuma, T. Yamamoto, *K. Mori, “EDEM2 Initiates Mammalian Glycoprotein ERAD by Catalyzing the First Mannose Trimming Step”, *J. Cell Biol.*, **206**(3), 347-356, (2014), [10.1083/jcb.201404075](https://doi.org/10.1083/jcb.201404075)
17. *H. Asanuma, H. Kashida, Y. Kamiya, “De Novo Design of Functional Oligonucleotides with Acyclic Scaffolds” *Chem. Rec.*, **14**(6), 1055-1069, (2014), [10.1002/tcr.201402040](https://doi.org/10.1002/tcr.201402040)
18. Y. Kamiya, *H. Asanuma, “Light-driven DNA Nanomachine with a Photoresponsive Molecular Engine” *Acc. Chem. Res.*, **47**(6), 1663-1672, (2014) , [10.1021/ar400308f](https://doi.org/10.1021/ar400308f)

【佐田和己】

1. *K. Sada, “Functional Polymers in Nonpolar Solvents induced by Dissociation of Macromolecular Complexes”, *Polym. J.*, **50**, 285-299, (2018), 10.1038/s41428-017-0016-4
2. M. Naya, Y. Hamano, K. Kokado, *K. Sada, “Organic Reaction as a Stimulus for Polymer Phase Separation.” *ACS Macro Lett.*, **6**, 898-902, (2017), 10.1021/acsmacrolett.7b00315
3. *K. Hirai, S. Isobe, *K. Sada, “Gas-generated Thermal Oxidation of a Coordination Cluster for an Anion-doped Mesoporous Metal Oxide”, *Sci. Rep.*, **5**, 18468, (2015), 10.1038/srep18468
4. D.H. Gharib, S. Amemori, M. Naya, *K. Kokado *K. Sada, “Gel Thermoresponsiveness Driven by Switching of the Charge-transfer Interaction”, *RSC Advances*, **5**, 89319, (2015), 10.1039/C5RA18388J
5. *K. Kokado, R.Taniguchi, *K. Sada, “Rigidity-induced Emission Enhancement of Network Polymers Crosslinked by Tetraphenylethene Derivatives”, *J. Mater. Chem. C*, **3**, 8504, (2015), 10.1039/C5TC01597A
6. K.Nishi, S. Tochioka, T. Hiroi, T. Yamada, K. Kokado, T.-H.Kim, E.P. Gilbert, K. Sada, *M.Shibayama, “Structural Analysis of Lipophilic Polyelectrolyte Solutions and Gels in Low-Polar Solvents”, *Macromolecules* **48**, 3613, (2015), 10.1021/acs.macromol.5b00753
7. S. Nagata, *K. Kokado, *K. Sada, “Metal-organic Framework Tethering PNIPAM for ON-OFF Controlled Release in Solution”, *Chem. Commun.*, **51**, 8614, (2015), 10.1039/C5CC02339D
8. R. Taniguchi, T. Yamada, K. Sada, *K. Kokado, “Stimuli-responsive Fluorescence of AIE Elastomer Based on PDMS and Tetraphenylethene”, *Macromolecules*, **47**(18), 6382-6388, (2014), 10.1021/ma501198d
9. T. Yamada, K. Kokado, Y. Higaki, A. Takahara, *K. Sada, “Preparation and Morphology Variation of Lipophilic Polyelectrolyte Brush Functioning in Non-Polar Solvents”, *Chem. Lett.*, **43**(8), 1300-1302, (2014), 10.1246/cl.140341
10. S. Sudo, S. Nagata, K. Kokado, *K. Sada, “Direct Synthesis of Liquid Metal Colloids and Their Transmetalation into Noble Metal Nanoparticles”, *Chem. Lett.*, **43**(8), 1207-1209, (2014), 10.1246/cl.140359
11. A. Md. R. Kabir, D. Inoue, Y. Hamano, H. Mayama, K. Sada, *A. Kakugo, “Biomolecular Motor Modulates Mechanical Property of Microtubule”, *Biomacromol.*, **15**(5), 1797-1805, (2014), 10.1021/bm5001789
12. M. Ito, A. Md. R. Kabir, D. Inoue, T. Torisawa, Y. Toyoshima, K. Sada, *A. Kakugo, “Formation of Ring-shaped Microtubule Assemblies Through Active Self-organization on Dynein”, *Polym. J.*, **46**(4), 220-225,

【佐藤宗太】

1. Y. Tachi, S. Sato, M. Yoneya, M. Fujita, Y. Kamoto, "Two Polyhedral Frameworks of an $M_{12}L_{24}$ Spherical Complex Revealed by Replica-Exchange Molecular Dynamics Simulations", in press, (2018)
2. T. Matsuno, M. Fujita, K. Fukunaga, S. Sato, *H. Isobe, "Concyclic CH- π Arrays for Single-Axis Rotations of a Bowl in a Tube" *Nature Commun.*, **9**, 3779, (2018), 10.1038/s41467-018-06270-6
3. K. Ikemoto, J. Lin, R. Kobayashi, S. Sato, *H. Isobe, "Fluctuating Carbonaceous Networks with a Persistent Molecular Shape: A Saddle-Shaped Geodesic Framework of 1,3,5-Trisubstituted Benzene (Phenine)" *Angew. Chem. Int. Ed.*, **57**, 8555-8559, (2018), [10.1002/anie.201803984](https://doi.org/10.1002/anie.201803984)
4. T. Matsuno, Y. Nakai, S. Sato, Y. Maniwa, *H. Isobe, "Ratchet-Free Solid-State Inertial Rotation of a Guest Ball in a Tight Tubular Host" *Nature Commun.*, **9**, 1907, (2018), [10.1038/s41467-018-04325-2](https://doi.org/10.1038/s41467-018-04325-2)
5. *Y. Nagata, T. Nishikawa, M. *Suginome, S. Sato, *M. Sugiyama, L. Porcar, A. Martel, R. Inoue, N. Sato "Elucidating the Solvent Effect on the Switch of the Helicity of Poly(quinoxaline-2,3-diyl)s: A Conformational Analysis by Small-Angle Neutron Scattering", *J. Am. Chem. Soc.*, **140**, 2722-2726, (2018), [10.1021/jacs.7b11626](https://doi.org/10.1021/jacs.7b11626)
6. S.-T. Pham, K. Ikemoto, K. Z. Suzuki, T. Izumi, H. Taka, H. Kita, S. Sato, H. Isobe, *S. Mizukami "Magneto-Electroluminescence Effects in the Single-Layer Organic Light-Emitting Devices with Macrocyclic Aromatic HydrocarbonsQ", *APL Materials*, **6**, 026103, (2018), [10.1063/1.5021711](https://doi.org/10.1063/1.5021711)
7. 佐藤宗太, 磯部寛之, "大環状芳香族分子のナノ細孔結晶を活用した全固体リチウムイオン電池の高容量電極材料" *応用物理(研究紹介)*, 2017, vol. 86, No. 12, p. 1065-1068
8. N. Miyamoto, Y. Nakazawa, T. Nakamura, K. Okano, S. Sato, Z. Sun, H. Isobe, *H. Tokuyama "Synthesis of 9,10-Diarylanthracenes via Mg(TMP)₂-2LiCl-Mediated Benzyne Generation/[4+2] Cycloaddition and Deoxygenation of 9,10-Epoxyanthracene Intermediates", *Synlett*, **29**, 513-518, (2017), [10.1055/s-0036-1591510](https://doi.org/10.1055/s-0036-1591510)
9. T. Matsuno, K. Kogashi, S. Sato, *H. Isobe "Enhanced yet Inverted Effects of π -Extension in Self-Assembly of Curved π -Systems with HelicityA", *Org. Lett.*, **19**, 6456-6459 (2017), [10.1021/acs.orglett.7b03534](https://doi.org/10.1021/acs.orglett.7b03534)
10. K. Kurihara, M. Matsuo, *T. Yamaguchi, *S. Sato, "Synthetic Approach to Biomolecular Science by Cyborg Supramolecular Chemistry", *Biochim. Biophys. Acta, (BBA) General Subjects.*, **1862**, 358-364, (2017), [10.1016/j.bbagen.2017.11.002](https://doi.org/10.1016/j.bbagen.2017.11.002)
11. J. Uchida, *M. Yoshio, *S. Sato, H. Yokoyama, *M. Fujita, *T. Kato, "Self-Assembly of Giant Spherical Liquid-Crystalline Complexes and Formation of Nanostructured Dynamic Gels Exhibiting Self-Healing Properties", *Angew. Chem. Int. Ed.*, **56**, 14085-14089, (2017), [10.1002/anie.201707740](https://doi.org/10.1002/anie.201707740)
12. *S. Sato, A. Yoshii, S. Takahashi, S. Furumi, M. Takeuchi, *H. Isobe, "Chiral Intertwined Spirals and Magnetic Transitiondipole Moments Dictated by Cylinder Helicity", *Proc. Natl. Acad. Sci. U.S.A.*, **114**, 13097-13101, (2017), [10.1073/pnas.1717524114](https://doi.org/10.1073/pnas.1717524114)
13. T. Matsuno, S. Kamata, S. Sato, A. Yokoyama, P. Sarkar, *H. Isobe, "Assembly, Thermodynamics, and Structure of a Two-Wheeled Composite of a Dumbbell-Shaped Molecule and Cylindrical Molecules with Different Edges", *Angew. Chem. Int. Ed.*, **56**, 15020-150247, (2017), [10.1002/anie.201709442](https://doi.org/10.1002/anie.201709442)
14. S. Hitosugi, S. Sato, T. Matsuno, T. Koretsune, R. Arita, *H. Isobe, "Pentagon-Embedded Cycloarylene Molecules with Cylindrical Shapes", *Angew. Chem. Int. Ed.*, **56**, 9106-9110, (2017), [10.1002/anie.201704676](https://doi.org/10.1002/anie.201704676)
15. Y. Tian, K. Ikemoto, *S. Sato, *H. Isobe, "[n]Cyclo-3,6-Phenanthrenylenes: Synthesis, Structure and Fluorescence", *Chem. Asian J.*, **12**, 2093-2097, (2017), [10.1002/asia.201700563](https://doi.org/10.1002/asia.201700563)
16. T. Izumi, Y. Tian, K. Ikemoto, A. Yoshii, T. Koretsune, R. Arita, H. Kita, H. Taka, *S. Sato, *H. Isobe, "Efficient Blue Electroluminescence from a Single-Layer Organic Device Composed Solely of Hydrocarbons", *Chem. Asian J.*, **12**, 730-733, (2017), [10.1002/asia.201700198](https://doi.org/10.1002/asia.201700198)
17. A. Yoshii, K. Ikemoto, T. Izumi, H. Kita, H. Taka, T. Koretsune, R. Arita, *S. Sato, *H. Isobe, "Structural Modulation of Macrocyclic Materials for Charge Carrier Transport Layers in Organic Light-Emitting Devices", *ECS J. Solid State Sci. Technol.*, **6**, M3065-M3067, (2017), [10.1149/2.0111706jss](https://doi.org/10.1149/2.0111706jss)
18. S. Kamata, S. Sato, *J. Wu, *H. Isobe, "Crystal Structure of 7,15-Bis-(4-tert-butylphenyl)-1,9-dimethylheptazethrene", *Acta Cryst.*, **E73**, 99-102, (2017), [10.1107/S2056989016020247](https://doi.org/10.1107/S2056989016020247)
19. K. Ikemoto, R. Kobayashi, S. Sato, *H. Isobe, "Entropy-Driven Ball-in-Bowl Assembly of Fullerene and

- Geodesic Phenylene Bowl”, *Org. Lett.*, **19** (9), 2362–2365, (2017), [10.1021/acs.orglett.7b00899](https://doi.org/10.1021/acs.orglett.7b00899)
20. G. Yan, T. Yamaguchi, T. Suzuki, S. Yanaka, S. Sato, M. Fujita, *K. Kato, “Hyper-assembly of Self-assembled Glycoclusters Mediated by Specific Carbohydrate-carbohydrate Interactions”, *Chem. Asian J.*, **12**(9), 968-972, (2017), [10.1002/asia.201700202](https://doi.org/10.1002/asia.201700202)
21. K. Ikemoto, R. Kobayashi, S. Sato, *H. Isobe, “Synthesis and Bowl-in-bowl Assembly of a Geodesic Phenylene Bowl”, *Angew. Chem. Int. Ed.*, **56**, 6511-6514, (2017), [10.1002/anie.201702063](https://doi.org/10.1002/anie.201702063)
(Very Important Paper(VIP)に採用)(Inside Back Cover Pictureに採用)
22. *K. Z. Suzuki, T. Izumi, X. Zhang, A. Sugihara, S.-T. Pham, H. Taka, S. Sato, H. Isobe, S. Mizukami, “Room Temperature Magnetoresistance in an Organic Spin Valve with an Aromatic Hydrocarbon macrocycle”, *APL Mater.*, **5**, 046101, (2017), [10.1063/1.4979548](https://doi.org/10.1063/1.4979548)
23. Y. Suzuki, S. Ishida, S. Sato, H. Isobe, *T. Iwamoto, “An Isolable Potassium Salt of a Borasilene–Chloride Adduct”, *Angew. Chem. Int. Ed.*, **56**, 4593-4597, (2017), [10.1002/anie.201612545](https://doi.org/10.1002/anie.201612545)
24. Z. Sun, N. Miyamoto, S. Sato, *H. Tokuyama, *H. Isobe, “An Obtuse-Angled Corner Unit for Fluctuating Carbon Nano hoops”, *Chem. Asian J.*, **12**, 271-275, (2017), [10.1002/asia.201601614](https://doi.org/10.1002/asia.201601614)
25. D. Fujita, Y. Ueda, S. Sato, H. Yokoyama, N. Mizuno, T. Kumazaka, *M. Fujita, “Self-Assembly of $M_{30}L_{60}$ Icosidodecahedron”, *Chem.*, **1**, 91-101, (2016), [10.1016/j.chempr.2016.06.007](https://doi.org/10.1016/j.chempr.2016.06.007)
26. *D. Fujita, Y. Ueda, S. Sato, N. Mizuno, T. Kumazaka, *M. Fujita, “Self-assembly of Tetraivalent Goldberg Polyhedra from 144 Small Components”, *Nature*, **540**, 563-566, (2016), [10.1038/nature20771](https://doi.org/10.1038/nature20771)
27. M. Sato, H. Azuma, A. Daigaku, S. Sato, K. Takasu, K. Okano, *H. Tokuyama, “Total Synthesis of (-)-Histrionicotoxin through a Stereoselective Radical Translocation-Cyclization Reaction”, *Angew. Chem. Int. Ed.*, **56**, 1087-1091, (2016), [10.1002/anie.201609941](https://doi.org/10.1002/anie.201609941)
28. T. Matsuno, S. Sato, A. Yokoyama, S. Kamata, *H. Isobe, “Self-Sorting of Two Hydrocarbon Receptors with One Carbonaceous Ligand”, *Angew. Chem. Int. Ed.*, **55**, 15339-15343, (2016), [10.1002/anie.201609444](https://doi.org/10.1002/anie.201609444)
29. P. Sarkar, Z. Sun, T. Tokuhira, M. Kotani, S. Sato, *H. Isobe, “Stereoisomerism in Nano hoops with Heterogeneous Biaryl Linkages of E/Z- and R/S-Geometries”, *ACS Cent. Sci.*, **2**, 740-747, (2016), [10.1021/acscentsci.6b00240](https://doi.org/10.1021/acscentsci.6b00240)
30. M. Kanto, S. Sato, N. Tsuda, *M. Sasaki, “Stereodivergent Synthesis and Configurational Assignment of the C1–C15 Segment of Amphirionin-5”, *J. Org. Chem.*, **81**, 9105-9121, (2016), [10.1021/acs.joc.6b01700](https://doi.org/10.1021/acs.joc.6b01700)
31. Z. Sun, T. Suenaga, P. Sarkar, S. Sato, M. Kotani, *H. Isobe, “Stereoisomerism, Crystal Structures, and Dynamics of Belt-shaped Cyclonaphthylenes”, *Proc. Natl. Acad. Sci. U.S.A.*, **113**, 8109-8114, (2016), [10.1073/pnas.1606530113](https://doi.org/10.1073/pnas.1606530113)
32. *S. Sato, A. Unemoto, T. Ikeda, *S. Orimo, *H. Isobe, “Carbon-rich Active Materials with Macroyclic Nanochannels for High-Capacity Negative Electrodes in All-Solid-State Lithium Rechargeable Battery”, *Small*, **12**, 3381-3387, (2016), [10.1002/smll.201600916](https://doi.org/10.1002/smll.201600916)
33. K. Ikemoto, M. Fujita, P. Too, Y. Tnay, S. Sato, S. Chiba, *H. Isobe, “Synthesis and Structures of π -Extended [n]Cyclo-para-phenylenes ($n = 12, 16, 20$) Containing $n/2$ Nitrogen Atoms”, *Chem. Lett.*, **45**, 658-660, (2016), [10.1246/cl.160258](https://doi.org/10.1246/cl.160258)
34. J. Xue, K. Ikemoto, S. Sato, *H. Isobe, “Introduction of Nitrogen Atoms in [n]Cyclo-meta-phenylenes via Cross Coupling Macrocyclization”, *Chem. Lett.*, **45**, 676-678, (2016), [10.1246/cl.160218](https://doi.org/10.1246/cl.160218)
35. *H. Isobe, K. Nakamura, S. Hitosugi, S. Sato, H. Tokoyama, H. Yamakado, K. Ohno, *H. Kono, “Reply to the ‘Comment on ‘Theoretical studies on a carbonaceous molecular bearing: association thermodynamics and dual-mode rolling dynamics’ by E. M. Cabaleiro-Lago, J. Rodriguez-Otero and A. Gil, *Chem. Sci.*, 2016, 7, DOI: 10.1039/C5SC04676A”, *Chem. Sci.*, **7**, 2929-2932 (2016), [10.1039/C6SC00550K](https://doi.org/10.1039/C6SC00550K)
36. K. Ikemoto, S. Sato, *H. Isobe, “One-pot Synthesis of [n]Cyclo-1,3-pyrenylenes via Ni-mediated Macrocyclization”, *Chem. Lett.*, **45**, 217-219, (2016), [10.1246/cl.151112](https://doi.org/10.1246/cl.151112)
37. K. Ikemoto, A. Yoshii, T. Izumi, H. Taka, H. Kita, J. Y. Xue, R. Kobayashi, *S. Sato, *H. Isobe, “Modular Synthesis of Aromatic Hydrocarbon Macrocycles for Simplified, Single-Layer Organic Light-Emitting Devices”, *J. Org. Chem.*, **81**, 662-666, (2016), [10.1021/acs.joc.5b02620](https://doi.org/10.1021/acs.joc.5b02620)
38. J. Y. Xue, T. Izumi, A. Yoshii, K. Ikemoto, T. Koretsune, R. Akashi, R. Arita, H. Taka, H. Kita, *S. Sato, *H. Isobe, “Aromatic Hydrocarbon Macrocycles for Highly Efficient Organic Light-emitting Devices with Single-layer Architectures”, *Chem. Sci.*, **7**, 896-904, (2016), [10.1039/C5SC03807C](https://doi.org/10.1039/C5SC03807C)

39. P. Sarkar, S. Sato, S. Kamata, T. Matsuno, *H. Isobe, "Synthesis and Dynamic Structures of a Hybrid Nanohoop Molecule Composed of Anthanthrenylene and Phenylene Panels", *Chem. Lett.*, **44**, 1581-1583, (2015), [10.1246/cl.150801](https://doi.org/10.1246/cl.150801)
40. Z. Sun, P. Sarkar, T. Suenaga, S. Sato, *H. Isobe, "Belt-shaped Cyclonaphthylenes", *Angew. Chem. Int. Ed.*, **54**, 12800-12804, (2015), [10.1002/anie.201506424](https://doi.org/10.1002/anie.201506424)
41. S. Hitosugi, K. Ohkubo, Y. Kawashima, T. Matsuno, S. Kamata, K. Nakamura, H. Kono, S. Sato, S. Fukuzumi, *H. Isobe, "Modulation of Energy Conversion Processes in Carbonaceous Molecular Bearings", *Chem. Asian J.*, **10**, 2404-2410, (2015), [10.1002/asia.201500673](https://doi.org/10.1002/asia.201500673)
42. *M. Kobayashi, T. Okuhara, H. Kato, S. Sato, M. Kakihana, "Novel Titanium Complexes with a Reversible Structure Change on Solvent Adsorption and Desorption", *Chem. Lett.*, **44**, 1050-1052, (2015), [10.1246/cl.150393](https://doi.org/10.1246/cl.150393)
43. P. Bonakdarzadeh, F. Topić, E. Kalenius, S. Bhowmik, S. Sato, M. Groessl, R. Knochenuuss, *K. Rissanen, "DOSY NMR, X-ray Structural and Ion-Mobility Mass Spectrometric Studies on Electron-Deficient and Electron-Rich M_6L_4 Coordination Cages", *Inorg. Chem.*, **54**, 6055-6061, (2015), [10.1021/acs.inorgchem.5b01082](https://doi.org/10.1021/acs.inorgchem.5b01082)
44. H. Yokoyama, Y. Ueda, D. Fujita, S. Sato, *M. Fujita, "Finely Resolved Threshold for the Sharp $M_{12}L_{24}/M_{24}L_{48}$ Structural Switch in Multi-Component M_nL_{2n} Polyhedral Assemblies: X-ray, MS, NMR, and Ultracentrifugation Analyses", *Chem. Asian J.*, **10**, 2292-2295, (2015), [10.1002/asia.201500519](https://doi.org/10.1002/asia.201500519)
45. *S. Sato, M. Ikemi, T. Kikuchi, S. Matsumura, *K. Shiba, *M. Fujita, "Bridging Adhesion of a Protein onto an Inorganic Surface Using Self-Assembled Dual Functionalized Spheres", *J. Am. Chem. Soc.*, **137**, 12890-12896, (2015), [10.1021/jacs.5b06184](https://doi.org/10.1021/jacs.5b06184)
46. E. Takahashi, K. Kamata, Y. Kikukawa, S. Sato, K. Suzuki, K. Yamaguchi, *N. Mizuno, "Synthesis and Oxidation Catalysis of a Ti-Substituted Phosphotungstate, and Identification of the Active Oxygen Species", *Catal. Sci. Technol.*, **5**, 4778-4789, (2015), [10.1039/C5CY01031D](https://doi.org/10.1039/C5CY01031D)
47. *S. Sato, Y. Yoshimasa, D. Fujita, M. Yagi-Utsumi, T. Yamaguchi, *K. Kato, *M. Fujita, "Self-Assembled Spherical Complex Displaying a Gangliosidic Glycan Cluster Capable of Interacting with Amyloidogenic Proteins", *Angew. Chem. Int. Ed.*, **54**, 8435-8439, (2015), [10.1002/anie.201501981](https://doi.org/10.1002/anie.201501981)
48. *H. Isobe, K. Nakamura, S. Hitosugi, S. Sato, H. Tokoyama, H. Yamakado, K. Ohno, *H. Kono, "Theoretical Studies on a Carbonaceous Molecular Bearing: Association Thermodynamics and Dual-mode Rolling Dynamics", *Chem. Sci.*, **6**, 2746-2753, (2015), [10.1039/c5sc00335k](https://doi.org/10.1039/c5sc00335k)
49. T. Matsuno, S. Sato, R. Iizuka, *H. Isobe, "Molecular Recognition in Curved π -systems: Effects of π -lengthening of Tubular Molecules on Structures and Thermodynamics", *Chem. Sci.*, **6**, 909-916, (2015), [10.1039/C4SC02812K](https://doi.org/10.1039/C4SC02812K)
50. *S. Sato, R. Takeuchi, M. Yagi-Utsumi, T. Yamaguchi, Y. Yamaguchi, K. Kato, *M. Fujita, "Self-Assembled, π -Stacked Complex as a Finely-Tunable Magnetic Aligner for Biomolecular NMR Applications", *Chem. Commun.*, **51**, 2540-2543, (2015), [10.1039/C4CC09354B](https://doi.org/10.1039/C4CC09354B)
51. D. Fujita, H. Yokoyama, Y. Ueda, S. Sato, *M. Fujita, "Geometrically Restricted Intermediates in the Self-Assembly of an $M_{12}L_{24}$ Cuboctahedral Complex", *Angew. Chem. Int. Ed.*, **54**, 155-158, (2015), [10.1002/anie.201409216](https://doi.org/10.1002/anie.201409216)
52. Q.-F. Sun, S. Sato, *M. Fujita, "An $M_{12}(L^1)_{12}(L^2)_{12}$ Cantellated Tetrahedron: A Case Study for Mixed-Ligand Self-Assembly", *Angew. Chem. Int. Ed.*, **53**, 13510-13513, (2014), [10.1002/anie.201408652](https://doi.org/10.1002/anie.201408652)
53. C. J. Bruns, D. Fujita, M. Hoshino, S. Sato, *J. F. Stoddart, *M. Fujita, "Emergent Ion-Gated Binding of Cationic Host–Guest Complexes within Cationic $M_{12}L_{24}$ Molecular Flasks", *J. Am. Chem. Soc.*, **136**, 12027-12034, (2014), [10.1021/ja505296e](https://doi.org/10.1021/ja505296e)
54. J. Y. Xue, K. Ikemoto, N. Takahashi, T. Izumi, H. Taka, H. Kita, *S. Sato, *H. Isobe, "Cyclo-meta-phenylenes Revisited: Nickel-mediated Synthesis, Mstructures and Device Applications", *J. Org. Chem.*, **79**, 9735-9739, (2014), [10.1021/jo501903n](https://doi.org/10.1021/jo501903n)
55. S. Hitosugi, K. Ohkubo, R. Iizuka, Y. Kawashima, K. Nakamura, S. Sato, H. Kono, S. Fukuzumi, *H. Isobe, "Photoinduced Electron Transfer in a Dynamic Supramolecular System with Curved π -structures", *Org. Lett.*, **16**, 3352-3355, (2014), [10.1021/ol501381x](https://doi.org/10.1021/ol501381x)
56. S. Sato, T. Yamasaki, *H. Isobe, "Solid-state Structures of Peapod Bearings Composed of Finite Single-wall Carbon Nanotube and Fullerene Molecules", *Proc. Natl. Acad. Sci. U.S.A.*, **111**, 8374-8379, (2014), [10.1073/pnas.1406518111](https://doi.org/10.1073/pnas.1406518111)
57. T. Matsuno, H. Naito, S. Hitosugi, S. Sato, M. Kotani, *H. Isobe, "Geometric Measures of Finite Carbon

Nanotube Molecules: A Proposal for Length Index and Filling Indexes”, *Pure Appl. Chem.*, **86**, 489-495,(2014), [10.1515/pac-2014-5006](https://doi.org/10.1515/pac-2014-5006)

58. T. Kikuchi, S. Sato, D. Fujita, *M. Fujita, “Stepwise DNA Condensation by a Histone-mimic Peptide-coated $M_{12}L_{24}$ Spherical Complex”, *Chem. Sci.*, **5**, 3257-3260, (2014), [10.1039/C4SC00656A](https://doi.org/10.1039/C4SC00656A)
59. *M. Yoneya, S. Tsuzuki, T. Yamaguchi, S. Sato, M. Fujita, “Coordination-Directed Self-Assembly of $M_{12}L_{24}$ Nanocage: Effects of Kinetic Trapping on the Assembly Process”, *ACS Nano*, **8**, 1290-1296, (2014), [10.1021/nn404595j](https://doi.org/10.1021/nn404595j)

【杉安和憲】

1. S. H. Jung, D. Bochicchio, *G. M. Pavan, *M. Takeuchi, *K. Sugiyasu, “A Block Supramolecular Polymer and Its Kinetically Enhanced Stability”, *J. Am. Chem. Soc.*, **140**, 10570-10577, (2018), [10.1021/jacs.8b06016](https://doi.org/10.1021/jacs.8b06016)
2. T. Fukui, S. Kawai, S. Fujinuma, Y. Matsushita, T. Yasuda, T. Sakurai, S. Seki, M. *Takeuchi, *K. Sugiyasu, “Control over Differentiation of a Metastable Supramolecular Assembly in One and Two Dimensions”, *Nature Chem.*, **9**, 493-499, (2017), [10.1038/NCHEM.2684](https://doi.org/10.1038/NCHEM.2684)
3. T. Fukui, *M. Takeuchi, *K. Sugiyasu, “Impact of a Subtle Structural Difference on the Kinetic Behavior of Metastable Supramolecular Assemblies”, *Polymer*, **128**, 311-316, (2017), [10.1016/j.polymer.2016.12.027](https://doi.org/10.1016/j.polymer.2016.12.027)
4. T. Fukui, *M. Takeuchi, *K. Sugiyasu, “Autocatalytic Time-Dependent Evolution of Metastable Two-Component Supramolecular Assemblies to Self-Sorted or Coassembled State”, *Sci. Rep.*, **7**, 2425, (2017), [10.1038/s41598-017-02524-3](https://doi.org/10.1038/s41598-017-02524-3)
5. M. Endo, T. Fukui, S. H. Jung, S. Yagai, M. Takeuchi, *K. Sugiyasu, “Photoregulated Living Supramolecular Polymerization of Established by Combining Energy Landscapes of Photoisomerization and Nucleation-elongation Processes”, *J. Am. Chem. Soc.*, **138**, 14347, (2016), [10.1021/jacs.6b08145](https://doi.org/10.1021/jacs.6b08145)
6. 福井智也, 大城宗一郎, 竹内正之, *杉安和憲, “アミロイド繊維のように成長する超分子集合体：メカニズムの解明と時間発展プログラム”, *生物物理*, **55**, 154-156, (2015), [10.2142/biophys.55.154](https://doi.org/10.2142/biophys.55.154)
7. *杉安和憲, “超分子ポリマーの長さをそろえる”, *現代化学*, 2015, 9月号, 32-36,
8. C. Pan, C. Zhao, M. Takeuchi, *K. Sugiyasu, “Conjugated Oligomers and Polymers Sheathed with Designer Side Chains”, *Chem. Asian J.*, (2015), **10**, 1820-1835, [10.1002/asia.201500452](https://doi.org/10.1002/asia.201500452)
9. S. Ogi, V. Stepanenko, K. Sugiyasu, M. Takeuchi, *F. Würthner “Mechanism of Self-Assembly Process and Seeded Supramolecular Polymerization of Perylene Bisimide Organogelator”, *J. Am. Chem. Soc.*, **137**, 3300-3307, (2015), [10.1021/ja511952c](https://doi.org/10.1021/ja511952c)
10. S. Ogi, T. Fukui, M. Jue, *M. Takeuchi, *K. Sugiyasu “Kinetic Control over Pathway Complexity In Supramolecular Polymerization through Modulating the Energy Landscape by Rational Molecular Design”, *Angew. Chem. Int. Ed.*, **53**, 14363-14367 (2014), [10.1002/anie.201407302](https://doi.org/10.1002/anie.201407302)
11. *K. Sugiyasu, S. Ogi, M. Takeuchi, , “Strapped Porphyrin-Based Polymeric Systems”, *Polymer J.*, **46**, 674-681, (2014), [10.1038/pj.2014.58](https://doi.org/10.1038/pj.2014.58)
12. C. Pan, *K. Sugiyasu, J. Aimi, A. Sato, M. Takeuchi “Picket-Fence Polythiophene and its Diblock Copolymers that Afford Microphase Separation Comprising a Stacked and an Isolated Polythiophene Ensemble” *Angew. Chem. Int. Ed.*, **53**, 8870-8875, (2014), [10.1002/anie.201402813](https://doi.org/10.1002/anie.201402813)
13. D. Sahoo, K. Sugiyasu, Y. Tian, M. Takeuchi, *I. G. Schblykin “Effect of Conjugated Backbone Protection on Intrinsic and Light-Induced Fluorescence Quenching in Polythiophene” *Chem. Mater.*, **26**, 4867-4875, (2014), [org/10.1021/cm5021959](https://doi.org/10.1021/cm5021959)
14. C. Pan, *K. Sugiyasu, *M. Takeuchi “Blending Conjugated Polymers without Phase Separation for Fluorescent Colour Tuning of Polymeric Materials through FRET” *Chem. Commun.*, **50**, 11814-11817, (2014), [10.1039/c4cc03594a](https://doi.org/10.1039/c4cc03594a)
15. *杉安和憲, 大城宗一郎, 竹内正之, “リビング超分子重合の実現” *高分子*, **63**, 851-854, (2014)

【鈴木大介】

1. S. Matsui, Y. Nishizawa, *T. Uchihashi, *D. Suzuki, “Monitoring Thermo-responsive Morphological Changes in Individual Hydrogel Microspheres”, *ACS Omega*, **3**(9), 10836–10842 , (2018), [10.1021/acsomega.8b01770](https://doi.org/10.1021/acsomega.8b01770)
2. T. Watanabe, C. Song, K. Murata, T. Kureha, *D. Suzuki, “Seeded Emulsion Polymerization of Styrene in the Presence of Water-Swollen Hydrogel Microspheres”, *Langmuir*, **34**, 8571-8580, (2018), [10.1021/acs.langmuir.8b01047](https://doi.org/10.1021/acs.langmuir.8b01047)
3. T. Kureha, Y. Nagase, *D. Suzuki, “High Reusability of Catalytically Active Gold Nanoparticles

- Immobilized in Core-Shell Hydrogel Microspheres”, *ACS Omega*, **3**, 6158-6165, (2018), 10.1021/acsomega.8b00819
4. H. Minato, M. Murai, T. Watanabe, S. Matsui, M. Takizawa, *T. Kureha, *D. Suzuki, “The Deformation of Hydrogel Microspheres at the Air/Water Interface”, *Chem. Commun.*, **54**, 932-935, (2018), 10.1039/C7CC09603H
 5. S. Minami, T. Watanabe, *D. Suzuki, *K. Urayama, “Viscoelasticity of Dense Suspensions of Thermosensitive Microgel Mixtures Undergoing Colloidal Gelation”, *Soft Matter*, **14**, 1596-1607, (2018), 10.1039/C7SM02411H
 6. M. Takizawa, Y. Sazuka, K. Horigome, Y. Sakurai, S. Matsui, H. Minato, T. Kureha, *D. Suzuki, “Self-organization of Soft Hydrogel Microspheres during the Evaporation of Aqueous Droplets”, *Langmuir*, accepted (2018), 10.1021/acs.langmuir.8b00230
 7. T. Kureha, *D. Suzuki “Nanocomposite Microgels for the Selective Separation of Halogen Compounds from Aqueous Solution”, *Langmuir*, **34**, 837-846, (2018), 10.1021/acs.langmuir.7b01485
 8. *D. Suzuki, “Control of Spatio-temporal Structures for Polymeric Hydrogel Microspheres”, 高分子, **66**, 6, (2017)
 9. *D. Suzuki, K. Horigome, T. Kureha, S. Matsui, T. Watanabe “Polymeric Hydrogel Microspheres; Design Synthesis, Characterization, Assembly and Applications”, *Polym. J.*, **49**, 695-702, (2017), 10.1038/pj.2017.39
 10. 渡邊拓巳、*鈴木大介 “シード乳化重合による新奇複合ゲル微粒子の合成とその構造制御”, *Colloid & Interface Communication*, **42**, 2, (2017)
 11. S. Hiroshige, T. Kureha, D. Aoki, J. Sawada, D. Aoki, *T. Takata, *D. Suzuki, “Formation of Tough Films via the Evaporation of Water from Dispersions of Elastomer Microspheres Crosslinked with Rotaxane Supramolecules”, *Chem. Eur. J.*, **23**, 8405-8408, (2017), 10.1002/chem.201702077
 12. S. Matsui, T. Kureha, S. Hiroshige, M. Shibata, *T. Uchihashi, *D. Suzuki “Fast Adsorption of Soft Hydrogel Microspheres on Solid Surfaces in Aqueous Solution”, *Angew. Chem. Int'l. Ed.*, **56**, 12146 -12149, (2017), 10.1002/anie.201705808
 13. T. Kureha, D. Aoki, S. Hiroshige, K. Iijima, D. Aoki, *T. Takata, *D. Suzuki, “Decoupled Thermo- and pH-responsive Hydrogel Microspheres Cross-linked by Rotaxane Networks”, *Angew. Chem. Int'l. Ed.*, **56**, 15393-15396, (2017), 10.1002/ange.201709633
 14. T. Kureha, Y. Nishizawa, *D. Suzuki, “Controlled Separation and Release of Organiodine Compounds using Poly(2-methoxyethyl acrylate)-analogue Microspheres”, *ACS Omega*, **2**, 7686-7694 (2017), 10.1021/acsomega.7b01556
 15. T. Kureha, S. Hiroshige, S. Matsui, *D. Suzuki, “Water-immiscible Bioinert Coatings and Film Formation from Aqueous Dispersions of Poly(2-methoxyethyl acrylate)Microspheres”, *Colloids Surf. B*, **155**, 166-172, (2017), 10.1016/j.colsurfb.2017.04.001
 16. C. Kobayashi, T. Watanabe, K. Murata, T. Kureha, *D. Suzuki, “Localization of Polystyrene Particles on the Surface of Poly(N-isopropylacrylamide-co-methacrylic acid) Microgels Prepared by Seeded Emulsion Polymerization of Styrene”, *Langmuir*, **32**, 1429-1439, (2016), 10.1021/acs.langmuir.5b03698
 17. S. Minami, T. Watanabe, *D. Suzuki, *K. Urayama, “Rheological Properties of Suspensions of Thermo-responsive Poly(N-isopropylacrylamide) Microgels Undergoing Volume Phase Transition”, *Polym. J.*, **48**, 1079-1086, (2016), 10.1038/pj.2016.79
 18. T. Watanabe, C. Kobayashi, C. Song, K. Murata, T. Kureha, *D. Suzuki, “Impact of Spatial Distribution of Charged Groups in Core Poly(N-isopropyl acrylamide)-Based Microgels on the Resultant Composite Structures Prepared by Seeded Emulsion Polymerization of Styrene”, *Langmuir*, **32**, 12760-12773, (2016), 10.1021/acs.langmuir.6b03172
 19. T. Kureha, T. Shibamoto, S. Matsui, T. Sato, *D. Suzuki, “Investigation of Changes in the Microscopic Structure of Anionic Poly(*N*-isopropylacrylamide-*co*-Acrylic acid) Microgels in the Presence of Cationic Organic Dyes toward Precisely Controlled Uptake/Release of Low-molecular-weight Chemical Compound”, *Langmuir*, **32** (18), 4575–4585, (2016), 10.1021/acs.langmuir.6b00760
 20. K. Horigome, T. Ueki, *D. Suzuki, “Direct Visualization of Swollen Microgels via Scanning Electron Microscopy Using Ionic Liquids”, *Polym. J.*, **48**, 273–279, (2016), 10.1038/pj.2015.103
 21. *鈴木大介, 呉羽拓真 “ソフトヒドロゲル微粒子の表面・内部構造と機能”
高分子1月号 Vol.64 「高分子科学最近の進歩」, 30-34 (2016), 高分子学会
 22. S. Matsui, T. Kureha, K. Okeyoshi, R. Yoshida, T. Sato, *D. Suzuki, “Small-angle X-ray Scattering

- Study on Internal Microscopic Structures of Poly(*N*-isopropylacrylamide-*cot*ris(2,2'-bipyridyl)ruthenium(II) Complex Microgels”, *Langmuir*, **31**, 7228-7237, (2015), [10.1021/acs.langmuir.5b01164](https://doi.org/10.1021/acs.langmuir.5b01164)
23. D. Suzuki, K. Shibata, A. Tsuchida, *T. Okubo, “Thermo-sensitive Colloidal Crystals Composed of Monodisperse Colloidal Silica- and Poly(*N*-isopropyl acrylamide) Gel-Spheres”, *Colloid Polym Sci.*, **293**, 2763-2769, (2015), [10.1007/s00396-015-3661-1](https://doi.org/10.1007/s00396-015-3661-1)
24. 鈴木大介、“刺激応答性ヒドロゲル微粒子の創製と構造評価-アクリルアミド誘導体から出発する単分散ゲル微粒子の展開-”, 化学と工業 6月号 Vol.68 「ゲル微粒子ソフトな微粒子が醸し出す豊かな機能」, 498-500 (2015), 日本化学会
25. *D. Suzuki, C. Kobayashi, “Raspberry-shaped Composite Microgel Synthesis by Seeded Emulsion Polymerization with Hydrogel Particles”, *Langmuir*, **30**(24), 7085-7092 (2014), [10.1021/la5017752](https://doi.org/10.1021/la5017752)
26. T. Kureha, T. Sato, *D. Suzuki, “Relationship between Temperature-Induced Changes in Internal Microscopic Structures of Poly(*N*-isopropylacrylamide) Microgels and Organic Dye Uptake Behavior” *Langmuir*, **30**(29), 8717-8725, (2014), [10.1021/la501838c](https://doi.org/10.1021/la501838c)
27. K. Urayama*, S. Cong, T. Saeki, S. Uratani, T. Takigawa, M. Murai, *D. Suzuki, “A Simple Feature of Yielding Behavior of Highly Dense Suspensions of Soft Micro-hydrogel Particles”, *Soft Matter*, **10**, 9486-9495, (2014), [10.1039/c4sm01841a](https://doi.org/10.1039/c4sm01841a)

【平岡秀一】

1. S. Kai, M. Nakagawa, T. Kojima, X. Li, M. Yamashina, M. Yoshizawa, *S. Hiraoka, “Steric Interaction between Neighboring Components Favors the Formation of Large Intermediates in the Self-Assembly Process of a Pd₂L₄ Capsule”, *Chem. Eur. J.*, **24**, 3965-3969, (2018), [1002/chem.201705253](https://doi.org/10.1002/chem.201705253)
2. T. Tateishi, W. Zhu, L. H. Foianesi-Takeshige, T. Kojima, K. Ogata, *S. Hiraoka, “Self-assembly of a Pd₄L₈ Double-walled Square Partly Takes Place through the Formation of Kinetically Trapped Species”, *Eur. J. Inorg. Chem.*, **2018**, 1192-1197 (2018), [10.1002/ejic.201800037](https://doi.org/10.1002/ejic.201800037)
3. Y. Matsumura, S. Iuchi, S. Hiraoka, *H. Sato, “Chiral Effects on the Final Step of an Octahedron-Shaped Coordination Capsule Self-Assembly”, *Phys. Chem. Chem. Phys.*, **20**, 7383-7386, (2018), [10.1039/C7CP08237A](https://doi.org/10.1039/C7CP08237A)
(selected as Hot Paper and Back Cover)
4. T. Tateishi, T. Kojima, *S. Hiraoka, “Multiple Pathways in the Self-assembly Process of a Pd₄L₈ Coordination Tetrahedron”, *Inorg. Chem.*, **57**, 2686-2694, (2018), [10.1021/acs.inorgchem.7b03085](https://doi.org/10.1021/acs.inorgchem.7b03085)
5. Y.-Y. Zhan, K. Ogata, T. Kojima, T. Koide, K. Ishii, T. Mashiko, M. Tachikawa, S. Uchiyama, *S. Hiraoka, “Hyperthermostable Cube-shaped Assembly in Water”, *Comms. Chem.*, **1**, 14, (2018), [10.1038/s42004-018-0014-2](https://doi.org/10.1038/s42004-018-0014-2)
6. S. Kai, S. P. Maddala, T. Kojima, S. Akagi, K. Harano, E. Nakamura, *S. Hiraoka, “Flexibility of Components Alters the Self-assembly Pathway of Pd₂L₄ Coordination Cages”, *Dalton Trans.*, **47**, 3258-3263, (2018), [10.1039/C8DT00112J](https://doi.org/10.1039/C8DT00112J)
7. N. Tanaka, Y.-Y. Zhan, Y. Ozawa, T. Kojima, T. Koide, T. Mashiko, U. Nagashima, M. Tachikawa, *S. Hiraoka, “Semi-quantitative Evaluation of Molecular Meshing by Surface Analysis with Varying Probe Radii”, *Chem. Commun.*, **54**, 3335-3338, (2018), [10.1039/c8cc00695d](https://doi.org/10.1039/c8cc00695d)
8. *R. Harada, T. Mashiko, M. Tachikawa, S. Hiraoka, *Y. Shigeta, “Programed Dynamical Ordering in the Self-organization Processes of a Nanocube: A Molecular Dynamics Study”, *Phys. Chem. Chem. Phys.*, **20**, 9115-9122, (2018), [10.1039/C8CP00284C](https://doi.org/10.1039/C8CP00284C)
9. T. Tateishi, T. Kojima, *S. Hiraoka, “Chiral Self-sorting Process in the Self-assembly of Homochiral Coordination Cages from Axially Chiral Ligands”, *Comms Chem.*, **1**, 20, (2018), [10.1038/s42004-018-0020-4](https://doi.org/10.1038/s42004-018-0020-4)
10. 増子貴子, 平岡秀一, 長嶋雲兵, *立川仁典, “歯車状両親媒性分子からなるナノキューブの置換基効果と溶媒効果の理論的研究”, *J. Comp. Chem. Jpn.*, **17**, 31-37, (2018), [10.2477/jccj.2018-0010](https://doi.org/10.2477/jccj.2018-0010)
11. *S. Hiraoka, “Unresolved Issues that Remain in Molecular Self-Assembly”, *Bull. Chem. Soc. Jpn.*, **91**, 957-978, (2018), [10.1246/bcsj.20180008](https://doi.org/10.1246/bcsj.20180008)
12. S. Kai, T. Kojima, F. L. Thorp-Greenwood, M. J. Hardie, *S. Hiraoka, “How Does Chiral Self-sorting Take Place in the Formation of Homochiral Pd₆L₈ Capsules Consisting of Cyclotrimeratrylene-based Chiral Tritopic Ligands?”, *Chem. Sci.*, **9**, 4104-4108, (2018), [10.1039/C8SC01062E](https://doi.org/10.1039/C8SC01062E)

13. Y.-Y. Zhan, N. Tanaka, Y. Ozawa, T. Kojima, T. Mashiko, U. Nagashima, M. Tachikawa, *S. Hiraoka, “Importance of Molecular Meshing for the Stabilization of Solvophobic Assemblies”, *J. Org. Chem.*, **83**, 5132–5137, (2018), [10.1021/acs.joc.8b00495](https://doi.org/10.1021/acs.joc.8b00495)
14. Y.-Y. Zhan, T. Kojima, T. Koide, M. Tachikawa, *S. Hiraoka. “A Balance Between van der Waals and Cation-π Interactions That Stabilizes Hydrophobic Assemblies”, *Chem. Eur. J.*, **83**, 5132–5137, (2018), [10.1002/chem.201801376](https://doi.org/10.1002/chem.201801376)
15. M. Nakagawa, S. Kai, T. Kojima, *S. Hiraoka, “Energy-Landscape-Independent Kinetic Trap of Incomplete Cage in the Self-assembly of a Pd₂L₄ Cage”, *Chem. Eur. J.*, **24**(35), 8696–8696, (2018), [10.1002/chem.201801183](https://doi.org/10.1002/chem.201801183)
16. Y. Matsumura, S. Iuchi, S. Hiraoka, *H. Sato, “Chiral Effects on the Final Step of an Octahedron-shaped Coordination Capsule Self-assembly”, *Phys. Chem. Chem. Phys.*, **20**, 7383–7386, (2018), [10.1039/C7CP08237A](https://doi.org/10.1039/C7CP08237A)
17. A. Baba, T. Kojima, *S. Hiraoka, “Quantitative Analysis of Self-Assembly Process of Hexagonal Pt(II) Macroyclic Complexes: Effect of Solvent and Components”, *Chem. Eur. J.*, **24**, 838–847, (2018), [10.1002/chem.201702955](https://doi.org/10.1002/chem.201702955)
18. S. Kai, V. Martí-Centelles, Y. Sakuma, T. Mashiko, T. Kojima, U. Nagashima, M. Tachikawa, P. J. Lusby, *S. Hiraoka, “Quantitative Analysis of Self-Assembly Process of a Pd₂L₄ Cage Consisting of Rigid Ditopic Ligands” *Chem. Eur. J.*, **24**, 663–671, (2018), [10.1002/chem.201704285](https://doi.org/10.1002/chem.201704285)
19. V. E. Pritchard, D. R. Martir, S. Oldknow, S. Kai, S. Hiraoka, N. J. Cookson, *E. Zysman-Colman, *M. J. Hardie, “Homochiral Self-Sorted and Emissive Ir^{III} Metallo-Cryptophanes” *Chem. Eur. J.*, **23**, 6290–6294, (2017), [10.1002/chem.201701348](https://doi.org/10.1002/chem.201701348)
20. Y. Matsumura, S. Hiraoka, *H. Sato, “A Reaction Model on the Self-assembly Process of Octahedron-shaped Coordination Capsules”, *Phys. Chem. Chem. Phys.*, **19**, 20338–20342, (2017), [10.1039/c7cp03493h](https://doi.org/10.1039/c7cp03493h)
21. S. Kai, Y. Sakuma, T. Mashiko, T. Kojima, M. Tachikawa, *S. Hiraoka, “The Effect of Solvent and Coordination Environment of Metal Source on the Self-Assembly Pathway of a Pd(II)-mediated Coordination Capsule”, *Inorg. Chem.*, **56**, 12652–12663, (2017), [10.1021/acs.inorgchem.7b02152](https://doi.org/10.1021/acs.inorgchem.7b02152)
22. S. Kai, T. Shigeta, T. Kojima, *S. Hiraoka, “Quantitative Analysis of Self-assembly Process of a Pd_{1.2}L₂ Coordination Sphere”, *Chem. Asian J.*, **12**, 3203–3207, (2017), [10.1002/asia.201701351](https://doi.org/10.1002/asia.201701351)
23. T. Mashiko, S. Hiraoka, U. Nagashima, *M. Tachikawa, “Theoretical Study on Substituent and Solvent Effects for Nanocubes Formed with Gear-shaped Amphiphile Molecules”, *Phys. Chem. Chem. Phys.*, **19**, 1627–1631, (2017), [10.1039/C6CP07754D](https://doi.org/10.1039/C6CP07754D)
24. V. E. Pritchard, D. R. Martir, S. Oldknow, S. Kai, S. Hiraoka, N. J. Cookson, *E. Zysman-Colman, *M. J. Hardie, “Homochiral Self-sorted and Emissive Ir^{III} metallo-cryptophanes”, *Chem. Eur. J.* **23**, (2017) , **23**, 6290-6294, [10.1002/chem.201701348](https://doi.org/10.1002/chem.201701348)
25. S. Hiraoka, “What Do We Learn from the Molecular Self-Assembly” *Chem. Rec.*, **15**, 1144–1147 (2015), [10.1002/tcr.201510005](https://doi.org/10.1002/tcr.201510005)
26. A. Baba, T. Kojiam, *S. Hiraoka, “Self-Assembly Process of Dodecanuclear Pt(II)-Linked Cyclic Hexagon”, *J. Am. Chem. Soc.*, **137**, 7664–7667, (2015), [10.1021/jacs.5b04852](https://doi.org/10.1021/jacs.5b04852)
27. *T. Kojima, *S. Hiraoka, “Mesityllithium and *p*(dimethylamino)Phenyllithium for the Selective Alternate Trilithiation of the Hexaphenylbenzene Framework”, *Chem. Commun.*, **50**, 10420–10423, (2014), [10.1039/C4CC04520C](https://doi.org/10.1039/C4CC04520C)
28. *T. Kojima, *S. Hiraoka, “Selective Alternate Derivatization of the Hexaphenylbenzene Framework through a Thermodynamically Controlled Halogen Dance”, *Org. Lett.*, **16**(3), 1024–1027, (2014), [10.1021/ol500041j](https://doi.org/10.1021/ol500041j)
29. T. Mashiko, K. Yamada, T. Kojima, S. Hiraoka, U. Nagashima, *M. Tachikawa, “Molecular Dynamics and Principal Component Analysis for a Self-assembled Nanocube in Aqueous Solution” *Chem. Lett.*, **43**(3), 366–368, (2014), [10.1246/cl.130928](https://doi.org/10.1246/cl.130928)
30. Y. Tsujimoto, T. Kojima, *S. Hiraoka, “Rate-determining Step in the Self-assembly Process of Supramolecular Coordination Capsules”, *Chem. Sci.*, **5**, 4167–4172, (2014), [10.1039/C4SC01652A](https://doi.org/10.1039/C4SC01652A)
31. T. Mashiko, K. Yamada, S. Hiraoka, U. Nagashima, *M. Tachikawa, “Molecular Dynamics Simulation of Self-assembled Nanocubes in Methanol”, *Mol. Simulation*, **41**, 845–849, (2014), [10.1080/08927022.2014.940523](https://doi.org/10.1080/08927022.2014.940523)
32. J. Koseki, Y. Kita, S. Hiraoka, U. Nagashima, *M. Tachikawa, “Temperature Dependence of Self-

assembled Molecular Capsules Consisting of Gear-shaped Amphiphile Molecules with Molecular Dynamics Simulations”, *Int. J. Quan. Chem.*, **113**(4), 397-400, (2013), 10.1002/qua.24108

【二井勇人】

1. *E. Futai, S. Osawa, T. Cai, T. Fujisawa, S. Ishiura, T. Tomita, “Suppressor Mutations for Presenilin 1 Familial Alzheimer Disease Mutants Modulate γ -secretase Activities,” *J. Biol. Chem.*, **291**, 435-446, (2016), 10.1074/jbc.M114.629287
2. 二井勇人 “膜内切断プロテアーゼによるタンパク質分解の制御機構” 日本応用酵素協会誌, **49**, 17-28, (2015)
3. T. Onodera, E. Futai, E. Kan, N. Abe, T. Uchida, Y. Kamio, *J. Kaneko, “Phosphatidylethanolamine Plasmalogen Enhances the Inhibiting Effect of Phosphatidylethanolamine on γ -secretase activity” *J. Biochem.*, **157** (5), 301-309, (2014), [10.1093/jb/mvu074](https://doi.org/10.1093/jb/mvu074)

【二木史朗】

1. K. Shinoda, S. Tsuji, S. Futaki, *M. Imanishi, “Nested PUF Proteins: Extending Target RNA Elements for Gene Regulation”, *ChemBioChem.*, **19**, 171, (2018), 10.1002/cbic.201700458
2. K. Sakagami, T. Masuda, K. Kawano, *S. Futaki, “Importance of Net Hydrophobicity in the Cellular Uptake of All-hydrocarbon Stapled Peptides,” *Mol. Pharm.*, **15**, 1332, (2018), 10.1021/acs.molpharmaceut.7b01130
3. *M. Imanishi, S. Tsuji, A. Suda, S. Futaki, “Detection of N^6 -methyladenosine Based on the Methyl-sensitivity of MazF RNA Endonuclease,” *Chem. Commun.*, **53**, 12930, (2017), 10.1039/c7cc07699a
4. *I. Nakase, K. Noguchi, A. Aoki, T. Takatani-Nakase, I. Fujii, S. Futaki, “Arginine-rich Cell-penetrating Peptide-modified Extracellular Vesicles for Active Macropinocytosis Induction”, *Sci. Rep.*, **7**, 1991, (2017), [10.1038/s41598-017-02014-6](https://doi.org/10.1038/s41598-017-02014-6)
5. M. Akishiba, T. Takeuchi, Y. Kawaguchi, K. Sakamoto, H. Yu, I. Nakase, T. Takatani-Nakase, F. Madani, A. Gräslund, *S. Futaki, “Cytosolic Antibody Delivery by Lipid-sensitive Endosomolytic Peptide”, *Nat. Chem.* **9**, 751, (2017), [10.1038/NCHEM.2779](https://doi.org/10.1038/NCHEM.2779)
6. T. Murayama, T. Masuda, S. Afonin, K. Kawano, T. Takatani-Nakase, H. Ida, Y. Takahashi, T. Fukuma, A. S. Ulrich, *S. Futaki, “Loosening of Lipid Packing Promotes Oligoarginine Entry into Cells”, *Angew. Chem. Int. Ed., Engl.*, **56**(26), 7644-7647, (2017), [10.1002/anie.201703578](https://doi.org/10.1002/anie.201703578) (German version: 10.1002/ange.201703578)
7. *二木史朗, “アルギニンに富む膜透過ペプチドの細胞内移行”, *生化学*, **89**(1), 8–14, (2017), [10.14952/SEIKAGAKU.2017.890008](https://doi.org/10.14952/SEIKAGAKU.2017.890008)
8. *S. Futaki, I. Nakase, “Cell-Surface Interactions on Arginine-Rich Cell-Penetrating Peptides Allow for Multiplex Modes of Internalization”, *Acc. Chem. Res.*, **50**, 2449-2456, (2017), [10.1021/acs.accounts.7b00221](https://doi.org/10.1021/acs.accounts.7b00221)
9. A. Oku, M. Imanishi, D. Noshiro, T. Murayama, T. Takeuchi, I. Nakase, *S. Futaki, “Use of Calmodulin EF-hand Peptides as Ca^{2+} -switchable Recognition Tags”, *Biopolymers, Pept. Sci.*, **108**(1), e22937, (2017), [10.1002/bip.22937](https://doi.org/10.1002/bip.22937)
10. *I. Nakase, K. Noguchi, I. Fujii, S. Futaki, “Vectorization of Biomacromolecules into Cells Using Extracellular Vesicles with Enhanced Internalization Induced by Macropinocytosis”, *Sci. Rep.*, **6**, 34937, (2016), [10.1038/srep34937](https://doi.org/10.1038/srep34937)
11. K. Motoyama, R. Nishiyama, Y. Maeda, T. Higashi, Y. Kawaguchi, S. Futaki, Y. Ishitsuka, Y. Kondo, T. Irie, T. Era, *H. Arima, “Cholesterol-lowering Effect of Octaarginine-appended β -Cyclodextrin in Npc1-trap-CHO Cells”, *Biol. Pharm. Bull.*, **39**, 1823, (2016), [10.1248/bpb.b16-00369](https://doi.org/10.1248/bpb.b16-00369)
12. *I. Nakase, N. Ueno, M. Katayama, K. Noguchi, T. Takatani-Nakase, N. B. Kobayashi, T. Yoshida, I. Fujii, S. Futaki, “Receptor Clustering and Activation by Multivalent Interaction through Recognition Peptides Presented on Exosomes”, *Chem. Commun.*, **53**, 317, (2016), [10.1039/C6CC06719K](https://doi.org/10.1039/C6CC06719K)
13. C. M. Backlund, F. Sgolastra, R. Otter, L. Minter, T. Takeuchi, S. Futaki, *G. N. Tew, “Increased Hydrophobic Block Length of PTDMs Promotes Protein Internalization”, *Polym. Chem.*, **7**, 7514, (2016), [10.1039/C6PY01615D](https://doi.org/10.1039/C6PY01615D)
14. C. M. Backlund, T. Takeuchi, S. Futaki, *G. N. Tew. “Relating structure and internalization for ROMP-based protein mimics,” *Biochim. Biophys. Acta*, **1858**, 1443-1450, (2016), [10.1016/j.bbamem.2016.03.024](https://doi.org/10.1016/j.bbamem.2016.03.024)
15. Y. Kawaguchi, T. Takeuchi, K. Kuwata, J. Chiba, Y. Hatanaka, I. Nakase, *S. Futaki, “Syndecan-4 Is a

- Receptor for Clathrin-Mediated Endocytosis of Arginine-Rich Cell-Penetrating Peptides,” *Bioconjug. Chem.*, **27**, 1119-1130, (2016), 10.1021/acs.bioconjchem.6b00082
16. T. Murayama, S. Pujals, H. Hirose, I. Nakase, *S. Futaki “Effect of Amino Acid Substitution in the Hydrophobic Face of Amphiphilic Peptides on Membrane Curvature and Perturbation: N-Terminal Helix Derived From Adenovirus Internal Protein VI As a Model,” *Biopolymers*, **106**, 430-439, (2015), 10.1002/bip.22797
 17. R. Miyajima, Y. Tsuda, T. Inokuma, A. Shigenaga, M. Imanishi, S. Futaki, *A. Otaka, “Preparation of Peptide Thioesters from Naturally Occurring Sequences Using Reaction Sequence Consisting of Regioselective S-cyanylation and Hydrazinolysis,” *Biopolymers*, **106**(4), 531-46, (2015), 10.1002/bip.22757
 18. Y. Kuroda, N. *Kato-Kogoe, E. Tasaki, M. Yuasa-Sunagawa, K. Yamanegi, K. Nakasyo, I. Nakase, S. Futaki, Y. Tohyama, M. Hirose, “Suppressive Effect of Membrane-permeable Peptides Derived from Autophosphorylation Sites of the IGF-1 Receptor on Breast Cancer Cells”, *Eur. J. Pharmacol.*, **765**, EJP41801, (2015), 10.1016/j.ejphar.2015.08.004
 19. I. Nakase, T. Takeuchi, *S. Futaki, “Cell Penetrating Peptides for Chemical Biological Studies”, *Methods Mol. Biol.*, **1324**, 387-396, (2015), 10.1007/978-1-4939-2806-4_26
 20. T. Takeuchi, M. Suzuki, N. Fujikake, H. A. Popiel, H. Kikuchi, S. Futaki, K. Wada, *Y. Nagai, “Intercellular Chaperone Transmission via Exosomes Contributes to Maintenance of Protein Homeostasis at the Organismal Level,” *Proc. Natl. Acad. Sci. U.S.A.*, **112**, E2497-2506, (2015), 10.1073/pnas.1412651112
 21. *I. Nakase, S. Futaki, “Combined Treatment with a pH-sensitive Fusogenic Peptide and Cationic Lipids Achieves Enhanced Cytosolic Delivery of Exosomes,” *Sci. Rep.*, **5**, 10112, (2015), 10.1038/srep10112
 22. *二木史朗, “細胞内・遺伝子デリバリー,” *日本防菌防黴学会誌*, 43, 259 (2015)
 23. I. Nakase, Y. Kawaguchi, M. Nomizu, *S. Futaki “Cellular Uptake of Arginine-Rich Cell-Penetrating Peptides and the Contribution of Membrane-Associated Proteoglycans,” *Trends Glycosci. Glycotech.*, **27**, 81, (2015), 10.4052/tigg.1420.1
 24. Y. Azuma, T. Kükenshöner, G. Ma, J. Yasunaga, M. Imanishi, G. Tanaka, I. Nakase, T. Maruno, Y. Kobayashi, *K. M. Arndt, *M. Matsuoka, *S. Futaki, “Controlling Leucine-zipper Partner Recognition in Cells Through Modification of α -g Interactions”, *Chem. Commun.*, **50**, 6364-6867, (2014), 10.1039/c4cc00555d
 25. *I. Nakase, K. Osaki, G. Tanaka, A. Utani, *S. Futaki, “Molecular Interplays Involved in the Cellular Uptake of Octaarginine on Cell Surfaces and the Importance of Syndecan-4 Cytoplasmic V Domain for the Activation of Protein Kinase Ca”. *Biochem. Biophys. Res. Commun.*, **446**, 857-862, (2014), 10.1016/j.bbrc.2014.03.018
 26. *T. Takeuchi, H. A. Popiel, S. Futaki, K. Wada, *Y. Nagai. “Peptide-based Therapeutic Approaches for Treatment of the Polyglutamine Diseases”, *Curr. Med. Chem.*, **21**, 2575-2582, (2014), 10.2174/092986732166140217124038

【芳坂貴弘】

1. Y. Mori, *H. Okumura, T. Watanabe, T. Hohsaka, “Antigen-dependent Fluorescence Response of Anti-c-Myc Quenchbody Studied by Molecular Dynamics Simulations”, *Chem. Phys. Lett.*, **698**, 223-226, (2018), 10.1016/j.cplett.2018.03.011
2. M. Yamaguchi, E. Ohta, T. Muto, T. Watanabe, T. Hohsaka, Y. Yamazaki, H. Kamikubo, *M. Kataoka, “Statistical Description of the Denatured Structure of a Single Protein, Staphylococcal Nuclease, by FRET Analysis”, *Biophys. Rev.*, **10**, 145-152, (2018), 10.1007/s12551-017-0334-y
3. S. Ahmed, T. Nakaji-Hirabayashi, T. Watanabe, T. Hohsaka, *K. Matsumura, “Freezing Assisted Gene Delivery Combined with Polyampholyte Nanocarriers”, *ACS Biomater. Sci. Eng.*, **3**, 1677-1689, (2017), 10.1021/acsbiomaterials.7b00176
4. A. Yamaguchi, T. Matsuda, K. Otake T. Yanagisawa, S. Yokoyama, Y. Fujiwara, T. Watanabe, *T. Hohsaka, *K. Sakamoto, “Incorporation of a Doubly Functionalized Synthetic Amino Acid into Proteins for Creating Chemical and Light-Induced Conjugates”, *Bioconjugate Chem.*, **27**, 198–206, (2016), 10.1021/acs.bioconjchem.5b00602
5. K. P. Huynh Nhat, T. Watanabe, K. Yoshikoshi, *T. Hohsaka, “Antibody-based Fluorescent and Fluorescent Ratiometric Indicators for Detection of Phosphotyrosine”, *J. Biosci. Bioeng.*, **122**, 146-154,

- (2016), 10.1016/j.jbiosc.2016.01.010
6. K. Yoshikoshi, T. Watanabe, *T. Hohsaka, "Double-Fluorescent-Labeled Single-Chain Antibodies Showing Antigen-Dependent Fluorescence Ratio Change", *Bull. Chem. Soc. Jpn.*, **89**, 573-580, (2016), 10.1246/bcsj.20150384
 7. A. Uyeda, T. Watanabe, Y. Kato, H. Watanabe, T. Yomo, *T. Hohsaka, *T. Matsuura, "Liposome-Based in Vitro Evolution of Aminoacyl-tRNA Synthetase for Enhanced Pyrrolysine Derivative Incorporation", *ChemBioChem.*, **16**, 1797-1802, (2015), 10.1002/cbic.201500174
 8. T. Ezure, K. Nanatani, Y. Sato, S. Suzuki, K. Aizawa, S. Souma, M. Ito, T. Hohsaka, G. von Heijine, T. Utsumi, K. Abe, E. Ando, N. Uozumi*, "A Cell-free Translocation System Using Extracts of Cultured Insect Cells to Yield Functional Membrane Proteins", *PLoS One.*, **9**, e112874, (2014), 10.1371/journal.pone.0112874
 9. Y. Ito, *T. Hohsaka, "Incorporation of Fluorescent Nonnatural Amino Acid into Sialic Acid-Binding Lectin for Fluorescence Detection of Ligand-Binding", *Bull. Chem. Soc. Jpn.*, **86**(6), 729-735, (2013), 10.1246/bcsj.20120345
 10. T. Matsubara, K. Iijima, T. Watanabe, T. Hohsaka, *T. Sato, "Incorporation of Glycosylated Amino Acid into Protein by an in Vitro Translation System", *Bioorg. Med. Chem. Lett.*, **23**(20), 5634-5636, (2013), 10.1016/j.bmcl.2013.08.035
- 【松浦友亮】
1. *A. Uyeda, T. Watanabe, T. Hohsaka, T. Matsuura, "Different Protein Localizations on the Inner and Outer Leaflet of Cell-sized Liposomes Using Cell-free Protein Synthesis", *Synth Biol.*, **3**, in press, (2018), 10.1093/synbio/ysy007
 2. *T. Matsuura, K. Hosoda, Y. Shimizu, "Robustness of a Reconstituted *Escherichia coli* Protein Translation System Analyzed by Computational Modeling", *ACS Synth Biol.*, **7**(8), 1964-1972, (2018), 10.1021/acssynbio.8b00228
 3. M. Iwamoto, M.A. Elfaramawy, M. Yamatake, T. Matsuura, S. Oiki, "Concurrent In Vitro Synthesis and Functional Detection of Nascent Activity of the KcsA Channel under a Membrane Potential", *ACS Synth Biol.*, **7**, 1004-1011, (2018), 10.1021/acssynbio.7b00454
 4. *T. Matsuura, N. Tanimura, K. Hosoda, Y. Yomo, Y. Shimizu, "Reaction Dynamics Analysis of a Reconstituted *Escherichia coli* Protein Translation System by Computational Modeling", *Proc. Natl. Acad. Sci. USA*, **114**, E1336-E1344, (2017), 10.1073/pnas.1615351114
 5. A. Uyeda, S. Nakayama, Y. Kato, H. Watanabe, T. *Matsuura, "Construction of an *in vitro* Gene Screening System of the *E. coli* EmrE Transporter Using Liposome Display", *Anal. Chem.*, **88**, 12028-12035, (2016), 10.1021/acs.analchem.6b02308
 6. N. Ohta, Y. Kato, H. Watanabe, H. Mori, *T. Matsuura, "In vitro Membrane Protein Synthesis Inside Sec Translocon-reconstituted Cell-sized Liposomes", *Sci. Rep.*, **6**, 36466, (2016), 10.1038/srep36466
 7. A. Uyeda, T. Watanabe, Y. Kato, H. Watanabe, T. Yomo, *T. Hohsaka, *T. Matsuura, "Liposome-Based in Vitro Evolution of Aminoacyl-tRNA Synthetase for Enhanced Pyrrolysine Derivative Incorporation", *Chembiochem*, **16**, 1797-1802, (2015), 10.1002/cbic.201500174
 8. S. Fujii, T. Matsuura, *T. Yomo, "Membrane Curvature Affects the Formation of alpha-Hemolysin Nanopores", *ACS Chem Biol.*, **10**, 1694-1701, (2015), 10.1021/acschembio.5b00107
 9. S. Fujii, T. Matsuura, *T. Yomo, "In Vitro Directed Evolution of Alpha-hemolysin by Liposome Display", *Biophysics*, **11**, 67-72, (2015), 10.2142/biophysics.11.67
 10. K. Usui, N. Ichihashi, Y. Kazuta, T. Matsuura, *T. Yomo, "Effects of Ribosomes on the Kinetics of Qbeta Replication", *FEBS Lett.*, **588**, 117-123, (2014), 10.1016/j.febslet.2013.11.018
 11. K. Uno, T. Sunami, N. Ichihashi, Y. Kazuta, T. Matsuura, *T. Yomo, "The Evolutionary Enhancement of Genotype-phenotype Linkages in the Presence of Multiple Copies of Genetic Material", *Chembiochem*, **15**, 2281-2288, (2014), 10.1002/cbic.201402299
 12. H. Soga, S. Fujii, T. Yomo, Y. Kato, H. Watanabe, *T. Matsuura, "In Vitro Membrane Protein Synthesis Inside Cell-sized Vesicles Reveals the Dependence of Membrane Protein Integration on Vesicle Volume", *ACS Synth. Biol.*, **3**, 372-379, (2014), 10.1021/sb400094c
 13. T. Okano, T. Matsuura, H. Suzuki, *T. Yomo, "Cell-free Protein Synthesis in a Microchamber Revealed the Presence of an Optimum Compartment Volume for High-order Reactions", *ACS Synth. Biol.*, **3**, 347-352, (2014), 10.1021/sb400087e
 14. K. Nishimura, T. Matsuura, T. Sunami, S. Fujii, K. Nishimura, H. Suzuki, *T. Yomo, "Identification of

- Giant Unilamellar Vesicles with Permeability to Small Charged Molecules”, *RSC. Advances*, **4**, 35224, (2014), [10.1039/c4ra05332j](https://doi.org/10.1039/c4ra05332j)
15. Y. Kazuta, T. Matsuura, N. Ichihashi, *T. Yomo, “Synthesis of Milligram Quantities of Proteins Using a Reconstituted in Vitro Protein Synthesis System”, *J. Biosci. Bioeng.*, **118**(5), 554-557, (2014), [10.1016/j.jbiosc.2014.04.019](https://doi.org/10.1016/j.jbiosc.2014.04.019)
 16. S. Fujii, T. Matsuura, T. Sunami, T. Nishikawa, Y. Kazuta, *T. Yomo, “Liposome Display for in Vitro Selection and Evolution of Membrane Proteins”, *Nature protocols*, **9**, 1578-1591, (2014), [10.1038/nprot.2014.107](https://doi.org/10.1038/nprot.2014.107)
- 【三宅弘之】
1. T. Sagami, S. Umemoto, Y. O Tahara, M. Miyata, Y. Yonamine, D. Ishikawa, T. Mori, K. Ariga, H. Miyake, *S. Shinoda, “pH-Responsive Cotton Effects in the d-d Transition Band of Self-Assembling Copper(II) Complexes with a Cholesteryl-armed Ligand”, *Bull. Chem. Soc. Jpn.*, **90**, 739-745 (2017), [10.1246/bcsj.20170054](https://doi.org/10.1246/bcsj.20170054)
 2. T. Sagami, Y. O. Tahara, M. Miyata, H. Miyake, *S. Shinoda, “Luminescence Sensing of Weakly-Hydrated Anions in Aqueous Solution by Self-assembled Europium(III) Complexes”, *Chem. Commun.*, **53**, 3967-3970, (2017), [10.1039/C7CC00477J](https://doi.org/10.1039/C7CC00477J)
 3. T. Sagami, S. Umemoto, Y. O Tahara, M. Miyata, Y. Yonamine, D. Ishikawa, T. Mori, K. Ariga, H. Miyake, *S. Shinoda, “pH-Responsive Cotton Effects in the d-d Transition Band of Self-Assembling Copper(II) Complexes with a Cholesteryl-armed Ligand”, *Bull. Chem. Soc. Jpn.*, **90**, 739-745, (2017), [10.1246/bcsj.20170054](https://doi.org/10.1246/bcsj.20170054)
 4. J. Gregoliński, M. Hikita, T. Sakamoto, H. Sugimoto, H. Tsukube, *H. Miyake, “Redox-Triggered Helicity Inversion in Chiral Cobalt Complexes in Combination with H⁺ and NO₃⁻ Stimuli”, *Inorg. Chem.*, **55**, 633-643, (2016), [10.1021/acs.inorgchem.5b01902](https://doi.org/10.1021/acs.inorgchem.5b01902)
 5. *K. Singh, P. Kaur, *H. Miyake, H. Tsukube, “Supramolecular Chemistry Strategies for Naked-eye Detection and Sensing”, In *Synergy in Supramolecular Chemistry*, 301–320, (2014), T. Nabeshima, ed., *CRC Press*, ISBN 9781466595026
 6. *H. Miyake, “Supramolecular Chirality in Dynamic Coordination Chemistry”, *Symmetry*, **6**, 880-895, (2014). [10.3390/sym6040880](https://doi.org/10.3390/sym6040880)
- A03 : 動的秩序の展開
- 【稻垣直之】
1. T. Minegishi, Y. Uesugi, N. Kaneko, W. Yoshida, K. Sawamoto, *N. Inagaki, “Shootin1b Mediates a Mechanical Clutch to Produce Force for Neuronal Migration”, *Cell Rep.*, in press, (2018)
 2. K. Baba, W. Yoshida, M. Toriyama, T. Shimada, C. F. Manning, M. Saito, K. Kohno, J. S. Trimmer, R. Watanabe, *N. Inagaki, “Gradient-reading and Mechano-effector Machinery for Netrin-1-induced Axon Guidance”, *eLife*, **7**, e34593, (2018), [10.7554/eLife.34593](https://doi.org/10.7554/eLife.34593)
 3. K. Abe, H. Katsuno, M. Toriyama, K. Baba, T. Mori, T. Hakoshima, Y. Kanemura, R. Watanabe, *N. Inagaki, “Grip and slip of L1-CAM on Adhesive Substrates Direct Growth Cone Haptotaxis”, *Proc. Natl. Acad. Sci. USA*, **115**, 2764-2769, (2018), [10.1073/pnas.1711667115](https://doi.org/10.1073/pnas.1711667115)
 4. *N. Inagaki, H. Katsuno, “Actin Waves: Origin of Cell Polarization and Migration?”, *Trends in Cell Biology*, **27**, 515-526, (2017), [10.1016/j.tcb.2017.02.003](https://doi.org/10.1016/j.tcb.2017.02.003)
 5. Y. Higashiguchi, K. Katsuta, T. Minegishi, S. Yonemura, A. Urasaki, *N. Inagaki, “Identification of a Shootin1 Isoform Expressed in Peripheral Tissues”, *Cell Tissue Res.*, **366**, 75-87, (2016), [10.1007/s00441-016-2415-9](https://doi.org/10.1007/s00441-016-2415-9)
 6. K. Tahara, M. Tsukui, T. Maeno, N. Inagaki, *J. Kikuchi, “Efficient Solid-Phase Gene Delivery Mediated by Cerasome: Effect of Reverse Procedure on Transfection Performances in Comparison with Solution-Based Method”, *Chem. Lett.*, **44**(12), 1643-1645, (2015), [10.1246/cl.150777](https://doi.org/10.1246/cl.150777)
 7. Y. Kubo, K. Baba, M. Toriyama, Y. Minegishi, T. Sugiura, S. Kozawa, K. Ikeda, *N. Inagaki, “Shootin1-cortactin Interaction Mediates Signal-force Transduction for Axon Outgrowth”, *J. Cell Biol.* **210**, 663-676, (2015), [10.1083/jcb.201505011](https://doi.org/10.1083/jcb.201505011)
 8. H. Katsuno, M. Toriyama, Y. Hosokawa, K. Mizuno, K. Ikeda, Y. Sakumura, N. Inagaki, “Actin Migration Driven by Directional Assembly and Disassembly of Membrane Anchored Actin Filaments”, *Cell Reports*, **12**, 648-660, (2015), [10.1016/j.celrep.2015.06.048](https://doi.org/10.1016/j.celrep.2015.06.048)
 9. 馬場健太郎, 浦崎明宏, 稲垣直之, ラージゲルプロテオミクスを基盤とした神経細胞の軸索形成とガイダンスの解析, 電気泳動 **58**, 49-52, (2014)

10. S. Kozawa, Y. Sakumura, M. Toriyama, N. Inagaki, *K. Ikeda, "Bayesian Cell Force Estimation Considering Force Directions", *Neural Process Lett.*, **41**, 191-200, (2013), [10.1007/s11063-013-9320-y](https://doi.org/10.1007/s11063-013-9320-y)
11. K. Tahara, T. Moriuchi, M. Tsukui, A. Hirota, T. Maeno, M. Toriyama, N. Inagaki, *J. Kikuchi, "Ceramic Coating of Liposomal Gene Carrier for Minimizing Toxicity to Primary Hippocampal Neurons", *Chem. Lett.*, **42**(10), 1265-1267, (2013), [10.1246/cl.130541](https://doi.org/10.1246/cl.130541)

【内山進】

1. T. Uchihashi, YH. Watanabe, Y. Nakazaki, T. Yamasaki, H. Watanabe, T. Maruno, K. Ishii, S. Uchiyama, C. Song, K. Murata, *R. Iino, T. Ando, "Dynamic Structural States of ClpB Involved in its Disaggregation function", *Nat. Commun.*, **9**, 2147, (2018), [10.1038/s41467-018-04587-w](https://doi.org/10.1038/s41467-018-04587-w)
2. Y. Zhan, K. Ogata, T. Kojima, T. Koide, K. Ishii, T. Mashiko, M. Tachikawa, S. Uchiyama, *S. Hiraoka, "Hyperthermostable Cube-shaped Assembly in Water", *Commun. Chem.*, **1**, 1-14, (2018), [10.1038/s42004-018-0014-2](https://doi.org/10.1038/s42004-018-0014-2)
3. R. Kim, S. Kanamaru, T. Mikawa, C. Prévost, K. Ishii, K. Ito, S. Uchiyama, M. Oda, H. Iwasaki, S. K Kim, *M. Takahashi, "RecA Requires Two Molecules of Mg²⁺ ions for its Optimal Strand Exchange Activity in Vitro", *Nucleic Acids Res.*, **46**: 2548-2559, (2018), [10.1093/nar/gky048](https://doi.org/10.1093/nar/gky048)
4. T. Maruno, H. Watanabe, S. Yoneda, T. Uchihashi, S. Adachi K. Arai, T. Sawaguchi, *S. Uchiyama, "Sweeping of Adsorbed Therapeutic Protein on Prefillable Syringes Promotes Micron Aggregate Generation", *J. Pharm. Sci.*, **107**, 1521-1529, (2018), [10.1016/j.xphs.2018.01.021](https://doi.org/10.1016/j.xphs.2018.01.021)
5. 石井健太郎、*内山 進, "タンパク質のネイティブ質量分析—動的なタンパク質複合体形成機構の解明にむけてー", *ぶんせき*, **10**, 472-475, (2017)
6. K. Ishii, M. Zhou, *S. Uchiyama, "Native Mass Spectrometry for Understanding Dynamic Protein Complex", *Biochim Biophys. Acta.*, **1862**, 275-286, (2017), [10.1016/j.bbagen.2017.09.019](https://doi.org/10.1016/j.bbagen.2017.09.019)
7. Q. Wang, R. Marchetti, S. Prsic, K. Ishii, Y. Arai, I. Ohta, S. Inuki, S. Uchiyama, A. Silipo, A. Molinaro, R. N. Husson, K. Fukase, *Y. Fujimoto, "A Comprehensive Study of the Interaction between Peptidoglycan Fragments and the Extracellular Domain of Mycobacterium tuberculosis Ser/Thr Kinase PknB", *ChemBioChem.*, **18**, 2094-2098, (2017), [10.1002/cbic.201700385](https://doi.org/10.1002/cbic.201700385)
8. *A. Ishii-Watabe, H. Shibata, A. Harazono, M. Hyuga, M. Kiyoshi, S. Saitoh, T. Iwura, T. Torisu, Y. Goda, S. Uchiyama, "Recent Topics of Research in the Characterization and Quality Control of Biopharmaceuticals in Japan", *J. Pharm. Sci.*, **106**, 3431-3437, (2017), [10.1016/j.xphs.2017.07.024](https://doi.org/10.1016/j.xphs.2017.07.024)
9. R. Phengchat, H. Takata, *S. Uchiyama, K. Fukui, "Calcium Depletion Destabilises Kinetochore Fibres by the Removal of CENP-F from the Kinetochore", *Sci. Rep.*, **7**, 7335, (2017), [10.1038/s41598-017-07777-6](https://doi.org/10.1038/s41598-017-07777-6)
10. R. Poonperm, H. Takata, S. Uchiyama, *K. Fukui, "Interdependency and Phosphorylation of KIF4 and Condensin I are Essential for Organization of Chromosome Scaffold", *PloS one*, **12**, e0183298, (2017), [10.1371/journal.pone.0183298](https://doi.org/10.1371/journal.pone.0183298)
11. N. Shinozaki, R. Hashimoto, K. Fukui, *S. Uchiyama, "Efficient Generation of Single Domain Antibodies with High Affinities and Enhanced Thermal Stabilities", *Sci. Rep.*, **7**, 5794, (2017), [10.1038/s41598-017-06277-x](https://doi.org/10.1038/s41598-017-06277-x)
12. T. Torisu, T. Maruno, Y. Hamaji, T. Ohkubo, *S. Uchiyama, "Synergistic Effect of Cavitation and Agitation on Protein Aggregation", *J Pharm Sci.*, **106**, 521-529, (2017), [10.1016/j.xphs.2016.10.015](https://doi.org/10.1016/j.xphs.2016.10.015)
13. T. Torisu, T. Maruno, S. Yoneda, Y. Hamaji, S. Honda, *T. Ohkubo, *S. Uchiyama, "Friability Testing as a New Stress-Stability Assay for Biopharmaceuticals", *J Pharm Sci.*, **106**, 2966-2978, (2017), [10.1016/j.xphs.2017.05.035](https://doi.org/10.1016/j.xphs.2017.05.035)
14. E. Krayukhina, M. Noda, K. Ishii, T. Maruno, H. Wakabayashi, M. Tada, T. Suzuki, A. Ishii-Watabe, M. Kato, *S. Uchiyama, "Analytical Ultracentrifugation with Fluorescence Detection System Reveals Differences in Complex Formation between Recombinant Human TNF and Different Biological TNF Antagonists in Various Environments", *MAbs.*, **9**, 664-679, (2017), [10.1080/19420862.2017.1297909](https://doi.org/10.1080/19420862.2017.1297909)
15. Z. Zhang, U. Ohto, T. Shibata, E. Krayukhina, M. Taoka, Y. Yamauchi, H. Tanji, T. Isobe, S. Uchiyama, K. Miyake, *T. Shimizu, "Structural Analysis Reveals that Toll-like Receptor 7 Is a Dual Receptor for Guanosine and Single-Stranded RNA," *Immunity*, **45**, 737-748, (2016), [10.1016/j.immuni.2016.09.011](https://doi.org/10.1016/j.immuni.2016.09.011)
16. S. Toma-Fukai, J. D. Kim, K. E. Park, N. Kuwabara, N. Shimizu, E. Krayukhina, S. Uchiyama, A. Fukamizu, *T. Shimizu, "Novel Helical Assembly in Arginine Methyltransferase 8," *J. Mol. Biol.*, **428**,

17. *M. Sugiyama, H. Yagi, K. Ishii, L. Porcar, A. Martel, K. Oyama, M. Noda, Y. Yunoki, R. Murakami, R. Inoue, N. Sato, Y. Oba, K. Terauchi, S. Uchiyama, *K. Kato, “Structural Characterization of the Circadian Clock Protein Complex Composed of KaiB and KaiC by Inverse Contrast-matching Small-angle Neutron Scattering,” *Sci Rep.*, **6**, 35567, (2016), [10.1038/srep35567](https://doi.org/10.1038/srep35567)
18. S. Seetaha, M. Yagi-Utsumi, T. Yamaguchi, K. Ishii, S. Hannongbua, *K. Choowongkomon, *K. Kato, “Application of Site-Specific Spin Labeling for NMR Detecting Inhibitor-Induced Conformational Change of HIV-1 Reverse Transcriptase,” *ChemMedChem.* **11**, 363-366, (2016), [10.1002/cmde.201500554](https://doi.org/10.1002/cmde.201500554)
19. *T. Satoh, T. Toshimori, M. Noda, S. Uchiyama, *K. Kato, “Interaction Mode between Catalytic and Regulatory Subunits in Glucosidase II Involved in ER Glycoprotein Quality Control,” *Protein Sci.*, **25**, 2095-2101, (2016), [10.1002/pro.3031](https://doi.org/10.1002/pro.3031)
20. R. Phengchat, *H. Takata, K. Morii, N. Inada, H. Murakoshi, S. Uchiyama, *K. Fukui, “Calcium Ions Function as a Booster of Chromosome Condensation,” *Sci. Rep.*, **6**, 38281, (2016), [10.1038/srep38281](https://doi.org/10.1038/srep38281)
21. *U. Ohto, H. Ishida, E. Krayukhina, S. Uchiyama, N. Inoue, *T. Shimizu, “Structure of IZUMO1-JUNO Reveals Sperm-oocyte Recognition During Mammalian Fertilization”, *Nature*, **534**, 566-569, (2016), [10.1038/nature18596](https://doi.org/10.1038/nature18596)
22. *M. Oda, Y. Tanabe, M. Noda, S. Inaba, E. Krayukhina, H. Fukada, S. Uchiyama, “Structural and Binding Properties of Laminarin Revealed by Analytical Ultracentrifugation and Calorimetric Analyses”, *Carbohydr Res.*, **431**, 33-38, (2016), [10.1016/j.carres.2016.05.008](https://doi.org/10.1016/j.carres.2016.05.008)
23. E. Nango, S. Akiyama, S. Maki-Yonekura, Y. Ashikawa, Y. Kusakabe, E. Krayukhina, T. Maruno, S. Uchiyama, N. Nuemket, K. Yonekura, M. Shimizu, N. Atsumi, N. Yasui, T. Hikima, M. Yamamoto, Y. Kobayashi, *A. Yamashita, “Taste Substance Binding Elicits Conformational Change of Taste Receptor T1r Heterodimer Extracellular Domains,” *Sci Rep.*, **6**, 25745, (2016), [10.1038/srep25745](https://doi.org/10.1038/srep25745)
24. A. Masato, F. Kiichi, *S. Uchiyama, “Suppression of Methionine Oxidation of a Pharmaceutical Antibody Stored in a Polymer-Based Syringe,” *J Pharm. Sci.*, **105**, 623-629, (2016), [10.1002/jps.24675](https://doi.org/10.1002/jps.24675)
25. *Y. Kabe, T. Nakane, I. Koike, T. Yamamoto, Y. Sugiura, E. Harada, K. Sugase, T. Shimamura, M. Ohmura, K. Muraoka, A. Yamamoto, T. Uchida, S. Iwata, Y. Yamaguchi, E. Krayukhina, M. Noda, H. Handa, K. Ishimori, S. Uchiyama, *T. Kobayashi, *M. Suematsu, “Haem-dependent Dimerization of PGRMC1/Sigma-2 Receptor Facilitates Cancer Proliferation and Chemoresistance,” *Nat. Commun.*, **7**, 11030, (2016), [10.1038/ncomms11030](https://doi.org/10.1038/ncomms11030)
26. *R. Inoue, T. Takata, N. Fujii, K. Ishii, S. Uchiyama, N. Sato, Y. Oba, K. Wood, K. Kato, N. Fujii, *M. Sugiyama, “New Insight into the Dynamical System of AlphaB-crystallin Oligomers,” *Sci Rep.*, **6**, 29208, (2016), [10.1038/srep29208](https://doi.org/10.1038/srep29208)
27. S. Uchiyama, “Biophysical Characterization of Biopharmaceuticals, Including Antibody Drugs”, *Yakugaku Zasshi*, **136**, 443-448, (2016), [10.1248/yakushi.15-00236-2](https://doi.org/10.1248/yakushi.15-00236-2)
28. A. Fujikawa, A. Nagahira, H. Sugawara, K. Ishii, S. Imajo, M. Matsumoto, K. Kuboyama, R. Suzuki, N. Tanga, M. Noda, S. Uchiyama, T. Tomoo, A. Ogata, M. Masumura, *M. Noda, “Small-molecule Inhibition of PTPRZ Reduces Tumor Growth in a Rat Model of Glioblastoma”, *Sci. Rep.*, **6**, 20473, (2016), [10.1038/srep20473](https://doi.org/10.1038/srep20473)
29. R. Thammaporn, K. Ishii, M. Yagi-Utsumi, S. Uchiyama, *S. Hannongbua, *K. Kato, “Mass Spectrometric Characterization of HIV-1 Reverse Transcriptase Interactions with Non-nucleoside Reverse Transcriptase Inhibitors”, *Biol Pharm Bull.*, **39**(3), 450-4, (2016), [10.1248/bpb.b15-00880](https://doi.org/10.1248/bpb.b15-00880)
30. K. Ishii, M. Noda, S. Uchiyama, “Mass Spectrometric Analysis of Protein-ligand Interaction”, *Biophys. Physicobiology*, **13**, 87-95, (2016), [10.2142/biophysico.13.0_87](https://doi.org/10.2142/biophysico.13.0_87)
31. K. Ishii, M. Noda, H. Yagi, R. Thammaporn, S. Seetaha, T. Satoh, *K. Kato, *S. Uchiyama, “Disassembly of the Self-assembled, Double-ring Structure of Proteasome α7 Homo-tetradecamer by α6”, *Sci. Rep.*, **5**, 18167, (2015), [10.1038/srep18167](https://doi.org/10.1038/srep18167)
32. S. Uchiyama, K. Kawahara, Y. Hosokawa, S. Fukakusa, H. Oki, S. Nakamura, Y. Kojima, M. Noda, R. Takino, Y. Miyahara, T. Maruno, Y. Kobayashi, T. Ohkubo, K. Fukui, “Structural Basis for Dimer Formation of Human Condensin Structural Maintenance of Chromosome Proteins and Its Implications for Single-stranded DNA Recognition”, *J Biol Chem.*, **290**(49), 29461-77, (2015), [10.1074/jbc.M115.670794](https://doi.org/10.1074/jbc.M115.670794)
33. R. Poonperm, H. Takata, T. Hamano, A. Matsuda, S. Uchiyama, Y. Hiraoka, *K. Fukui, “Chromosome

- Scaffold is a Double-Stranded Assembly of Scaffold Proteins” *Sci. Rep.* **5**, 11916, (2015), [10.1038/srep11916](https://doi.org/10.1038/srep11916)
34. H. Zhao, S. Uchiyama, *P. Schuck, “Multilaboratory Comparison of Calibration Accuracy and the Performance of External References in Analytical Ultracentrifugation” *PLoS ONE*, **10**(5), e0126420, (2015), [10.1371/journal.pone.0126420](https://doi.org/10.1371/journal.pone.0126420)
 35. S. Totoki, G. Yamamoto, K. Tsumoto, *S. Uchiyama, K. Fukui, “Quantitative Laser Diffraction Method for the Assessment of Protein Subvisible Particles”, *J. Pharm. Sci.*, **104**, 618-626, (2015), [10.1002/jps.24288](https://doi.org/10.1002/jps.24288)
 36. *S. Kita, *H. Matsubara, Y. Kasai, T. Tamaoki, Y. Okabe, H. Fukuwara, J. Kamishikiryo, E. Krayukhina, S. Uchiyama, T. Ose, K. Kuroki, K. Maenaka, “Crystal Structure of Extracellular Domain of Human Lectin-like Transcript 1 (LLT1), the Ligand for Natural Killer Receptor-P1A.” *Eur. J. Immunol.*, **45**(6), 1605-13, (2015), [10.1002/eji.201545509](https://doi.org/10.1002/eji.201545509).
 37. S. Harada, Y. Hiromori, S. Nakamura, K. Kawahara, S. Fukakusa, T. Maruno, M. Noda, S. Uchiyama, K. Fukui, J. Nishikawa, H. Nagase, Y. Kobayashi, *T. Yoshida, T. Ohkubo, *T. Nakanishi, “Structural Basis for PPARgamma Transactivation by Endocrine Disrupting Organotin Compounds.”, *Sci. Rep.*, **5**, 8520, (2015), [10.1038/srep08520](https://doi.org/10.1038/srep08520).
 38. U. Ohto, T. Shibata, H. Tanji, E. Krayukhina, S. Uchiyama, K. Miyake, *T. Shimizu, “Structural Basis of CpG and Inhibitory DNA Recognition by Toll-like receptor 9.” *Nature* **520**, 702-705, (2015), [10.1038/nature14138](https://doi.org/10.1038/nature14138)
 39. E. Krayukhina, K. Tsumoto, *S. Uchiyama, K. Fukui, “Effects of Syringe Material and Silicone Oil Lubrication on the Stability of Pharmaceutical Proteins.” *J. Pharm. Sci.*, **104**, 527-535, (2014), [10.1002/jps.24184](https://doi.org/10.1002/jps.24184)
 40. *S. Uchiyama, “Liquid Formulation for Antibody Drugs.” *BBA - Proteins and Proteomics*, **1844**, 2041-2052, (2014), [10.1016/j.bbapap.2014.07.016](https://doi.org/10.1016/j.bbapap.2014.07.016)
 41. T. Hamano, A. Dwiranti, K. Kaneyoshi, S. Fukuda, R. Kometani, M. Nakao, H. Takata, S. Uchiyama, N. Ohmido, *K. Fukui. “Chromosome Interior Observation by Focused Ion Beam/Scanning Electron Microscopy (FIB/SEM) Using Ionic Liquid Technique” *Microsc. Microanal.*, **20**, 1340-1347, (2014), [10.1017/S143192761401280X](https://doi.org/10.1017/S143192761401280X)
 42. M. Amano, N. Kobayashi, M. Yabuta, S. Uchiyama, *K. Fukui, “Detection of Histidine Oxidation in a Monoclonal Immunoglobulin Gamma (IgG) 1 Antibody.” *Anal. Chem.* **86**, 7536-7543, (2014), [10.1021/ac501300m](https://doi.org/10.1021/ac501300m)
 43. D. Volkin, S. Hershenson, R. Ho, S. Uchiyama, G. Winter, *J. Carpenter, “Two Decades of Publishing Excellence in Pharmaceutical Biotechnology”, *J. Pharm. Sci.*, **104**, 290-300, (2014), [10.1002/jps.24285](https://doi.org/10.1002/jps.24285)
 44. W. Han, M. Yamauchi, U. Hasegawa, M. Noda, K. Fukui, A.J. van der Vlies, S. Uchiyama, *H. Uyama, “Pepsin Immobilization on an Aldehyde-modified Polymethacrylate Monolith and Its Application for Protein Analysis,” *J. Biosci. Bioeng.*, **119**, 505-510, (2014), [10.1016/j.jbiosc.2014.10.018](https://doi.org/10.1016/j.jbiosc.2014.10.018)

【老木成稔】

1. M. Iwamoto, M.A. Elfaramawy, M. Yamatake, T. Matsuura, S. Oiki, “Concurrent *in vitro* Synthesis and Functional Detection of Nascent Activity of the KcsA Channel under the Membrane Potential”, *ACS Synth. Biol.*, **7** (4), 1004-1011, (2018), [10.1021/acssynbio.7b00454](https://doi.org/10.1021/acssynbio.7b00454)
2. S. Kojima, M. Iwamoto, S. Oiki, S. Tochigi, H. Takahashi, “Thylakoid Membranes Contain a Non-selective Channel Permeable to Amall Organic Molecules”, *J. Biol. Chem.*, **293**, 7777-7785, (2018), [10.1074/jbc.RA118.002367](https://doi.org/10.1074/jbc.RA118.002367)
3. S. Oiki, M. Iwamoto, “Channel-Membrane Interplay in Lipid Bilayer Membranes Manipulated through Monolayer Technologies”, *Biological & Pharmaceutical Bulletin*, **41**, 303-311, (2018), [10.1248/bpb.b17-00708](https://doi.org/10.1248/bpb.b17-00708)
4. 炭竈享司, 老木成稔, “チャネルの入口がイオンの透過速度を決める”, *生物物理*, **58**,(1), 012-016, (2018), [10.2142/biophys.58.012](https://doi.org/10.2142/biophys.58.012)
5. M. Kalathingal, T. Sumikama, T. Mori, S. Oiki, S. Saito, ”Structure and Dynamics of Solvent Molecules Inside Polytheonamide B Channel in Different Environments: A Molecular Dynamics Study”, *Phys. Chem. Chem. Phys.*, **20**, 3334-3348, (2017), [10.1039/c7cp06299k](https://doi.org/10.1039/c7cp06299k)
6. M. Iwamoto, A. Sumino, E. Shimada, M. Kinoshita, N. Matsumori, S. Oiki, “Channel Formation and Membrane Deformation via Sterol-Aided Polymorphism of Amphidinol 3”, *Sci. Rep.*, **7**, 10782, (2017), [10.1038/s41598-017-11135-x](https://doi.org/10.1038/s41598-017-11135-x)

7. M. Iwamoto, S. Oiki, "Membrane Perfusion of Hydrophobic Substances Around Channels Embedded in the Contact Bubble bilayer", *Sci. Rep.*, **7**, 6857, (2017), [10.1038/s41598-017-07048-4](https://doi.org/10.1038/s41598-017-07048-4)
 8. 岩本真幸, 老木成稔, "脂質平面膜とパッチクランプのハイブリッドとしての CBB (接触液胞 2 重膜) 法", *生物物理*, **57**(6), 313-317, (2017), [10.2142/biophys.57.313](https://doi.org/10.2142/biophys.57.313)
 9. A. Sumino, T. Uchihashi, *S. Oiki, "Oriented Reconstitution of the Full-length KcsA Potassium Channel in a Lipid Bilayer for AFM Imaging", *J. Phys. Chem. Lett.*, **8**, 785-793, (2017), [10.1021/acs.jpclett.6b03058](https://doi.org/10.1021/acs.jpclett.6b03058)
 10. Y. Matsuki, M. Iwamoto, K. Mita, K. Shigemi, S. Matsunaga, *S. Oiki, "Rectified Proton Grotthuss Conduction Across a Long Water-Wire in the Test Nanotube of the Polytheonamide B Channel", *J. Am. Chem. Soc.*, **138**, (12), 4168-4177, (2016), [10.1021/jacs.5b13377](https://doi.org/10.1021/jacs.5b13377)
 11. T. Sumikama, *S. Oiki, "Digitalized K⁺ Occupancy in the Nanocavity Holds and Releases Queues of K⁺ in a Channel", *J. Am. Chem. Soc.*, **138**, 10284-10292, (2016), [10.1021/jacs.6b05270](https://doi.org/10.1021/jacs.6b05270)
 12. 老木成稔: イオンチャネルの分子構造「特集 電解質の新しい見方・考え方」【水電解質と機能蛋白調節(基礎)】腎と透析, **80**, 390-396, 2016
 13. H.-K. Chang, M. Iwamoto, *S. Oiki, *R.-C Shieh, "Mechanism for Attenuated Outward Conductance Induced by Mutations in the Cytoplasmic Pore of Kir2.1 Channels", *Sci. Rep.*, **5**, 18404 (1-14), (2015), [10.1038/srep18404](https://doi.org/10.1038/srep18404)
 14. *Y. Furutani, H. Shimizu, Y. Asai, S. Oiki, H. Kandori, "Specific Interactions between Alkali Metal Cations and the KcsA Channel Studied Using ATR-FTIR Spectroscopy", *Biophys. Physicobiology*, **12**, 37-45, (2015), [10.2142/biophysico.12.0_37](https://doi.org/10.2142/biophysico.12.0_37)
 15. *S. Oiki, "Channel Function Reconstitution and Re-animation: A Single-channel Strategy in the Post-crystal Age", *J. Physiol.*, **593**, 2553-2573, (2015), [10.1113/JP270025](https://doi.org/10.1113/JP270025)
 16. A. Yamakata, H. Shimizu, *S. Oiki, "Surface-Enhanced IR Absorption Spectroscopy of the KcsA Potassium Channel upon Application of an Electric Field", *Phys. Chem. Chem. Phys.*, **17**, 21104 – 21111, (2015), [10.y1039/C5CP02681D](https://doi.org/10.1039/C5CP02681D)
 17. Y. Furutani, H. Shimizu, Y. Asai, *S. Oiki, H. Kandori, "Specific Interactions between Alkali Metal Cations and the KcsA Channel Studied Using ATR-FTIR Spectroscopy", *Biophysics and Physicobiology*, **12**, 37-45, (2015), [10.2142/biophysico.12.0_37](https://doi.org/10.2142/biophysico.12.0_37)
 18. H. Nakao, K. Ikeda, M. Iwamoto, H. Shimizu, *S. Oiki, Y. Ishihara, M. Nakano, "pH-Dependent Promotion of Phospholipid Flip-Flop by the KcsA Potassium Channel", *BBA Biomemb.*, **1848**(1), Part.A, 145-150, (2015), [10.1016/j.bbamem.2014.10.001](https://doi.org/10.1016/j.bbamem.2014.10.001)
 19. 老木成稔, "膜内 KcsA カリウムチャネルの原子間力顕微鏡による構造・動態解析" 生物物理 **55**(1), 005-010, (2015), [10.2142/biophys.55.005](https://doi.org/10.2142/biophys.55.005)
 20. M. Iwamoto, S. Oiki, "Contact Bubble Bilayers with Flush Drainage", *Sci. Rep.*, **5**, 9110, (2015), [10.1038/srep09110](https://doi.org/10.1038/srep09110)
 21. S. Phongphanphanee, N. Yoshida, *S. Oiki, F. Hirata, "The 'Ambivalent' Snug-Fit Sites in the KcsA Potassium Channel Probed by '3D-RISM Microscopy'", *Pure and Applied Chemistry*, **86**, 97-104, (2014), [10.1515/pac-2014-5018](https://doi.org/10.1515/pac-2014-5018)
 22. M. Iwamoto, S. Matsunaga, *S. Oiki, "Paradoxical One-ion Pore Behavior of a Long β -helical Peptide of Marine Cytotoxic Polytheonamide B", *Sci. Rep.*, **4**, 3636, (2014), [10.1038/srep03636](https://doi.org/10.1038/srep03636)
 23. A. Sumino, D. Yamamoto, M. Iwamoto, T. Dewa, *S. Oiki, "Gating-Associated Clustering-Dispersion Dynamics of the KcsA Potassium Channel in a Lipid Membrane", *J. Phys. Chem. Lett.*, **5**, 578-584, (2014), [10.1021/jz402491t](https://doi.org/10.1021/jz402491t), (ACS Live Slides)
 24. S. Phongphanphanee, N. Yoshida, *S. Oiki, F. Hirata, "Distinct Configurations of Cations and Water in the Selectivity Filter of the KcsA Potassium Channel Proved by 3D-RISM Theory" *J. Mol. Liq.*, **200**, Part.A, 52-58 , (2014), [10.1016/j.molliq.2014.03.050](https://doi.org/10.1016/j.molliq.2014.03.050)
 25. 老木成稔, "KcsA カリウムチャネルでみるチャネル-膜相互作用", 膜 **39**, 309-315, (2014), 日本膜学会, 東京、日本
【岡本祐幸】
1. S. Kawano, Y. Tamura, R. Kojima, S. Bala, E. Asai, A.H. Michel, B. Kornmann, I. Riezman, H. Riezman, Y. Sakae, Y. Okamoto, *T. Endo, "Structure-function Insights into Phospholipid Transfer by Mmm1-Mdm12 of ERMES between Membranes", *J. Cell Biol.*, **217**, 959-974 (2018), [10.1083/jcb.201704119](https://doi.org/10.1083/jcb.201704119)

2. S. Tsukamoto, Y. Sakae, Y. Itoh, T. Suzuki, and *Y. Okamoto, "Computational Analysis for Selectivity of Histone Deacetylase Inhibitor by Replicaexchange Umbrella Sampling Molecular Dynamics Simulations", *J. Chem. Phys.*, **148**, 125102 , (6 pages), (2018), 10.1063/1.5019209
3. S. Ito, D.G. Fedorov, *Y. Okamoto, and *S. Irle, "Implementation of Replica-exchange Umbrella Sampling in GAMESS", *Comput. Phys. Commun.*, **228**, 152-162, (2018), 10.1016/j.cpc.2018.01.014
4. Y. Mori, *Y. Okamoto, "Conformational Changes of Ubiquitin under High Pressure Conditions: A Pressure Simulated Tempering Molecular Dynamics Study", *J. Comput. Chem.*, **38**, 1167-1173, (2017), [10.1002/jcc.24767](#)
5. *Y. Sakae, T. Satoh, H. Yagi, S. Yanaka, T. Yamaguchi, Y. Isoda, S. Iida, Y. Okamoto, *K. Kato, "Conformational Effects of N-glycan Core Fucosylation of Immunoglobulin G Fc Region on its Interaction with Fcγ Receptor IIIa", *Sci. Rep.*, **7**, 13780 (10 pages), (2017), [10.1038/s41598-017-13845-8](#)
6. *N. Nishikawa, Y. Sakae, T. Gouda, Y. Tsujimura, Y. Okamoto, "Two Major Stable Structures of Amyloid-forming Peptides: Amorphous Aggregates and Amyloid fibrils", *Molecular Simulation*, **43**, 1370-1376, (2017), [10.1080/08927022.2017.1359746](#)
7. S. Ito, S. Irle, Y. Okamoto, "Implementation of Replica-exchange Umbrella Sampling in the DFTB+ Semiempirical Quantum Chemistry Package", *Comput. Phys. Commun.*, **204**, 1-10, (2016), [10.1016/j.cpc.2016.02.010](#)
8. G. La Penna, Y. Mori, R. Kitahara, K. Akasaka, Y. Okamoto, "Modeling ¹⁵N NMR Chemical Shift Changes in Protein Backbone with Pressure", *J. Chem. Phys.*, **145**, 085104 (12 pages), (2016), [10.1063/1.4961507](#)
9. 榎慶丈, 西川直宏, 塚本修一朗, 鈴木孝禎, 岡本祐幸, "分子動力学シミュレーションによる医学・創薬に向けたタンパク質の構造解析", *YAKUGAKU ZASSHI*, **136**, 113-120, (2016), [10.1248/yakushi.15-00230-4](#)
10. 岡本祐幸, 卷頭言 : 計算生物物理学の将来, 生物物理 (日本生物物理学会誌) , **56** 卷, 75, (2016), [10.2142/biophys.56.075](#)
11. X. Lu, D. Fang, S. Ito, Y. Okamoto, V. Ovchinnikov, Q. Cui, "QM/MM Free Energy Simulations: Recent Progress and Challenges", *Molecular Simulation*, **42**, 1056-1078, (2016), [10.1080/08927022.2015.1132317](#)
12. R. Urano, Y. Okamoto, "New Implementations of Replica-exchange Method for Simulations of Complex Systems: Designed-walk and Deterministic Replica-exchange Methods", *Phys. Procedia.*, **68**, 100-104, (2015), [10.1016/j.phpro.2015.07.116](#)
13. R. Urano, *H. Kokubo, Y. Okamoto, "Predictions of Tertiary Structures of α-helical Membrane Proteins by Replica-exchange Method with Consideration of Helix Deformations", *J. Phys. Soc. Jpn.*, **84**, 084802 (12 pages), (2015), [10.7566/JPSJ.84.084802](#)
14. *R. Urano, Y. Okamoto, "Designed-walk Replica-exchange Method for Simulations of Complex Systems", *Comput. Phys. Commun.*, 380-383, (2015), [10.1016/j.cpc.2015.07.007](#)
15. *R. Urano, Y. Okamoto, "Deterministic Replica-exchange Method without Pseudo Random Numbers for Simulations of Complex Systems", *Comput. Phys. Commun.*, **197**, 128-135, (2015), [10.1016/j.cpc.2015.08.020](#)
16. R. Urano, *Y. Okamoto "Observation of Helix Associations for Insertion of a Retinal Molecule and Distortions of Helix Structures in Bacteriorhodopsin", *J. Chem. Phys.*, **143**, 235101 (10 pages), (2015), [10.1063/1.4935964](#)
17. Y. Okamoto, "Editorial: ICMS2013", in Special Issue: ICMS2013 *Molecular Simulation* **41**, 779 (2015), [10.1080/08927022.2015.1048075](#)
18. S. Somani, Y. Okamoto, A.J. Ballard, *D.J. Wales, "Equilibrium Molecular Thermodynamics from Kirkwood Sampling", *Journal of Physical Chemistry B*, **119**, 6155-6169, (2015), [10.1021/acs.jpcb.5b01800](#)
19. Y. Sakae, T. Hiroyasu, M. Miki, K. Ishii, *Y. Okamoto, "Conformational Search Simulations of Trp-cage Using Genetic Crossover", *Molecular Simulation* , **41**, 1045-1049, (2015), [1080/08927022.2015.1016937](#)
20. *N. Nishikawa, P.H. Nguyen, P. Derreumaux, Y. Okamoto, "Replica-exchange Molecular Dynamics Simulation for Understanding the Initial Process of Amyloid Peptide Aggregation", *Molecular Simulation* **41**, 1041-1044, (2015), [10.1080/08927022.2014.938445](#)
21. *N. Nishikawa, Y. Sakae, Y. Okamoto, "Molecular Dynamics Simulations to Clarify the Concentration Dependency of Protein Aggregation", *JPS Conference Proceedings*, **5**, 011020, (7 pages), (2015), [10.7566/JPSCP.5.011020](#)

22. Y. Sakae, T. Hiroyasu, M. Miki, K. Ishii, *Y. Okamoto, "A Conformational Search Method for Protein Systems Using Genetic Crossover and Metropolis Criterion", *J. Phys. Conference Series*, **487**, 012003 (5 pages), (2014), 10.1088/1742-6596/487/1/012003
23. *T. Yoda, Y. Sugita, Y. Okamoto, "Salt Effects on Hydrophobic-core Formation in Folding of a Helical Miniprotein Studied by Molecular Dynamics Simulations", *PROTEINS: Structure, Function, and Bioinformatics*, **82**, 933-943, (2014), 10.1002/prot.24467
24. *Y. Okamoto, H. Kokubo, T. Tanaka, "Prediction of Ligand Binding Affinity by the Combination of Replica-exchange Method and Double-decoupling Method", *Journal of Chemical Theory and Computation* **10**, 3563-3569, (2014), 10.1021/ct500539u
25. T. Yamaguchi, Y. Sakae, Y. Zhang, S. Yamamoto, Y. Okamoto, *K. Kato, "Exploration of Conformational Spaces of High-mannose-type Oligosaccharides by an NMR-validated Simulation", *Angewandte Chemie International Edition* **53**, 10941-10944, (2014), 10.1002/anie.201406145
26. *H. Kokubo, T. Tanaka, Y. Okamoto, "Prediction of Protein-ligand Binding Structures by Replica-exchange Umbrella Sampling Simulations: Application to Kinase Systems", *Journal of Chemical Theory and Computation* **9**, 4660-4671, (2013), 10.1021/ct4004383
27. *H. Kokubo, T. Tanaka, Y. Okamoto, "Two-dimensional Replica-exchange Method for Predicting Protein-ligand Binding Structures", *Journal of Computational Chemistry*, **34**, 2601-2614, (2013), 10.1002/jcc.23427

【奥村久士】

1. Y. Mori, *H. Okumura, T. Watanabe, T. Hohsaka, "Antigen-dependent Fluorescence Response of Anti-c-Myc Quenched Body Studied by Molecular Dynamics Simulations", *Chem. Phys. Lett.*, **698**, 223, (2018), 10.1016/j.cplett.2018.03.01
2. H. Nishizawa, *H. Okumura, "Classical Molecular Dynamics Simulation to Understand Role of a Zinc Ion for Aggregation of Amyloid- β Peptides", *J. Comput. Chem. Jpn.*, **17**, 76, (2018), 10.2477/jccj.2018-0005
3. *H. Okumura, M. Higashi, Y. Yoshida, H. Sato, R. Akiyama, "Theoretical Approaches for Dynamical Ordering of Biomolecular Systems", *Biochim. Biophys. Acta, (BBA), Gen. Subj.*, **1862**, 212-228, (2018), 10.1016/j.bbagen.2017.10.001
4. *M. Yamauchi, *H. Okumura, "Replica-permutation Method for Molecular Dynamics and Monte Carlo Simulations and its Application to Reveal Temperature and Pressure Dependence of Folded, Misfolded, and Unfolded States of Chignolin", *J. Chem. Phys.*, **147**, 184107, (15 pages), (2017), 10.1063/1.4996431
5. *H. Okumura, S. G. Itoh, "Structural and Fluctuational Difference between Two Ends of A β Amyloid Fibril: MD Simulation Predicts Only One End has Open Conformations", *Sci. Rep.*, **6**, 38422, (2016), 10.1038/srep38422
6. R. Gupta, *S. Saito, Y. Mori, S. G. Itoh, H. Okumura, *M. Tominaga, "Structural Basis of TRPA1 Inhibition by HC-030031 Utilizing Species-specific Differences", *Sci. Rep.*, **6**, 37460, (2016), 10.1038/srep37460
7. H. Nishizawa, *H. Okumura, "Rapid QM/MM Approach for Biomolecular Systems Under Periodic Boundary Conditions: Combination of the Density-functional Tight-binding Theory and Particle Mesh Ewald Method", *J. Comput. Chem.*, **37**, 2701-2711, (2016), 10.1002/jcc.24497
8. S. G. Itoh, *H. Okumura, "Oligomer Formation of Amyloid- β (29-42) from its Monomers Using the Hamiltonian Replica-permutation Molecular Dynamics Simulation", *J. Phys. Chem. B*, **120**, 6555-6561, (2016), 10.1021/acs.jpcb.6b03828
9. W. Khuntawee, T. Rungrotmongkol, P. Wolschann, P. Pongsawasdi, N. Kungwan, *H. Okumura, *S. Hannongbua, "Conformation Study of ϵ -cyclodextrin: Replica-exchange Molecular Dynamics Simulations", *Carbohydr. Polym.*, **141**, 99-105, (2016), 10.1016/j.carbpol.2015.10.018
10. *伊藤暁, *奥村久士:「レア・イベントを捕えるための新たな分子シミュレーション手法—アミロイド線維形成の理解に向けた取り組みー」日本物理学会誌 **71**(7), 463-468, (2016), 10.11316/butsuri.71.7_463
11. Y. Mori, *H. Okumura, "Simulated Tempering Based on Global Balance or Detailed Balance Conditions: Suwa-Todo, Heat bath, and Metropolis algorithms", *J. Comput. Chem.*, **36**, (2015), 2344-2349, 10.1002/jcc.24213
12. H. Nishizawa, *H. Okumura, "Comparison of Replica-permutation Molecular Dynamics Simulations with and without Detailed Balance Condition" *J. Phys. Soc. Jpn.*, **84**, 074801 (6 pages), (2015), 10.7566/JPSJ.84.074801

13. S. G. Itoh, *H. Okumura, "Replica-permutation Method to Enhance Sampling Efficiency" *Mol. Sim.*, **41**, 1021-1026, (2015), [10.1080/08927022.2014.923576](https://doi.org/10.1080/08927022.2014.923576)
14. Y. Mori, *H. Okumura, "Molecular Dynamics Simulation Study on the High-pressure Behaviour of an AK16 Peptide" *Mol. Sim.*, **41**, 1035-1040, (2015), [10.1080/08927022.2014.938071](https://doi.org/10.1080/08927022.2014.938071)
15. K. Inagaki, T. Satoh, S. G. Itoh, H. Okumura, *K. Kato, "Redox-dependent Conformational Transition of Catalytic Domain of Protein Disulfide Isomerase Indicated by Crystal Structure-based Molecular Dynamics Simulation", *Chem. Phys. Lett.*, **618**, 203-207, (2015), [10.1016/j.cplett.2014.11.017](https://doi.org/10.1016/j.cplett.2014.11.017)
16. S. G. Itoh, *H. Okumura, "Dimerization Process of Amyloid- β (29-42) Studied by the Hamiltonian Replica-permutation Molecular Dynamics Simulations", *J. Phys. Chem. B*, **118**, 11428-11436, (2014), [10.1021/jp505984e](https://doi.org/10.1021/jp505984e)
17. *H. Okumura, S. G. Itoh, "Amyloid Fibril Disruption by Ultrasonic Cavitation: Nonequilibrium Molecular Dynamics Simulations", *J. Am. Chem. Soc.*, **136**(30), 10549-10552, (2014), [10.1021/ja502749f](https://doi.org/10.1021/ja502749f)
18. Y. Mori, *H. Okumura, "Molecular Dynamics Study on the Structural Changes of Helical Peptides Induced by Pressure", *Proteins*, **82**, 2970-2981, (2014), [10.1002/prot.24654](https://doi.org/10.1002/prot.24654)
19. H.-L. Chiang, C.-J. Chen, H. Okumura, *C.-K. Hu, "Transformation between α -helix and β -sheet Structures of One and Two Polyglutamine Peptides in Explicit Water Molecules by Replica-exchange Molecular Dynamics Simulations", *J. Comput. Chem.*, **35**, 1430-1437, (2014), [10.1002/jcc.23633](https://doi.org/10.1002/jcc.23633)
20. *奥村久士: “タンパク質の折りたたみ、変性、凝集、アミロイド線維：生体分子動力学シミュレーションの最前線”, *分子研レターズ*, **70**, 4-7, (2014)

【加藤晃一】

1. K. Morita, Y.Y. Yamamoto, A. Hori, T. Obata, Y. Uno, K. Shinohara, K. Noguchi, K. Noi, T. Ogura, K. Ishii, K. Kato, M. Kikumoto, R. Arranz, J.M. Valpuesta, M. Yohda, "Expression, Functional Characterization, and Preliminary Crystallization of the Cochaperone Prefoldin from the Thermophilic Fungus Chaetomium Thermophilum", *Int. J. Mol. Sci.*, **19**, 2452, (2018), [10.3390/ijms19082452](https://doi.org/10.3390/ijms19082452)
2. Y. Hisamatsu, N. Umezawa, H. Yagi, K. Kato, T. Higuchi, "Design and Synthesis of a 4-Aminoquinoline-based Molecular Tweezer that Recognizes Protoporphyrin IX and Iron(III) Protoporphyrin IX and its Application as a Supramolecular Photosensitizer", *Chem. Sci.*, in press (2018), [10.1039/C8SC02133C](https://doi.org/10.1039/C8SC02133C)
3. N. Sriwilaijaroen, S. Nakakita, S. Kondo, H. Yagi, K. Kato, T. Murata, H. Hiramatsu, T. Kawahara, Y. Watanabe, Y. Kanai, T. Ono, J. Hirabayashi, K. Matsumoto, Y. Suzuki, "N-glycan Structures of Human Alveoli Provide Insight into Influenza A virus Infection and Pathogenesis", *FEBS J.*, **285**, 1611-1634, (2018), [10.1111/febs.14431](https://doi.org/10.1111/febs.14431)
4. K. Kato, T. Satoh, "Structural Insights on the Dynamics of Proteasome Formation", *Biophys. Rev.*, **10**, 597-604, (2018), [10.1007/s12551-017-0381-4](https://doi.org/10.1007/s12551-017-0381-4)
5. 山口拓実, 渡邊東紀男, 矢木宏和, 加藤晃一, "分子動力学計算とNMR計測を用いた糖鎖の配座空間探査", *J. Comput. Chem. Jpn.*, **17**, 1-7 (2018), [10.2477/jcc.2018-0011](https://doi.org/10.2477/jcc.2018-0011)
6. 矢木宏和, 加藤晃一, "常磁性NMR法と計算科学を組み合わせた糖鎖の動的コンホメーション解析", *生化学*, **90**, 198-202, (2018), [10.14952/SEIKAGAKU.2018.900198](https://doi.org/10.14952/SEIKAGAKU.2018.900198)
7. T. Yamaguchi, K. Kato, "Molecular Dynamics of Gangliosides", *Methods Mol Biol.*, Humana Press (New York), Gangliosides (S.Sonnino and A.Prinetti ed.), **1804**, 411-417 (2018), [10.1007/978-1-4939-8552-4_20](https://doi.org/10.1007/978-1-4939-8552-4_20)
8. K. Kato, H. Yagi, T. Yamaguchi, "NMR Characterization of the Dynamic Conformations of Oligosaccharides", *Modern Magnetic Resonance, 2nd Edition* (G.A.Webb ed.), 737-754, Springer International Publishing, (2018), [10.1007/978-3-319-28275-6_35-1](https://doi.org/10.1007/978-3-319-28275-6_35-1)
9. H. Yagi, S. Yanaka, K. Kato, "Structure and Dynamics of Immunoglobulin G Glycoproteins Glycobiophysics", *Advances in Experimental, Medicine and Biology* 1104, Springer Nature Singapore Pte Ltd. in press (2018)
10. T. Satoh, K. Kato, "Structural Aspects of ER Glycoprotein Quality-Control System Mediated by Glucose Tagging Glycobiophysics", *Advances in Experimental, Medicine and Biology* 1104, Springer Nature Singapore Pte Ltd. in press (2018)
11. H. Yagi, G. Yan, T. Suzuki, S. Tsuge, T. Yamaguchi, *K. Kato, "Lewis X-carrying Neoglycolipids Evoke Selective Apoptosis in Neural Stem Cells", *Neurochem. Res.*, **43**, 212-218, (2018), [10.1007/s11064-017-2415-5](https://doi.org/10.1007/s11064-017-2415-5)
12. M. Yagi-Utsumi, A. Sikdar, T. Kozai, R. Inoue, M. Sugiyama, T. Uchihashi, H. Yagi, T. Satoh, *K. Kato,

- “Conversion of Functionally Undefined Homopentameric Protein PbaA into a Proteasome Activator by Mutational Modification of its C-terminal Segment Conformation”, *Protein Eng. Des. Sel.*, **31**, 29-36,(2018),10.1093/protein/gzx066
13. J. Kicuntod, K. Sangpheak, M. Mueller, P. Wolschann, H. Viernstein, S. Yanaka, *K. Kato, W. Chavasiri, P. Pongsawasdi, N. Kungwan, T. Rungrotmongkol, “Theoretical and Experimental Studies on Inclusion Complexes of Pinostrobin and β -Cyclodextrins”, *Sci. Pharm.*, **86**, 5, (2018),10.3390/scipharm86010005
 14. H. Yagi, D. Takakura, L. T. Roumenina, W. H. Fridman, C. Sautès-Fridman, N. Kawasaki, *K. Kato, “Site-specific N-glycosylation Analysis of Soluble Fc γ Receptor IIIb in Human Serum, *Sci. Rep.*, **8**, Article number: 2719, (2018),10.1038/s41598-018-21145-y
 15. K. Kato, T. Furuhashi, K. Kato, A. Oda, *E. Kurimoto, “The Assembly Mechanism of Coiled-coil Domains of the Yeast Cargo Receptors Emp46p/47p and the Mutational Alteration of pH-dependency of Complex Formation”, *J Biochem.*, **163**, 441-446, (2018), [10.1093/jb/mvy011](https://doi.org/10.1093/jb/mvy011)
 16. R. Yogo, S. Yanaka, *K. Kato, “Backbone ^1H , ^{13}C , and ^{15}N Assignments of the Extracellular Region of Human Fc γ Receptor IIIb”, *Biomol. NMR Assign.*, **12**, 201-204, (2018), [10.1007/s12104-018-9809-4](https://doi.org/10.1007/s12104-018-9809-4)
 17. K. Mukaigasa, T. Tsujita, V. Thanh Nguyen, L. Li, H. Yagi, Y. Fuse, Y. Nakajima-Takagi, K. Kato, M. Yamamoto, *M. Kobayashi, “Nrf2 Activation Attenuates Genetic Endoplasmic Reticulum Stress Induced by a Mutation in the Phosphomannomutase 2 Gene in Zebrafish”, *Proc. Natl. Acad. Sci. USA*, **115**, 2758-2763, (2018), 10.1073/pnas.1714056115
 18. S. Yanaka, H. Yagi, R. Yogo, M. Yagi-Utsumi, *K. Kato, “Stable Isotope Labeling Approaches for NMR Characterization of Glycoproteins Using Eukaryotic Expression Systems”, *J. Biomol. NMR*, in press, (2018), [10.1007/s10858-018-0169-2](https://doi.org/10.1007/s10858-018-0169-2)
 19. T. Ikeya, D. Ban, D. Lee, Y. Ito, K. Kato, *Christian Griesinger, “Solution NMR Views of Dynamical Ordering of Biomacromolecules”, *Biochim. Biophys. Acta –General Subjects*, **1862**, 287-306 (2018), 10.1016/j.bbagen.2017.08.020
 20. K. Kurihara, M. Matsuo, *T. Yamaguchi, *S. Sato, ‘Synthetic Approach to biomolecular science by cyborg supramolecular chemistry,’ *Biochimica et Biophysica Acta (BBA) - General Subjects*, **1862**, 358, (2018), 10.1016/j.bbagen.2017.11.002
 21. K. Matsuzaki, K. Kato, *K. Yanagisawa, “Ganglioside-mediated Assembly of Amyloid β -protein: Roles in Alzheimer's Disease”, *Prog. Mol. Biol. Transl. Sci.*, **156**, 413-434, (2018), [10.1016/bs.pmbts.2017.10.005](https://doi.org/10.1016/bs.pmbts.2017.10.005)
 22. T. Suzuki, M. Kajino, S. Yanaka, T. Zhu, H. Yagi, T. Satoh, T. Yamaguchi, *K. Kato, “Conformational Analysis of a High-mannose-type Oligosaccharide Displaying Glucosyl Determinant Recognised by Molecular Chaperones Using NMR-validated Molecular Dynamics Simulation”, *ChemBioChem*, **18**, 396-410, (2017), 10.1002/cbic.201600595
 23. T. Yoshimura, A. Hayashi, M. Handa-Narumi, H. Yagi, N. Ohno, T. Koike, Y. Yamaguchi, K. Uchiyama, K. Kadomatsu, J. Sedzik, K. Kitamura, K. Kato, B.D. Trapp, H. Baba, *K. Ikenaka, “GlcNAc6ST-1 Regulates Sulfation of N-glycans and Myelination in the Peripheral Nervous System”, *Sci. Rep.*, **7**, Article number: 42257, (2017), 10.1038/srep42257
 24. M. Nagae, S. K. M., M. Neyazaki, R. Oi, A. Ikeda, N. Matsugaki, S. Akashi, H. Manya, M. Mizuno, H. Yagi, K. Kato, T. Senda, T. Endo, T. Nogi, *Y. Yamaguchi, “3D structural Analysis of Protein O-mannosyl Kinase, POMK, a Causative Gene Product of Dystroglycanopathy”, *Genes Cells*, **22**, 348-359, (2017), 10.1111/gtc.12480
 25. S. Sawaguchi, S. Varshney, M. Ogawa, Y. Sakaidani, H. Yagi, K. Takeshita, T. Murohara, K. Kato, S. Sundaram, P. Stanley, *T. Okajima, “GlcNAc on NOTCH1 EGF Repeats Regulates Ligand-induced Notch Signaling and Vascular Development in Mammals”, *eLife*, **6**, e24419, (2017), 10.7554/eLife.24419
 26. E. Kurimoto, T. Satoh, Y. Ito, E. Ishihara, K. Okamoto, M. Yagi-Utsumi, K. Tanaka, *K. Kato, “Crystal Structure of Human Proteasome Assembly Chaperone PAC4 Involved in Proteasome Formation”, *Protein Sci.*, **26**, 1080-1085, (2017), 10.1002/pro.3153
 27. T. Kato, N. Kako, K. Kikuta, T. Miyazaki, S. Kondo, H. Yagi, K. Kato, *E. Y. Park, “N-Glycan Modification of a Recombinant Protein via Coexpression of Human Glycosyltransferases in Silkworm Pupae”, *Sci. Rep.*, **7**, Article number: 1409, (2017),10.1038/s41598-017-01630-6
 28. G. Yan, T. Yamaguchi, T. Suzuki, S. Yanaka, S. Sato, M. Fujita, *K. Kato, “Hyper-assembly of Self-Assembled Glycoclusters Mediated by Specific Carbohydrate–carbohydrate Interactions”, *Chem. Asian J.*

- J.*, **12**, 968-972, (2017), 10.1002/asia.201700202
29. T. Kato, K. Kikuta, A. Kanematsu, S. Kondo, H. Yagi, K. Kato, *E. Y. Park, "Alteration of a Recombinant Protein N-glycan Structure in Silkworms by Partial Suppression of N-acetylglucosaminidase Gene Expression", *Biotechnol. Lett.*, **39**, 1299-1308, (2017), 10.1007/s10529-017-2361-y
30. H. Yagi, H. Tateno, K. ayashi, T. Hayashi, K. Takahashi, J. Hirabayashi, K. Kato, *M. Tsuboi, "Lectin Microarray Analysis of Isolated Polysaccharides from *Sasa Veitchii*", *Biosci. Biotechnol. Biochem.*, **81**, 1687-1689, (2017), 10.1080/09168451.2017.1340089
31. R. Yogo, S. Yanaka, H. Yagi, A. Martel, L. Porcar, Y. Ueki, R. Inoue, N. Sato, M. Sugiyama, *K. Kato, "Characterization of Conformational Deformation-coupled Interaction between Immunoglobulin G1 Fc Glycoprotein and a Low-affinity Fcγ Receptor by Deuteration-assisted Small-angle Neutron Scattering", *Biochem. Biophys. Rep.*, **12**, 1-4, (2017), 10.1016/j.bbrep.2017.08.004
32. T. Satoh, C. Song, T. Zhu, T. Toshimori, K. Murata, Y. Hayashi, H. Kamikubo, T. Uchihashi, *K. Kato, "Visualisation of a flexible modular structure of the ER folding-sensor enzyme UGGT", *Sci. Rep.*, **7**, Article number: 12142, (2017), 10.1038/s41598-017-12283-w
33. S. Kitazawa, M. Yagi-Utsumi, K. Kato, *R. Kitahara, "Interactions Controlling the Slow Dynamic Conformational Motions of Ubiquitin", *Molecules*, **22**, 1414, (2017), 10.3390/molecules22091414
34. S. Yanaka, T. Yamazaki, R. Yogo, M. Noda, S. Uchiyama, H. Yagi, *K. Kato, "NMR Detection of Semi-specific Antibody Interactions in Serum Environments", *Molecules*, **22**, 1619, (2017), 10.3390/molecules22101619
35. Y. Sakae, T. Satoh, H. Yagi, S. Yanaka, T. Yamaguchi, Y. Isoda, S. Iida, Y. Okamoto, *K. Kato, "Conformational Effects of N-glycan Core fucosylation of Immunoglobulin G Fc Region on its Interaction with Fcγ Receptor IIIa", *Sci. Rep.*, **7**, Article number: 13780, (2017), 10.1038/s41598-017-13845-8
36. T. Takenaka, T. Nakamura, S. Yanaka, M. Yagi-Utsumi, M. S. Chandak, K. Takahashi, S. Paul, K. Makabe, M. Arai, K. Kato, *K. Kuwajima, "Formation of the Chaperonin Complex Studied by 2D NMR Spectroscopy", *PLOS ONE*, **12**, e0187022, (2017), 10.1371/journal.pone.0187022
37. T. Kozai, T. Sekiguchi, T. Satoh, H. Yagi, K. Kato, *T. Uchihashi, "Two-step Process for Disassembly Mechanism of Proteasome α7 Homo-tetradecamer by α6 Revealed by High-speed Atomic Force Microscopy", *Sci. Rep.*, **7**, Article number: 15373, (2017), 10.1038/s41598-017-15708-8
38. 矢木宏和, *加藤晃一, NMR を利用して糖タンパク質糖鎖の構造動態と相互作用を観る
医学のあゆみ, **262**, 467-, (2017)
39. T. Suzuki, M. Kajino, S. Yanaka, T. Zhu, H. Yagi, T. Satoh, T. Yamaguchi, *K. Kato, "Conformational Analysis of a High-mannose-type Oligosaccharide Displaying Glucosyl Determinant Recognised by Molecular Chaperones Using NMR-validated Molecular Dynamics Simulation," *ChemBioChem.*, **18**, 396-410, (2017), 10.1002/cbic.201600595
40. T. Yoshimura, A. Hayashi, M. Handa-Narumi, H. Yagi, N. Ohno, T. Koike, Y. Yamaguchi, K. Uchimura, K. Kadomatsu, J. Sedzik, K. Kitamura, K. Kato, B.D. Trapp, H. Baba, *K. Ikenaka, "GlcNAc6ST-1 Regulates Sulfation of N-glycans and Myelination in the Peripheral Nervous System," *Sci. Rep.*, **7**, Article number: 42257, (2017), 10.1038/srep42257
41. M. Nagae, S.K. Mishra, M. Neyazaki, R. Oi, A. Ikeda, N. Matsugaki, S. Akashi, H. Manya, M. Mizuno, H. Yagi, K. Kato, T. Senda, T. Endo, T. Nogi, *Y. Yamaguchi, "3D Structural Analysis of Protein O-Mannosyl Kinase, POMK, a Causative Gene Product of Dystroglycanopathy," *Genes Cells.*, **22**(4), 348-359, (2017), 10.1111/gtc.12480
42. E. Kurimoro, T. Satoh, Y. Ito, E. Ishihara, K. Okamoto, M. Yagi-Utsumi, K. Tanaka, *K. Kato, "Crystal Structure of Human Proteasome Assembly Chaperone PAC4 Involved in Proteasome Formation," *Protein Sci.*, **26**(5), 1080-1085, (2017), 10.1002/pro.3153
43. S. Sawaguchi, S. Varshney, M. Ogawa, Y. Sakaidani, H. Yagi, K. Takeshita, T. Murohara, K. Kato, S. Sundaram, P. Stanley, *T. Okajima, "O-GlcNAc on NOTCH1 EGF Repeats Regulates Ligand-Induced Notch Signaling and Vascular Development in Mammals," *eLife*, in press, (2017), 10.7554/eLife.24419,
44. G. Yan, T. Yamaguchi, T. Suzuki, S. Yanaka, S. Sato, M. Fujita, *K. Kato, "Hyper-Assembly of Self-assembled Glycoclusters Mediated by Specific Carbohydrate-carbohydrate Interactions," *Chem. Asian J.*, **12**, 968-972, (2017), 10.1002/asia.201700202

45. H. Yagi, K. Kato, "Functional Roles of Glycoconjugates in the Maintenance of Stemness and Differentiation Process of Neural Stem Cells," *Glycoconjugate J.*, **34**, 757-763, (2017)
46. R.Inoue, T.Takata, N.Fujii, K.Ishii, S.Uchiyama, N.Sato, Y.Oba, K.Wood, K.Kato, N.Fujii, *M.Sugiyama, "New Insight into the Dynamical System of α B-crystallin Oligomers," *Sci. Rep.*, **6**, Article number: 29208, (2016), [10.1038/srep29208](https://doi.org/10.1038/srep29208)
47. H. Ito, H. Kaji, A. Togayachi, P. Azadi, M. Ishihara, R. Geyer, C. Galuska, H. Geyer, K. Kakehi, M. Kinoshita, N.G. Karlsson, C. Jin, K. Kato, H. Yagi, S. Kondo, N. Kawasaki, N. Hashii, D. Kolarich, K. Stavenhagen, N.H. Packer, M. Nakano, N. Taniguchi, A. Kurimoto, Y. Wada, M. Tajiri, P. Yang, W. Cao, H.Li, P.M.Rudd, *H. Narimatsu, "Comparison of Analytical Methods for Profiling *N*- and *O*-Linked Glycans from Cultured Cell Lines: HUPO Human Disease Glycomics/Proteome Initiative Multi-institutional Study," *Glycoconjugate J.*, **33**, 405-415 (2016), [10.1007/s10719-015-9625-3](https://doi.org/10.1007/s10719-015-9625-3)
48. M.Sugiyama, H. Yagi, K. Ishii, L. Porcare, A. Martele, K. Oyama, M. Noda, Y. Yunoki, R. Murakami, R. Inoue, N. Sato, Y. Oba, K. Terauchi, S. Uchiyama, *K. Kato, "Structural Characterization of the Circadian Clock Protein Complex Composed of KaiB and KaiC by Inverse Contrast-matching Small-angle Neutron Scattering," *Sci. Rep.*, **6**, Article number: 35567, (2016), [10.1038/srep35567](https://doi.org/10.1038/srep35567)
49. T. Satoh, T. Toshimori, M. Noda, S. Uchiyama, *K. Kato, "Interaction Mode between Catalytic and Regulatory Subunits in Glucosidase II Involved in ER Glycoprotein Quality Control," *Protein Sci.*, **25**, 2095-2101, (2016), [10.1002/pro.3031](https://doi.org/10.1002/pro.3031)
50. H. Yagi, C.-W. Kuo, T. Obayashi, S. Ninagawa, K.-H.Khoo, *K. Kato, "Direct Mapping of Additional Modifications on Phosphorylated *O*-glycans of α -dystroglycan by Mass Spectrometry Analysis in Conjunction with Knocking out of Causative Genes for Dystroglycanopathy," *Mol. Cell Proteomics*, **15**, 3424-3434, (2016), [10.1074/mcp.M116.062729](https://doi.org/10.1074/mcp.M116.062729)
51. M.Yagi-Utsumi, K. Kato, *K. Nishimura, "Membrane-induced Dichotomous Conformation of Amyloid B with the Disordered N-terminal Segment Followed by the Stable C-terminal β Structure", *PLoS ONE*, **11**, e0146405, (2016), [10.1371/journal.pone.0146405](https://doi.org/10.1371/journal.pone.0146405)
52. S. Seetaha, M.Yagi-Utsumi, T. Yamaguchi, K. Ishii, S. Hannongbua, *K. Choowongkomon, *K. Kato, "Application of Site-specific Spin Labeling for NMR Detecting Inhibitor-induced Conformational Change of HIV-1 Reverse Transcriptase", *Chem.Med.Chem.*, **11**, 363-366, (2016), [10.1002/cmdc.201500554](https://doi.org/10.1002/cmdc.201500554)
53. *T. Satoh, T. Toshimori, G. Yan, T. Yamaguchi, *K. Kato, "Structural Basis for Two-step Glucose Trimming by Glucosidase II Involved in ER Glycoprotein Quality Control", *Sci. Rep.*, **6**, Article number: 20575, (2016), [10.1038/srep20575](https://doi.org/10.1038/srep20575)
54. R. Thammaporn, K. Ishii, M. Yagi-Utsumi, S. Uchiyama, *S. Hannongbua, *K. Kato, "Mass Spectrometric Characterization of HIV-1 Reverse Transcriptase Interactions with Non-Nucleoside Reverse Transcriptase Inhibitors", *Biol. Pharm. Bull.*, **39**, 450-454, (2016), [10.1248/bpb.b15-00880](https://doi.org/10.1248/bpb.b15-00880)
55. 加藤晃一, 谷中冴子, 矢木・内海真穂, "NMR構造生物学がもたらす新たな創薬研究のツール", *MEDCHEM NEWS*, **26**, 195-200, (2016)
56. M.Yagi-Utsumi, T. Satoh, *K. Kato, "Structural Basis of Redox-dependent Substrate Binding of Protein Disulfide Isomerase", *Sci. Rep.*, **5**, Article number: 13909, (2015), [10.1038/srep13909](https://doi.org/10.1038/srep13909)
57. K. Inagaki, T. Satoh, M.Yagi-Utsumi, A.-C. Le Gulluche, T. Anzai, Y. Uekusa, Y. Kamiya, *K. Kato, "Redox-coupled Structural Changes of the Catalytic a Domain of Protein Disulfide Isomerase", *FEBS Lett.*, **589**, 2690-2694, (2015), [10.1016/j.febslet.2015.07.041](https://doi.org/10.1016/j.febslet.2015.07.041)
58. Y. Isoda, H. Yagi, T. Satoh, M. Shibata-Koyama, K. Masuda, M. Satoh, K. Kato, *S. Iida, "Importance of the Side Chain at Position 296 of Antibody Fc in Interactions with Fcy RIIIa and Other Fcy Receptors", *PLoS ONE*, **10**, e0140120, (2015), [10.1371/journal.pone.0140120](https://doi.org/10.1371/journal.pone.0140120)
59. K. Ishii, H. Enda, M. Noda, M. Kajino, A. Kim, E. Kurimoto, K. Sato, A. Nakano, Y. Kobayashi, H. Yagi, S. Uchiyama, *K. Kato, "pH-dependent Assembly and Segregation of the Coiled-coil Segments of Yeast Putative Cargo Receptors Emp46p and Emp47p", *PLoS ONE*, **10**, e0140287, (2015), [10.1371/journal.pone.0140287](https://doi.org/10.1371/journal.pone.0140287)
60. R. Thammaporn, M.Yagi-Utsumi, T. Yamaguchi, P. Boonsri, P. Saparpakorn, K. Choowongkomon, S. Techasakul, *K. Kato, *S. Hannongbua, "NMR Characterization of HIV-1 Reverse Transcriptase Binding to Various Non-nucleoside Reverse Transcriptase Inhibitors with Different Activities", *Sci. Rep.*, **5**, Article number: 15806, (2015), [10.1038/srep15806](https://doi.org/10.1038/srep15806)
61. T. Zhu, T. Yamaguchi, T. Satoh, *K. Kato, "A Hybrid Strategy for the Preparation of ^{13}C -labeled High-

- mannose-type Oligosaccharides with Terminal Glucosylation for NMR Study”, *Chem. Lett.*, **44**, 1744-1746, (2015), 10.1246/cl.150898
62. H. Yagi, Y. Zhang, M. Yagi-Utsumi, T. Yamaguchi, S. Iida, Y. Yamaguchi, *K. Kato, “Backbone ^1H , ^{13}C , and ^{15}N Resonance Assignments of the Fc Fragment of Human Immunoglobulin G Glycoprotein”, *Biomol. NMR Assign.*, **9**, 257-260, (2015), 10.1007/s12104-014-9586-7
63. H. Yagi, N. Fukuzawa, Y. Tasaka, K. Matsuo, Y. Zhang, T. Yamaguchi, S. Kondo, S. Nakazawa, N. Hashii, N. Kawasaki, T. Matsumura, *K. Kato, “NMR-based Structural Validation of Therapeutic Antibody Produced in *Nicotiana benthamiana*”, *Plant Cell Rep.*, **34**, 959-968, (2015), 10.1007/s00299-015-1757-1
64. H. Yagi, M. Nakamura, J. Yokoyama, Y. Zhang, T. Yamaguchi, S. Kondo, J. Kobayashi, T. Kato, E. Y. Park, S. Nakazawa, N. Hashii, N. Kawasaki, *K. Kato, “Stable Isotope Labeling of Glycoprotein Expressed in Silkworms Using Immunoglobulin G as a Test Molecule”, *J. Biomol. NMR*, **62**, 157-167, (2015), 10.1007/s10858-015-9930-y
65. 加藤晃一, *稻垣直之, “離合集散が織りなす生命分子機能の研究フロンティア”, *実験医学*, 5月号, **33**, 8, (2015)
66. M. Yagi-Utsumi, *K. Kato, “Structural and Dynamic Views of GM1 Ganglioside”, *Glycoconjugate J.*, **32**, 105-112, (2015), 10.1007/s10719-015-9587-5
67. M. Ogawa, S. Sawaguchi, T. Kawai, D. Nadano, T. Matsuda, H. Yagi, K. Kato, K. Furukawa, *T. Okajima, “Impaired O-linked N-acetylglucosaminylation in the Endoplasmic Reticulum by Mutated EGF Domain-specific O-linked N-acetylglucosamine Transferase Found in Adams-Oliver Syndrome”, *J. Biol. Chem.*, **290**, 2137-2149, (2015), 10.1074/jbc
68. K. Inagaki, T. Satoh, S. G. Itoh, H. Okumura, *K. Kato, “Redox-dependent Conformational Transition of Catalytic Domain of Protein Disulfide Isomerase Indicated by Crystal Structure-based Molecular Dynamics Simulation”, *Chem. Phys. Lett.*, **618**, 203-207, (2015), 10.1016/j.cplett.2014.11.017
69. Y. Kitago, M. Nagae, Z. Nakata, M. Yagi-Utsumi, S. Takagi-Niidome, E. Mihara, T. Nogi, K. Kato, *J. Takagi, “Structural Basis for Amyloidogenic Peptide Recognition by SorLA”, *Nat. Struct. Mol. Biol.*, **22**, 199-206, (2015), 10.1038/nsmb.2954
70. *S. Sato, R. Takeuchi, M. Yagi-Utsumi, T. Yamaguchi, Y. Yamaguchi, K. Kato, *M. Fujita, “Self-Assembled, π -Stacked Complex as a Finely-Tunable Magnetic Aligner for Biomolecular NMR Applications”, *Chem. Comm.*, **51**, 2540-2543, (2015), 10.1039/C4CC09354B
71. *T. Satoh, T. Yamaguchi, *K. Kato, “Emerging Structural Insights into Glycoprotein Quality Control Coupled with N -glycan Processing in the Endoplasmic Reticulum”, *Molecules*, **20**, 2475-2491, (2015), 10.3390/molecules20022475
72. 山口拓実, *加藤晃一, “糖鎖の立体構造を描き出す”, *生物物理*, **55**, 81-83, (2015), 10.2142/biophys.55.081
73. *加藤晃一, “糖鎖構造学研究の新展開”, *BIOTOVO*, **23**, 2-3 (2015)
74. T. Yamaguchi, *K. Kato, “Paramagnetic NMR probes for characterization of the dynamic conformations and interactions of oligosaccharides”, *Glycoconjugate J.*, **32**, 505-513, (2015), 10.1007/s10719-015-9599-1
75. N. Nakagawaa, H. Yagi, K. Kato, H. Takematsu, *S. Oka, “Ectopic Clustering of Cajal-Retzius and Subplate Cells is an Initial Pathological Feature in Pompant knockout Mice, a Model of Dystroglycanopathy”, *Sci. Rep.*, **5**, 11163, (2015), 10.1038/srep11163
76. S. H. Kang, H. S. Jung, S. J. Lee, C. I. Park, S. M. Lim, H. Park, B. S. Kim, K. H. Na, G. J. Han, J. W. Bae, H. J. Park, K. C. Bang, B. T. Park, H. S. Hwang, I.-Soo Jung, J. I. Kim, D. B. Oh, D. I. Kim, H. Yagi, K. Kato, D. K. Kim, *H. H. Kim, “Glycan Structure and Serum Half-life of Recombinant CTLA4Ig, an Immunosuppressive Agent, Expressed in Suspension-cultured Rice Cells with Coexpression of Human B1,4-galactosyltransferase and Human CTLA4Ig”, *Glycoconjugate J.*, **32**, 161-72, (2015), 10.1007/s10719-015-9590-x,
77. 蟹川暁, 加藤晃一, *森和俊, “糖鎖依存の構造異常タンパク質分解に必須な糖鎖刈り込み機構を解明～革新的ゲノム編集技術によって従来のモデルを一新～”, *化学と生物*, **53**, 571-573 (2015)
78. *加藤晃一, *佐藤匡史, “生命分子の自己組織化のダイナミクス”, *化学工業*, **66**, 32-37, (2015)
79. *S. Sato, Y. Yoshimasa, D. Fujita, M. Yagi-Utsumi, T. Yamaguchi, *K. Kato, *M. Fujita, “Self-Assembled Spherical Complex Displaying a Gangliosidic Glycan Cluster Capable of Interacting with Amyloidogenic Proteins”, *Angew. Chem. Int. Ed.*, **54**, 8435-8439, (2015), 10.1002/anie.201501981
80. M. Tagawa, K. Shirane, L. Yu, T. Sato, S. Furukawa, H. Mizuguchi, R. Kuji, K. Kawamura, N.

- Takahashi, K. Kato, S. Hayakawa, S. Sawada, *K. Furukawa, "Enhanced Expression of the 84-galactosyltransferase 2 Gene Impairs Mammalian Tumor Growth", *Cancer Gene Therapy*, **21**, 219-227, (2014), 10.1038/cgt.2014.21
81. N. Kawasaki, T. Okumoto, Y. Yamaguchi, N. Takahashi, W. H. Fridman, C. Sautès-Fridman, H. Yagi, *K. Kato, "Site-specific Classification of *N*-linked Oligosaccharides of the Extracellular Regions of Fcγ Receptor IIIb Expressed in Baby Hamster Kidney Cells", *J. Glycomics Lipidomics* **4**, 116, (2014), 10.4172/2153-0637.1000116
 82. A. Sikdar, T. Satoh, M. Kawasaki, *K. Kato, "Crystal Structure of Archaeal Homolog of Proteasome-assembly Chaperone PbaA", *Biochem. Biophys. Res. Commun.*, **453**, 493-497, (2014), 10.1016/j.bbrc
 83. *T. Doi, M. Yoshida, K. Ohsawa, K. Shin-ya, M. Takagi, Y. Uekusa, T. Yamaguchi, K. Kato, T. Hirokawa, T. Natsume, "Total Synthesis and Characterization of Thielocin B1 as a Protein-protein Interaction Inhibitor of PAC3 Homodimer", *Chem. Sci.*, **5**, 1860-1868, (2014), 10.1039/C3SC53237B
 84. Y. Kamiya, T. Satoh, *K. Kato, "Recent Advances in Glycoprotein Production for Structural Biology: Toward Tailored Design of Glycoforms", *Curr. Opin. Struct. Biol.*, **26**, 44-53, (2014), 10.1016/j.sbi.2014.03.008
 85. S. Kitazawa, T. Kameda, A. Kumo, M. Yagi-Utsumi, N. J. Baxter, K. Kato, M. P. Williamson, *R. Kitahara, "Close Identity between Alternatively Folded State N2 of Ubiquitin and the Conformation of the Protein Bound to the Ubiquitin-activating Enzyme", *Biochemistry*, **53**(3), 447-449, (2014), 10.1021/bi401617n
 86. T. Satoh, Y. Saeki, T. Hiromoto, Y. H. Wang, Y. Uekusa, H. Yagi, H. Yoshihara, M. Yagi-Utsumi, T. Mizushima, K. Tanaka, *K. Kato, "Structural Basis for Proteasome Formation Controlled by an Assembly Chaperone Nas2", *Structure*, **22**(5), 731-743, (2014), 10.1016/j.str.2014.02.014
 87. T. Satoh, K. Suzuki, T. Yamaguchi, *K. Kato, "Structural Basis for Disparate Sugar-binding Specificities in the Homologous Cargo Receptors ERGIC-53 and VIP36", *PLoS One*, **9**(2), e87963, (2014), 10.1371/journal.pone.0087963
 88. M. Sugiyama, H. Yagi, T. Yamaguchi, K. Kumoi, M. Hirai, Y. Oba, N. Sato, L. Porcar, A. Martel, *K. Kato, "Conformational Characterization of a Protein Complex Involving Intrinsically Disordered Protein by Small-angle Neutron Scattering Using the Inverse Contrast Matching Method: a Case Study of Interaction between α-synuclein and PbaB Tetramer as a Model Chaperone", *J. Appl. Cryst.*, **47**(1), 430-435, (2014), 10.1107/S1600576713033475
 89. K. Takagi, Y. Saeki, H. Yashiroda, H. Yagi, A. Kaiho, S. Murata, T. Yamane, K. Tanaka, T. Mizushima, *K. Kato, "Pba3-Pba4 Heterodimer Acts as a Molecular Matchmaker in Proteasome α-ring Formation", *Biochem. Biophys. Res. Commun.*, **450**(2), 1110-1114, (2014), 10.1016/j.bbrc.2014.06.119
 90. Y. Uekusa, K. Okawa, M. Yagi-Utsumi, O. Serve, Y. Nakagawa, T. Mizushima, H. Yagi, Y. Saeki, K. Tanaka, *K. Kato, "Backbone ¹H, ¹³C, and ¹⁵N Assignments of Yeast Ump1, an Intrinsically Disordered Protein that Functions as a Proteasome Assembly Chaperone", *Biomol. NMR Assign.*, **8**, 383-386, (2014), 10.1007/s12104-013-9523-1
 91. S. Ninagawa, T. Okada, Y. Sumitomo, Y. Kamiya, K. Kato, S. Horimoto, T. Ishikawa, S. Takeda, T. Sakuma, T. Yamamoto, *K. Mori, "EDEM2 Initiates Mammalian Glycoprotein ERAD by Catalyzing the First Mannose Trimming Step," *J. Cell Biol.*, **206**, 347-356, (2014), 10.1083/jcb.201404075
 92. 矢木宏和, 矢木・内海真穂, 加藤晃一, "糖鎖構造生物学の最前線", ファルマシア, **50**, 746-750, (2014)
 93. T. Yamaguchi, Y. Sakae, Y. Zhang, S. Yamamoto, Y. Okamoto, *K. Kato, "Exploration of Conformational Spaces of High-Mannose-Type Oligosaccharides by an NMR-Validated Simulation," *Angew. Chem. Int. Ed.*, **126**, 11121-11124, (2014), 10.1002/ange.201406145
 94. T. Satoh, A. Sumiyoshi, M. Yagi-Utsumi, E. Sakata, H. Sasakawa, E. Kurimoto, Y. Yamaguchi, W. Li, C.A.P. Joazeiro, T. Hirokawa, *K. Kato, "Mode of Substrate Recognition by the Josephin Domain of Ataxin-3, which has an Endo-type Deubiquitinase Activity," *FEBS Lett.*, **588**, 4422-4430, (2014), 10.1016/j.febslet.2014.10.013
 95. T. Zhu, *T. Satoh, *K. Kato, "Structural Insight into Substrate Recognition by the Endoplasmic Reticulum Folding-sensor Enzyme: Crystal Structure of Third Thioredoxin-like Domain of UDP-glucose:Glycoprotein Glucosyltransferase", *Sci. Rep.*, **4**, 7322, (2014), 10.1038/srep07322
 96. H. Yagi, N. Nakagawa, T. Saito, H. Kiyonari, T. Abe, T. Toda, S-W. Wu, K-H. Khoo, *S. Oka, *K. Kato, "AGO61-dependent GlcNAc Modification Primes the Formation of Functional Glycans on α-dystroglycan", *Sci. Rep.*, **3**, 3288, (2013), 10.1038/srep03288
 97. Y. Zhang, T. Yamaguchi, *K. Kato, "New NMR Tools for Characterizing the Dynamic Conformations

- and Interactions of Oligosaccharides”, *Chem. Lett.*, **42**(12), 1455-1462, (2013), [10.1246/cl.130789](https://doi.org/10.1246/cl.130789)
98. K. Araki, S. Iemura, Y. Kamiya, D. Ron, K. Kato, T. Natsume, *K. Nagata, “Ero1- α and PDIs Constitute a Hierarchical Electron Transfer Network of Endoplasmic Reticulum Oxidoreductases” *J. Cell Biol.*, **202**(6), 861-874, (2013), [10.1083/jcb.201303027](https://doi.org/10.1083/jcb.201303027)
99. S. Horimoto, S. Ninagawa, T. Okada, H. Koba, T. Sugimoto, Y. Kamiya, K. Kato, S. Takeda, *K. Mori, “The Unfolded Protein Response Transducer ATF6 Represents a Novel Transmembrane-type Endoplasmic Reticulum-associated Degradation Substrate Requiring Both Mannose Trimming and SEL1L Protein”, *J. Biol. Chem.*, **288**, 31517-31527, (2013), [10.1074/jbc.M113.476010](https://doi.org/10.1074/jbc.M113.476010)
100. 矢木宏和, *加藤晃一, “IgG-Fc と Fc 受容体の複合体形成における糖鎖の役割”, *実験医学*, **31**, 1602-1606, (2013)
101. 矢木宏和, *加藤晃一, “神経幹細胞の幹細胞性維持における複合糖質の役割”, *生化学*, **85**, 1012-1016, (2013)
- 【菊地和也】
1. Y. Hori, N. Otomura, A. Nishida, M. Nishiura, M. Umeno, I. Suetake, K. Kikuchi*, “Synthetic-Molecule/Protein Hybrid Probe with Fluorogenic Switch for Live-Cell Imaging of DNA Methylation”, *J. Am. Chem. Soc.*, **140**, 1686-1690, (2018), [10.1021/jacs.7b09713](https://doi.org/10.1021/jacs.7b09713)
 2. R. Sato, J. Kozuka, *M. Ueda, R. Mishima, *Y. Kumagai, A. Yoshimura, M. Minoshima, S. Mizukami, *K. Kikuchi, “Intracellular Protein Labeling Probes for Multicolor Single-molecule Imaging of Immune Receptor-adaptor Molecular Dynamics”, *J. Am. Chem. Soc.*, **139**, 17397-17404, (2017), [10.1021/jacs.7b08262](https://doi.org/10.1021/jacs.7b08262)
 3. Y. Matsui, *S. Mizukami, *K. Kikuchi, “Ratiometric Imaging of Intracellular Mg²⁺ Dynamics Using a Red Fluorescent Turn-off Probe and a Green Fluorescent Turn-on Probe”, *Chem. Lett.*, **47**, 23-26, (2017), [10.1246/cl.170918](https://doi.org/10.1246/cl.170918)
 4. Y. Hori, S. Hirayama, *K. Kikuchi, “Development of Cyanine Probes with Dinitrobenzene Quencher for Rapid Fluorogenic Protein Labeling”, *Philos. Trans. R. Soc. A*, **375**, 20170018, (2017), [10.1098/rsta.2017.0018](https://doi.org/10.1098/rsta.2017.0018)
 5. Y. Matsui, Y. Funato, H. Imamura, H. Miki, *S. Mizukami, *K. Kikuchi, “Visualization of Long-term Mg²⁺ Dynamics in Apoptotic Cells with a Novel Targetable Fluorescent Probe”, *Chem. Sci.*, **8**, 8255-8264, (2017), [10.1039/c7sc03954a](https://doi.org/10.1039/c7sc03954a)
 6. Y. Matsui, K. K. Sadhu, *S. Mizukami, *K. Kikuchi, “Highly Selective Tridentate Fluorescent Probes for Visualizing Intracellular Mg²⁺ Dynamics without Interference from Ca²⁺ Fluctuation”, *Chem. Commun.*, **53**, 10644-10647, (2017), [10.1039/c7cc06141b](https://doi.org/10.1039/c7cc06141b)
 7. M. Minoshima, *K. Kikuchi, “Photostable and Photoswitching Fluorescent Dyes for Super-resolution Imaging”, *J. Biol. Inorg. Chem.*, **22**, 639-652, (2017), [10.1007/s00775-016-1435-y](https://doi.org/10.1007/s00775-016-1435-y)
 8. S. Mizukami, M. Kashibe, K. Matsumoto, Y. Hori, *K. Kikuchi, “Enzyme-triggered Compound Release Using Functionalized Antimicrobial Peptide Derivatives”, *Chem. Sci.*, **8**, 3047-3052, (2017), [10.1039/C6SC04435B](https://doi.org/10.1039/C6SC04435B)
 9. S. Hirayama, Y. Hori, Z. Benedek, T. Suzuki, *K. Kikuchi, “Fluorogenic Probes Reveal a Role of GLUT4 N-glycosylation in Intracellular Trafficking”, *Nat. Chem. Biol.*, **12**, 853-859, (2016), [10.1038/nchembio.2156](https://doi.org/10.1038/nchembio.2156)
 10. H. Maeda, T. Kowada, J. Kikuta, M. Furuya, M. Shirazaki, S. Mizukami, M. Ishii, *K. Kikuchi, “Real-time Intravital Imaging of pH Variation Associated with Osteoclast Activity and Motility Using Designed Small Molecular Probe”, *Nat. Chem. Biol.*, **12**, 579-585, (2016), [10.1038/nchembio.2096](https://doi.org/10.1038/nchembio.2096)
 11. S. Sotoma, J. Iimura, R. Igarashi, K.M. Hirosawa, H. Ohnishi, S. Mizukami, K. Kikuchi, T.K. Fujiwara, *M. Shirakawa, *H. Tochio, “Selective Labeling of Proteins on Living Cell Membranes Using Fluorescent Nanodiamond Probes”, *Nanomaterials*, **6**, 56, 9p (2016), [10.3390/nano6040056](https://doi.org/10.3390/nano6040056)
 12. T. Wasin, K. Enomoto, T. Sakurai, V. Padalkar, H. Cheng, M. Tang, A. Horio, D., Sakamaki, M. Omichi, A. Saeki, K. Kikuchi, Y. Hori, A. Chiba, Y. Saito, T. Kamiya, M. Sugimoto, *S. Seki, “Fabrication of ‘Clickable’ Polyfluorene Nanowires with High Aspect Ratio as Biological Sensing Platforms”, *ACS Sensors*, **6**, 766-774, (2016), [10.1021/acssensors.6b00070](https://doi.org/10.1021/acssensors.6b00070)
 13. Y. Kamikawa, Y. Hori, K. Yamashita, L. Jin, S. Hirayama, D.M. Standley, *K. Kikuchi, “Design of a Protein tag and Fluorogenic Probe with Modular Structure for Live-Cell Imaging of Intracellular Proteins”, *Chem. Sci.*, **7**, 308-314, (2016), [10.1039/C5SC02351C](https://doi.org/10.1039/C5SC02351C)
 14. Y. Hori, S. Hirayama, M. Sato, *K. Kikuchi, “Redesign of Fluorogenic Labeling System to Improve

- Surface Charges, Brightness, and Binding Kinetics for Imaging Functional Localization of Bromodomains”, *Angew. Chem. Int. Ed.*, **54**, 14368-14371, (2015), 10.1002/anie.201506935
15. Z. Zhang, S. Mizukami, K. Fujita, *K. Kikuchi, “An Enzyme-Responsive Metal-Enhanced Near-Infrared Fluorescence Sensor Based on Functionalized Gold Nanoparticles”, *Chem. Sci.*, **6**, 4934-4939, (2015), 10.1039/C5SC01850A
 16. K. Mochizuki, L. Shi, S. Mizukami, M. Yamanaka, M. Tanabe, W.T. Gong, A.F. Palonpon, S. Kawano, S. Kawata, K. Kikuchi, *K. Fujita, “Nonlinear Fluorescence Imaging by using Photoinduced Charge Separation”, *Jpn. J. Appl. Phys.*, **54**, 042403, (2015), <http://dx.doi.org/10.7567/JJAP.54.042403>
 17. *K. Kikuchi, “¹⁹F MRI Probes with Tunable Switches and Highly Sensitive ¹⁹F MRI Nano-probes”, *Bull. Chem. Soc. Japan*, **88**, 518-522, (2015), 10.1246/bcsj.20140392
 18. M. Minoshima, *K. Kikuchi, “Chemical Probes for Elucidating Histone Deacetylase Function”, *Anal. Sci.*, **31**, 287-292, (2015), <http://doi.org/10.2116/analsci.31.287>
 19. T. Kowada, Y. Hori, *K. Kikuchi, “BODIPY-based Fluorescent Probes for Biological Applications”, *Chem. Soc. Rev.*, **44**, 4953-4972, (2015), 10.1039/c5cs00030k
 20. T. Nakamura, F. Sugihara, H. Matsushita, Y. Yoshioka, S. Mizukami, *K. Kikuchi, “Mesoporous Silica Nanoparticles for ¹⁹F Magnetic Resonance Imaging, Fluorescence Imaging, and Drug Delivery”, *Chem. Sci.*, **6**, 1986-1990, (2015), 10.1039/C4SC03549F
 21. R. Baba, Y. Hori, *K. Kikuchi, “Intramolecular Long-distance Nucleophilic Reactions as a Rapid Fluorogenic Switch Applicable to detection of Enzymatic Activity”, *Chem. Eur. J.*, **21**, 4695-4702, (2015), 10.1002/chem.201406093
 22. T. Nakamura, H. Matsushita, F. Sugihara, Y. Yoshioka, S. Mizukami, *K. Kikuchi, “Activatable ¹⁹F MRI Nanoparticle Probes for the Detection of Reducing Environments”, *Angew. Chem. Int. Ed.*, **54**, 1007-1010, (2015), 10.1002/anie.201409365
 23. M. Minoshima, T. Matsumoto, *K. Kikuchi, “Development of Fluorogenic Probe based on a DNA Staining Dye for Continuous Monitoring of the Histone Deacetylase Reaction”, *Anal. Chem.*, **86**(15), 7925-7930, (2014), [10.1021/ac501881s](https://doi.org/10.1021/ac501881s)
 24. S. Okada, S. Mizukami, T. Sakata, Y. Matsumura, Y. Yoshioka, *K. Kikuchi, “Ratiometric MRI Sensors Based on Core-Shell Nanoparticles for Quantitative pH Imaging”, *Adv. Mater.*, **26**(19), 2989-2992, (2014), [10.1002/adma.201305018](https://doi.org/10.1002/adma.201305018)
 25. H. Matsushita, S. Mizukami, F. Sugihara, Y. Nakanishi, Y. Yoshioka, *K. Kikuchi, “Multifunctional Core-shell Silica Nanoparticles for Highly Sensitive ¹⁹F MRI”, *Angew. Chem. Int. Ed.*, **53**(4), 1008-1011, (2014), [10.1002/anie.201308500](https://doi.org/10.1002/anie.201308500)
 26. A. Yoshimura, S. Mizukami, Y. Mori, Y. Yoshioka, *K. Kikuchi, “¹H MRI Detection of Gene Expression in Living Cells by Using Protein Tag and Biotinylation Probe”, *Chem. Lett.*, **43**, 219-221, (2014), [10.1246/cl.130942](https://doi.org/10.1246/cl.130942)
 27. S. Mizukami, Y. Hori, *K. Kikuchi, “Small-Molecule-Based Protein-Labeling Technology in Live Cell Studies: Probe-Design_Concepts and Applications”, *Acc. Chem. Res.*, **47**(1), 247–256, (2014), [10.1021/ar400135f](https://doi.org/10.1021/ar400135f)

【佐甲靖志】

1. M. Yanagawa., M. Hiroshima., Y. Togashi,, T. Yamashita, Y. Shichida, M. Murat., M. Ueda, *Y. Sako, “Single-molecule Diffusion-based Estimation of GPCR Activity”, *Sci. Sig.*, in press, (2018), 10.1101/205161
2. M. Yasui, M. Hiroshima, J. Kozuka, *Y. Sako, *M. Ueda, “Automated Single-molecule Imaging in Living Cells”, *Nat. Comm.*, **9**, 3061, (2018), 10.1038/s41467-018-05524-7
3. S. Magi, K. Iwamoto, N. Yumono, M. Hiroshima, T. Nagashima, R. Ohki, A. Garcia-Munoz, N. Volinsky, A. Von Kriegsheim, Y. Sako, K. Takahashi, S. Kimura, B. N. Kholodenko, *M. Okada-Hatakeyama, “Transcriptionally Inducible Pleckstrin Homology-like Domain Family A Member 1 Attenuates ErbB Receptor Activity by Inhibiting Receptor Oligomerization,” *J. Biol. Chem.*, **293**, 2206, (2018), [10.1074/jbc.M117.778399](https://doi.org/10.1074/jbc.M117.778399)
4. R. Maeda, T. Sato, K. Okamoto, M. Yanagawa, *Y. Sako, “Lipid-protein Interplay in Dimerization of the Juxtamembrane Domains of Epidermal Growth Factor Receptor,” *Biophys. J.*, **114**, 893, (2018), 10.1016/j.bpj.2017.12.029
5. M. Hiroshima, C.-g. Pack, K. Kaizu, K. Takahashi, M. Ueda, *Y. Sako, “Transient Acceleration of Epidermal Growth Factor Receptor Dynamics Produces Higherorder Signaling Clusters,” *J. Mol. Biol.*,

430(9), 1386-1401, (2018), 10.1016/j.jmb.2018.02.018

6. *Y. Shindo, Y. Kondo, *Y. Sako, "Inferring a Nonlinear Biochemical Network Model from a Heterogeneous Single-cell Time Course Data," *Sci. Rep.*, **8**, 6790, (2018), [10.1038/s41598-018-25064-w](https://doi.org/10.1038/s41598-018-25064-w)
7. R. Maeda, M. Hiroshima, T. Yamashita, A. Wada, Y. Sako, Y. Shichida, *Y. Imamoto, "Shift in Conformational Equilibrium Induces Constitutive Activity of G-Protein Coupled Receptor, Rhodopsin," *J. Phys. Chem. B*, **122**, 4838-4843, (2018), [10.1021/acs.jpcb.8b02819](https://doi.org/10.1021/acs.jpcb.8b02819)
8. K. Okamoto., M. Hiroshima, *Y. Sako, "Single-molecule Fluorescence Based Analysis of Protein Conformation, Interaction, and Oligomerization in Cellular Systems," *Biophys. Rev.*, **10**, 317-326, (2017), [10.1007/s12551-017-0366-3](https://doi.org/10.1007/s12551-017-0366-3)
9. Y. Nakamura, N. Umeki, M. Abe *Y. Sako, "Mutation-specific Mechanisms of Hyperactivation of Noonan Syndrome SOS Molecules Detected with Single-molecule Imaging in Living Cells," *Sci. Rep.* **7**, 14153, (2017), [10.1038/s41598-017-14190-6](https://doi.org/10.1038/s41598-017-14190-6)
10. *R. Iino, T. Iida, A. Nakamura, E. Saita, H. You, Y. Sako, "Single-molecule Imaging and Manipulation of Biomolecular Machines and Systems", *Biochim. Biophys. Acta, (BBA), Gen. Subj.*, **1862**, 241-252, (2017), [10.1016/j.bbagen.2017.08.008](https://doi.org/10.1016/j.bbagen.2017.08.008)
11. R. Yoshizawa, *N. Umeki, M. Yanagawa, M. Murata, *Y. Sako, "Single-molecule Fluorescence Imaging of RalGDS on Cell Surfaces during Signal Transduction to Ras to Ral," *Biophys. Physicobiol.* **14**, 75-84, (2017), [10.2142/biophysico.14.0_75](https://doi.org/10.2142/biophysico.14.0_75)
12. *K. Okamoto, Y. Sako, "Recent Advances in FRET for the Study of Protein Interactions and Dynamics", *Curr. Opin. Struct. Biol.*, **46**, 16-23, (2017), [10.1016/j.sbi.2017.03.010](https://doi.org/10.1016/j.sbi.2017.03.010)
13. *Y. Arata, M. Hiroshima, C.-G. Pack, R. Ramanujam, F. Motegi, K. Nakazato, P. W. Wiseman, H. Sawa, T. J. Kobayashi, H. B. Brandão, T. Shibata, *Y. Sako, "Cortical Polarity of the RING Protein PAR-2 is Maintained by Exchange Rate Kinetics at the Cortical-cytoplasm Boundary", *Cell Rep.*, **16**, 2156-2168, (2016), [10.1016/j.celrep.2016.07.047](https://doi.org/10.1016/j.celrep.2016.07.047)
14. 新土優樹、小迫英尊、佐甲靖志、高橋恒一, "細胞内シグナルのアナログ・デジタル変換" *生物物理*, **56**, 334, (2016), [10.2149/biophys.56.334](https://doi.org/10.2149/biophys.56.334)
15. K. Okamoto, *Y. Sako, "State Transition Analysis of Spontaneous Branch Migration of the Holliday Junction by Photon-based Single-molecule Fluorescence Resonance Energy Transfer", *Biophys. Chem.*, **209**, 21-27, (2016), [10.1016/j.bpc.2015.11.004](https://doi.org/10.1016/j.bpc.2015.11.004)
16. Y. Nakamura, K. Hibino, T. Yanagida, *Y. Sako, "Switching of the Positive Feedback for RAS Activation by a Concerted Function of SOS Membrane Association Domains", *Biophys. Physicobiol.*, **13**, 1-11, (2016), [10.2142/biophysico.13.0_1](https://doi.org/10.2142/biophysico.13.0_1)
17. S. Iwata, K. Masuhara, N. Umeki, Y. Sako, *S. Maruta, "Interaction of a Novel fluorescent GTP Analogue with the Small G-protein K-Ras", *J. Biochem.*, **159**, 41-48, (2015), [10.1093/jb/mvv071](https://doi.org/10.1093/jb/mvv071)
18. M. Yanagawa, K. Kojima, T. Yamashita, Y. Imamoto, T. Matsuyama, K. Nakanishi, Y. Yamano, A. Wada, Y. Sako, *Y. Shichida, "Origin of the Low Thermal Isomerization Rate of Rhodopsin Chromophore", *Sci. Rep.* **5**, 11081, (2015), [10.1038/srep11081](https://doi.org/10.1038/srep11081)
19. Y. Zhou, H. Mao, B. Joddar, N. Umeki, Y. Sako, C. Nishioka, E. Takahashi, K. Wada, Y. Wang, *Y. Ito, "The Significance of Membrane Fluidity of Feeder Cell-derived Substrates for Maintenance of iPS Cell Stemness", *Sci. Rep.* **5**, 11386 (1-13), (2015), [10.1038/srep11386](https://doi.org/10.1038/srep11386)
20. H. Park, S.-S. Han, Y. Sako, *C.-G. Pack, "Dynamic and Unique Nucleolar Microenvironment Revealed by Fluorescence Correlation Spectroscopy", *FASEB J.* **29**, 837-848, (2015), [10.1096/fj.14-254110](https://doi.org/10.1096/fj.14-254110)
21. H. Shinohara, M. Behar, K. Inoue, M. Hiroshima, T. Yasusda, T. Nagashima, S. Kimura, H. Sanjo, S. Maeda, N. Yumoto, S. Ki, S. Akira, Y. Sako, A. Hoffman, T. Kurosaki, *M. Okada-Hatakeyama, "Positive Feedback within a Kinase Signaling Complex Functions as a Switch Mechanism for NF- κ B Activation", *Science*, **344**(6185), 760-764, (2014), [10.1126/science.1250020](https://doi.org/10.1126/science.1250020)

【笛井理生】

1. T. Okuno, K. Kato, S. Minami, T. P. Terada, M. Sasai, *G. Chikenji, "Importance of Consensus Region of Multiple-ligand Templates in a Virtual Screening Method", *Biophys. Physicobiol.* **13**, 149-156, (2016), [10.2142/biophysico.13.0_149](https://doi.org/10.2142/biophysico.13.0_149)
2. *笛井理生, 寺田智樹, "真核細胞のルースな遺伝子制御とクロマチン動態", *生物物理*, **56**, 106-108, (2016)
3. *K. Maeshima, S. Ide, K. Hibino, M. Sasai, "Liquid-like Behavior of Chromatin", *Curr Opin Genet Dev.*, **37**, 36-45, (2016), [10.1016/j.gde.2015.11.006](https://doi.org/10.1016/j.gde.2015.11.006)
4. S. S. Ashwin, *M. Sasai, "Effects of Collective Histone State Dynamics on Epigenetic Landscape and

- Kinetics of Cell Reprogramming”, *Sci. Rep.*, **5**, 16746, (2015), 10.1038/srep16746
5. C. Chen, K. Zhang, H. Feng, M. Sasai, *J. Wang, “Multiple Coupled Landscapes and Non-diabatic Dynamics with Applications to Self Activating Genes”, *Phys. Chem. Chem. Phys.*, **17**, 29036-29044, (2015), 10.1039/C5CP04780C
 6. T. Okuno, K. Kato, T. Terada, M. Sasai, *G. Chikenji, “VS-APPLE: A Virtual Screening Algorithm Using Promiscuous Protein-Ligand Complexes”, *J. Chem. Inf. Model.*, **55**(6), 1108–1119, (2015), [10.1021/acs.jcim.5b00134](https://doi.org/10.1021/acs.jcim.5b00134)
 7. T. Inanami, T. P. Terada, *M. Sasai, “Coupling of Lever Arm Swing and Biased Brownian Motion in Actomyosin”, *Proc.Natl.Acad.Sci., USA*, **111**(45), 15969–15974, (2014), [10.1371/journal.pcbi.1003552](https://doi.org/10.1371/journal.pcbi.1003552)

【佐藤健】

1. A. Sakaguchi, M. Sato, K. Sato, K. Gengyo-Ando, T. Yorimitsu, J. Nakai, T. Hara, K. Sato, *K. Sato, “REI-1 is a Guanine Nucleotide Exchange Factor Regulating RAB-11 Localization and Function in *C. elegans* embryos”, *Dev. Cell.*, **35**, 211-221, (2015), [10.1016/j.devcel.2015.09.013](https://doi.org/10.1016/j.devcel.2015.09.013)
2. K. Ishii, H. Enda, M. Noda, M. Kajino, A. Kim, E. Kurimoto, K. Sato, A. Nakano, Y. Kobayashi, H. Yagi, S. Uchiyama, *K. Kato, “pH-dependent Assembly and Segregation of the Coiled-coil Segments of Yeast Putative Cargo Receptors Emp46p and Emp47p”, *PLoS ONE*, **10**, e140287, (2015), [10.1371/journal.pone.0140287](https://doi.org/10.1371/journal.pone.0140287)
3. H. Iwasaki, T. Yorimitsu, *K. Sato, “Distribution of Sec24 Isoforms to each ER Exit Site is Dynamically Regulated in *Saccharomyces Cerevisiae*”, *FEBS Lett.*, **589**, 1234-1239, (2015), [10.1016/j.febslet.2015.04.006](https://doi.org/10.1016/j.febslet.2015.04.006)
4. T. Yorimitsu, K. Sato, *M. Takeuchi, “Molecular Mechanisms of Sar/Arf GTPases in Vesicular Trafficking in Yeast and Plant.”, *Front.Plant Sci.*, **5**, 411, (2014), [10.3389/fpls.2014.00411](https://doi.org/10.3389/fpls.2014.00411)
5. C. Kodera, T. Yorimitsu, *K. Sato, “Sec23 Homolog Nel1 is a Novel GTPase-activating Protein for Sar1 but Does Not Function as a Subunit of the COPII Coat.”, *J. Biol. Chem.*, **289**, 21423-21432, (2014), [10.1074/jbc.M114.553917](https://doi.org/10.1074/jbc.M114.553917)
6. K. Ebine, T. Inoue, J. Ito, E. Ito, T. Uemura, T. Goh, H. Abe, K. Sato, A. Nakano, *T. Ueda, “Plant Vacuolar Trafficking Occurs through Distinctly Regulated Pathway.”, *Curr. Biol.*, **24**(12), 1375-1382, (2014), [10.1016/j.cub.2014.05.004](https://doi.org/10.1016/j.cub.2014.05.004)

【申惠媛】

1. *申惠媛、高津宏之細胞膜ホスファチジルセリン - フリップペイゼの活性調節機構. *生化学*, **90**, 486-490, (2018), [10.14952/SEIKAGAKU.2018.900486](https://doi.org/10.14952/SEIKAGAKU.2018.900486).
2. *申惠媛脂質二重層間のリン脂質の移動（フリップーフロップ）による細胞膜の変形. *実験医学*, **36**, 2239-2242, (2018)
3. N. Takada, T. Naito, T. Inoue, K. Nakaama, H. Takatsu, *H.-W. Shin, “Phospholipid-flipping Activity of P4-ATPase Drives Membrane Curvature,” *EMBO J.*, **37**, e97705, (2018), [10.15252/embj.201797705](https://doi.org/10.15252/embj.201797705)
4. H. Takatsu, M. Takayama, T. Naito, N. Takada, K. Tsumagari, Y. Ishihama, K. Nakayama, *H.-W. Shin, “Phospholipid Flippase ATP11C is Endocytosed and Downregulated Following Ca2+-mediated Protein Kinase C Activation,” *Nat. Commun.*, **8**, 1423, (2017), [10.1038/s41467-017-01338-1](https://doi.org/10.1038/s41467-017-01338-1)
5. K. H. Tomaszowski, N. Hellmann, V. Ponath, H. Takatsu, H.-W. Shin, *B. Kaina, “Uptake of Glucose-conjugated MGMT Inhibitors in Cancer Cells: Role of Flippases and Type IV P-type ATPases,” *Sci. Rep.*, **7**, 13925, (2017), [10.1038/s41598-017-14129-x](https://doi.org/10.1038/s41598-017-14129-x)
6. Y. Tanaka, N. Ono, T. Shima, G. Tanaka, Y. Katoh, K. Nakayama, H. Takatsu, *H.-W. Shin, “The Phospholipid Flippase ATP9A is Required for Recycling Pathway from Endosomes to the Plasma Membrane”, *Mol. Biol. Cell.*, **27**, 3883, (2016), [10.1091/mbc.E16-08-0586](https://doi.org/10.1091/mbc.E16-08-0586)
(Selected for highlights)
7. R. Miyano, T. Matsumoto, H. Takatsu, K. Nakayama, *H.-W. Shin, “Alteration of Transbilayer Phospholipid Compositions is Involved in Cell Adhesion, Cell Spreading, and Focal Adhesion Formation”, *FEBS Lett.*, **590**, 2138, (2016), [10.1002/1873-3468.12247](https://doi.org/10.1002/1873-3468.12247)
8. A. Hanai, M. Ohgi, C. Yagi, T. Ueda, H.-W. Shin, *K. Nakayama, “Class I Arfs (Arf1 and Arf3) and Arf6 are Localized to the Flemming Body and Play Important Roles in Cytokinesis”, *J. Biochem.*, **159**, 201, (2016), [10.1093/jb/mvv088](https://doi.org/10.1093/jb/mvv088)

【真行寺千佳子】

1. H. Yoke, *C. Shingyoji, “Effects of External Strain on the Regulation of Microtubule Sliding Induced by Outer arm Dynein of Sea Urchin Sperm Flagella”, *J. Exp. Biology*, 1122-1134, (2017), [10.1242/jeb.147942](https://doi.org/10.1242/jeb.147942)

2. *C. Shingyoji, I. Nakano, Y. Inoue, H. Higuchi, “Dynein Arms are Strain-dependent Direction-switching Force Generators,” *Cytoskeleton*, **72**, 388-401, (2015), 10.1002/cm.21232
【杉山正明】
1. *Y. Nagata, T. Nishikawa, *M. Suginome, S. Sato, *M. Sugiyama, L. Porcar, A. Martel, R. Inoue, N. Sato, “Elucidating the Solvent Effect on the Switch of the Helicity of Poly(quinoxaline-2,3-diyl)s: A Conformational Analysis by Small-Angle Neutron Scattering”, *J. Am. Chem. Soc.*, **140**, 2722-2726, (2018), 10.1021/jacs.7b11626
2. M. Yagi-Utsumi, A. Sikdar, T. Kozai, R. Inoue, M. Sugiyama, T. Uchihashi, H. Yagi, T. Satoh, K. Kato, “Conversion of Functionally Undefined Homopentameric Protein PbaA into a Proteasome Activator by Mutational Modification of its C-terminal Segment Conformation”, *Protein Eng. Des.Sel.*, **31**, 29-36, (2018), 10.1093/protein/gzx066
3. P. Bernadó, N. Shimizu, G. Zaccai, H. Kamikubo, M. Sugiyama, “Solution Scattering Approaches to Dynamical Ordering in Biomolecular Systems”, *Biochim. Biophys. Acta, (BBA), Gen. Subj.*, **1862**, 253-274, (2017), 10.1016/j.bbagen.2017.10.015
4. E. Hibino, R. Inoue, M. Sugiyama, J. Kuwahara, K. Matsuzaki, M. Hoshino, “Identification of Heteromolecular Binding Sites in Transcription Factors Sp1 and TAF4 Using High-resolution NMR Spectroscopy”, *Protein Science*, **26**, 2280-2290, (2017), 10.1002/pro.3287
5. R. Yogo, S. Yanaka, H. Yagi, A. Martel, L. Porcar, Y. Ueki, R. Inoue, N. Sato, M. Sugiyama, K. Kato, “Characterization of Conformational Deformation-coupled Interaction between Immunoglobulin G1 Fc Glycoprotein and a Low-affinity Fcγ Receptor by Deuteration-assisted Small-angle Neutron Scattering”, *Biochem.Biophys. Rep.*, **12**, 1-4, (2017), 10.1016/j.bbrep.2017.08.004
6. J. Trewhella, A. P. Duff, D. Durand, F. Gabel, J. M. Guss, W. A. Hendrickson, G. L. Hura, D. A. Jacques, N. M. Kriby, A. H. Kwan, J. Perez, L. Pollack, T. M. Ryan, A. Sali, D. Schneidman-Duhovny, T. Schwede, D. I. Svergun, M. Sugiyama, J. A. Tainer, P. Vachette, J. Westbrook, A. E. Whitten, “2017 Publication Guidelines for Structural Modelling of Small-angle Scattering Data from Biomolecules in Solution: an Update”, *Acta Crystallographica*, **D73**, (2017), 710-728, 10.1107/S2059798317011597
7. M. Sugiyama, H. Nakagawa, R. Inoue, and Y. Kawakita, “Neutron Biology for Next Generation”, *J-PARC-Review*, 2017-024, (2017), 1-46, (Japanese)
8. *M. Sugiyama, H. Yagi, K. Ishii, L. Porcar, A. Martel, K. Oyama, M. Noda, Y. Yunoki, R. Murakami, R. Inoue, N. Sato, Y. Oba, K. Terauchi, S. Uchiyama, *K. Kato, “Structural Characterization of the Circadian Clock Protein Complex Composed of KaiB and KaiC by Inverse Contrast-matching Small-angle Neutron Scattering”, *Sci Rep.*, **6**, 35567, (2016), 10.1038/srep35567
9. E. Hibino, R. Inoue, M. Sugiyama, J. Kuwahara, K. Matsuzaki, *M. Hoshino, “Interaction between Intrinsically Disordered Regions in Transcription Factors Sp1 and TAF4”, *Protein Sci.*, **25**, 2006-2017, (2016), 10.1002/pro.3013
10. R. Inoue, T. Takata, N. Fujii, K. Ishii, D. N. Sato, Y. Oba, K. Wood, K. Kato, N. Fujii, *M. Sugiyama, “New Insight into the Dynamical System of αB-crystallin Oligomers”, *Sci Rep.*, **6**, 29208, (2016), 10.1038/srep29208
11. *杉山正明, “中性子小角散乱による溶液中のタンパク質の構造解析”, 展望(日本アイソトープ協会学会誌)、3月号, **743**, 2-6, 2016年
12. Y. Takemoto, T. Yamamoto, N. Ikuma, Y. Uchida, K. Suzuki, S. Shimono, H. Takahashi, N. Sato, Y. Oba, R. Inoue, M. Sugiyama, H. Tsue, T. Kato, J. Yamauchi, R. Tamura, “Preparation, Characterization and Magnetic Behavior of a Spin-labelled Physical Hydrogel Containing a Chiral Cyclic Mitroxide Radical Unit Fixed Inside the Gelator Molecule”, *Soft Matter*, **11**, 5563-5570, (2015), 10.1039/C5SM01216C
13. *E. Chatani, *R. Inoue, H. Imamura, M. Sugiyama, Mi. Kato, M. Yamamoto, K. Nishida, T. Kanaya, “Early Aggregation Preceding the Nucleation of Insulin Amyloid Fibrils as Monitored by Small Angle X-ray Scattering”, *Sci. Rep.*, **5**, 15485, (2015), 10.1038/srep15485
14. *M. Sugiyama, N. Horikoshi, Y. Suzuki, H. Taguchib, T. Kujirai, R. Inoue, Y. Oba, N. Sato, A. Martel, L. Porcar, *H. Kurumizaka, “Solution Structure of Variant H2A.Z.1 Nucleosome Investigated by Small-angle X-ray and Neutron Scatterings” *Biochem. Biophys. Rep.*, **4**, 28-32, (2015), 10.1016/j.bbrep.2015.08.019
15. *N. Sato, A. Matsumiya, Y. Higashino, S. Funaki, Y. Kitao, Y. Oba, R. Inoue, F. Arisaka, Fumio, *M. Sugiyama, *R. Urade, “Molecular Assembly of Wheat Gliadins into Nanostructures: A Small-Angle X-Ray Scattering Study of Gliadins in Distilled Water over a Wide Concentration Range”, *J. Agric. Food*

Chem., **63**, 8715–8721, (2015), 10.1021/acs.jafc.5b02902

16. *M. Sugiyama, Y. Arimura, K. Shirayama, R. Fujita, Y. Oba, N. Sato, R. Inoue, T. Oda, M. Sato, R. K. Heenan, H. Kurumizaka, “Distinct Features of the Histone Core Structure in Nucleosomes Containing the Histone H2A.B Variant”, *Biophys. J.*, **106**, 2206-2213, (2014), 10.1016/j.bpj.2014.04.007
17. *N. Rahman, N. Sato, M. Sugiyama, Y. Hidaka, H. Okabe, K. Hara, “The Effect of Hot DMSO Treatment on the γ -ray-induced Grafting of Acrylamide onto PET films”, *Polym. J.*, **46**, 412–421, (2014), 10.1038/pj.2014.12
18. N. Rahman, N. Sato, M. Sugiyama, Y. Hidaka, H. Okabe, K. Hara, “Selective Hg(II) Adsorption from Aqueous Solutions of Hg(II) and Pb(II) by Hydrolyzed Acrylamide-grafted PET films”, *J. Environ. Sci. Health., Part A*, **49**, 798-806, (2014), 10.1080/10934529.2014.882209
19. *K. Mori, K. Iwase, Y. Oba, T. Fukunaga, M. Sugiyama, “Surface Observation of LaNi5 under Deuterium Atmosphere Using Small-Angle Neutron Scattering”, *Mater. Trans.*, **55**, 1643-1646, (2014), 10.2320/matertrans.M2014009
20. Y. Oba, S. Abe, M. Ohnuma, N. Sato, M. Sugiyama, “Temperature Dependence of the Nanostructure in a PbSe–ZnSe Composite Thin Film”, *J. Phys. D: Appl. Phys.*, **47**, (2014), 435102/1-6
21. *M. Sugiyama*, H. Yagi, T. Yamaguchi, K. Kumoi, M. Hirai, Y. Oba, N. Sato, L. Porcar, A. Martel, *K. Kato, “Conformational Characterization of a Protein Complex Involving Intrinsically Disordered Protein by Small-angle Neutron Scattering Using the Inverse Contrast Matching Method: a Case Study of Interaction between α -synuclein and PbaB Tetramer as a Model Chaperone”, *J. Appl. Crystallogr.*, **47**, 430–435, (2014), 10.1107/S1600576713033475

【茶谷 紘理】

1. N. Yamamoto, S. Ito, M. Nakanishi, E. Chatani, K. Inoue, H. Kandori, K. Tominaga, “Effect of Temperature and Hydration Level on Purple Membrane Dynamics Studied Using Broadband Dielectric Spectroscopy from sub-GHz to THz Regions” *J. Phys. Chem. B*, **122**, 1367-1377, (2018), 10.1021/acs.jpcb.7b10077
2. N. Yamamoto, S. Tsuhara, A. Tamura, *E. Chatani, “A Specific Form of Prefibrillar Aggregates that Functions as a Precursor of Amyloid Nucleation” *Sci. Rep.*, **8**, 62, (2018), 10.1038/s41598-017-18390-y
3. E. Chatani, N. Yamamoto, “Recent Progress on Understanding the Mechanisms of Amyloid Nucleation” *Biophys. Rev.*, **10**, 527-534, (2018), [10.1007/s12551-017-0353-8](https://doi.org/10.1007/s12551-017-0353-8)
4. T.H. Liu, K.I. Yuyama, T. Hiramatsu, N. Yamamoto, *E. Chatani, *H. Miyasaka, T. Sugiyama *H. Masuhara, “Femtosecond-Laser-Enhanced Amyloid Fibril Formation of Insulin” *Langmuir*, **33**, 8311-8318, (2017), [10.1021/acs.langmuir.7b01822](https://doi.org/10.1021/acs.langmuir.7b01822)
5. A. Nitani, H. Muta, M. Adachi, M. So, K. Sasahara, K. Sakurai, E. Chatani, K. Naoe, H. Ogi, D. Hall, *Y. Goto, “Heparin-dependent Aggregation of Hen Egg White Lysozyme Reveals two Distinct Mechanisms of Amyloid Fbrillation” *J. Biol. Chem.*, in press, (2017), [10.1074/jbc.M117.813097](https://doi.org/10.1074/jbc.M117.813097)
6. *E. Chatani, *R. Inoue, H. Imamura, M. Sugiyama, M. Kato, M. Yamamoto, K. Nishida, T. Kanaya, “Early Aggregation Preceding the Nucleation of Insulin Amyloid Fibrils as Monitored by Small Angle X-ray Scattering”, *Sci. Rep.*, **5**, 15485, (2015), 10.1038/srep15485
7. *井上 倫太郎、茶谷 紘理、金谷 利治, 「小角X線散乱によるアミロイド線維形成機構に関する研究」, *SPring-8/SACLA利用研究成果集* **3**, 2011B1996/BL40B2, (2015)
8. **茶谷 紘理, 「プログラムされていないフォールディングーアミロイド線維の形成ー」 *生化学* **87**, 292-297, (2015)
9. *E. Chatani, H. Imamura, N. Yamamoto, *M. Kato, “Stepwise Organization of the β -structure Identifies Key Regions Essential for the Propagation and Cytotoxicity of Insulin Amyloid Fibrils”, *J. Biol. Chem.*, **289**, 10399-10410, (2014), [10.1074/jbc.M113.520874](https://doi.org/10.1074/jbc.M113.520874)
10. *E. Chatani, Y. Tsuchisaka, Y. Masuda, *R. Tsenkova, “Water Molecular System Dynamics Associated with Amyloidogenic Nucleation as Revealed by Real Time Near Infrared Spectroscopy and Aquaphotomics”, *PLoS One* **9**, e101997, (2014), [10.1371/journal.pone.0101997](https://doi.org/10.1371/journal.pone.0101997)

【寺内一姫】

1. *T. Teramoto, C. Azai, K. Terauchi, M. Yoshimura, T. Ohta, “Soft X-ray Imaging of Cellular Carbon and Nitrogen Distributions in Heterocystous Cyanobacterium”, *Plant Physiology*, **177**, 52-61 (2018), [10.1104/pp.17.01767](https://doi.org/10.1104/pp.17.01767)
2. K. Oyama, C. Azai, J. Matsuyama, * K. Terauchi, “Phosphorylation at Thr432 Induces Structural

- Destabilization of the CII Ring in the Circadian Oscillator KaiC”, *EBS Lett*, **592**, 36–45, (2018), 10.1002/1873-3468.12945
3. C. Azai, N. Kobayashi, T. Mizoguchi, H. Tamiaki, K. Terauchi, *Y. Tsukatani, “Rapid C8-vinyl Reduction of Divinyl-chlorophyllide a by BciA from *Rhodobacter capsulatus*”, *J. Photochem. Photobiol. A: Chemistry*, **353**, 661-666, (2018), [10.1016/j.jphotochem.2017.09.010](https://doi.org/10.1016/j.jphotochem.2017.09.010)
 4. *寺内一姫, 大山克明, 浅井智広, 「ブルーネイティブ電気泳動による時計タンパク質 KaiC の動的構造解析」, 電気泳動, **61**, 107-110, (2017)
 5. *T. Teramoto, M. Yoshimura, C. Azai, K. Terauchi, T. Ohota, “Determination of Carbon-to-nitrogen Ratio in the Filamentous and Heterocystous Cyanobacterium *Anabaena* sp. PCC 7120 with Single-cell soft X-ray Imaging”, *J. Phys.:Conf. Ser.*, **849**, 012005, (2017), [10.1088/1742-6596/849/1/012005](https://doi.org/10.1088/1742-6596/849/1/012005)
 6. K. Terauchi, R. Sobue, Y. Furutani, R. Aoki, *Y. Fujita, “Isolation of Cyanobacterial Mutants Exhibiting Growth Defects under Microoxic Conditions by Transposon Tagging Mutagenesis of *Synechocystis* sp. PCC 6803.” *J. Gen. Appl. Microbiol.*, **63**, 131-138, (2017), [10.2323/jgam.2016.08.004](https://doi.org/10.2323/jgam.2016.08.004)
 7. K. Oyama, C. Azai, K. Nakamura, S. Tanaka, *K. Terauchi, “Conversion between Two Conformational States of KaiC is Induced by ATP Hydrolysis as a Trigger for Cyanobacterial Circadian Oscillation” *Sci. Rep.*, **6**, 32443, (2016), [10.1038/srep32443](https://doi.org/10.1038/srep32443)
 8. M. Sugiyama, H. Yagi, K. Ishii, L. Porcar, A. Martel, K. Oyama, M. Noda, Y. Yunoki, R. Murakami, R. Inoue, N. Sato, Y. Oba, K. Terauchi, S. Uchiyama, *K. Kato, ‘Structural Characterization of the Circadian Clock Protein Complex Composed of KaiB and KaiC by Inverse Contrast Matching Small-angle Neutron Scattering”, *Sci. Rep.*, **6**, 35567, (2016), [10.1038/srep35567](https://doi.org/10.1038/srep35567)
 9. J. Nomata, K. Terauchi, *Y. Fujita, “Stoichiometry of ATP Hydrolysis and Chlorophyllide Formation of Dark-operative Protochlorophyllide Oxidoreductase from Rhodobacter Capsulatus”, *Biochem. Biophys. Res. Commun.*, **470**, 704-709, (2016), [10.1016/j.bbrc.2016.01.070](https://doi.org/10.1016/j.bbrc.2016.01.070)
 10. 大山克明, 浅井智広, *寺内一姫 “3つの時計タンパク質によるシアノバクテリア生物時計再構成系の解析”, 光合成研究, **25**, 175-180, (2015)
 11. Y. Hiraide, K. Oshima, T. Fujisawa, K. Uesaka, Y. Hirose, R. Tsujimoto, H. Yamamoto, S. Okamoto, Y. Nakamura, K. Terauchi, T. Omata, K. Ihara, M. Hattori, *Y. Fujita, “Loss of Cytochrome cM Stimulates Cyanobacterial Heterotrophic Growth in the Dark”, *Plant Cell Physiol.*, **56**, 334-345, (2015), [10.1093/pcp/pcu165](https://doi.org/10.1093/pcp/pcu165)

【水野健作】

1. K. Takahashi, T. Nagai, S. Chiba, K. Nakayama, *K. Mizuno, “Glucose Deprivation Induces Primary Cilium Formation through mTORC1 Inactivation,” *J. Cell Sci.*, **131**, jcs208769, (2018), [10.1242/jcs.208769](https://doi.org/10.1242/jcs.208769)
2. C. Wille, T. Eiseler, S. T. Langenberger, J. Richter, K. Mizuno, P. Radermacher, M. Huber-Lang, T. Seufferlein, S. Paschke, “PKD Regulates Actin Polymerization, Neutrophil Deformability and Transendothelial Migration in Response to fMLP and Trauma,” *J. Leukocyte Biol.*, in press, (2018), [10.1002/JLB.4A0617-251RR](https://doi.org/10.1002/JLB.4A0617-251RR)
3. S. Fujiwara, T. Matsui, K. Ohashi, S. Deguchi, *K. Mizuno, “Solo, a RhoA-targeting Guanine Nucleotide Exchange Factor, is Critical for Hemidesmosome Formation and Acinar Development in Epithelial Cells,” *PLoS One*, **13**, e0195124, (2018), [10.1371/journal.pone.0195124](https://doi.org/10.1371/journal.pone.0195124)
4. R. Nishimura, K. Kato, S. Fujiwara, K. Ohashi, *K. Mizuno, “Solo and Keratin Filaments Regulate Epithelial Tubule Morphology,” *Cell Struct. Funct.*, **43**, 95-105, (2018), [10.1247/csf.18010](https://doi.org/10.1247/csf.18010)
5. T. Nagai, *K. Mizuno, “Jasplakinolide Induces Primary Cilium Formation through Cell Rand YAP Inactivation”, *PLoS ONE*, **12**, e0183030, (2017), [10.1371/journal.pone.0183030](https://doi.org/10.1371/journal.pone.0183030)
6. K. Geng, S. Kumar, S. G. Kimani, V. Kholodovych, C. Kasikara, K. Mizuno, O. Sandiford, P. Rameshwar, S. V. Kotenko, R. B. Birge, “Requirement of Gamma-carboxyglutamic Acid Modification and Phosphatidylserine Binding for the Activation of Tyro3, Axl, and Mertk Receptors by Growth Arrest-specific 6,” *Front. Immunol.* **8**, 1521, (2017), [10.3389/fimmu.2017.01521](https://doi.org/10.3389/fimmu.2017.01521)
7. K. Takahashi, H. Okabe, S. Kanno, T. Nagai, *K. Mizuno, “A Pleckstrin Homology-like Domain is Critical for F-actin Binding and Cofilin-phosphatase Activity of Slingshot-1”, *Biochem. Biophys. Res. Commun.*, **482**, 686-692, (2017), [10.1016/j.bbrc.2016.11.095](https://doi.org/10.1016/j.bbrc.2016.11.095)
8. *K. Ohashi, S. Fujiwara, K. Mizuno, “Roles of the Cytoskeleton, Cell Adhesion and Rho Signaling in Mechanosensing and Mechanotransduction,” *J. Biochem.*, **161**, 245-254, (2017), [10.1093/jb/mvw082](https://doi.org/10.1093/jb/mvw082)

9. R. Morita, M. Kihara, Y. Nakatsu, Y. Nomoto, M. Ogawa, K. Ohashi, K. Mizuno, T. Tachikawa, Y. Ishimoto, Y. Morishita, *T. Tsuji, "Coordination of Cellular Dynamics Contributes to Tooth Epithelium Deformations," *PLoS One*, **11**, e0161336, (2016), [10.1371/journal.pone.0161336](https://doi.org/10.1371/journal.pone.0161336)
10. G. Konotop, E. Bausch, T. Nagai, A. Turchinovich, N. Becker, A. Benner, M. Boutros, K. Mizuno, *A. Krämer, M. S. Raab, "Pharmacological Inhibition of Centrosome Clustering by Slingshot-Mediated Cofilin Activation and Actin Cortex Destabilization," *Cancer Res.*, **76**, 6690-6700, (2016), [10.1158/0008-5472.CAN-16-1144](https://doi.org/10.1158/0008-5472.CAN-16-1144)
11. 藤原佐知子, 大橋一正, 水野健作, “メカノセンシングにおける細胞骨格、細胞接着の機能,” *生化学* **88**, 443-451, (2016)
12. S. Fujiwara, *K. Ohashi, T. Mashiko, H. Kondo, *K. Mizuno, “Interplay between Solo and Keratin Filaments is Crucial for Force-induced Stress Fiber Reinforcement.”, *Mol. Biol. Cell.*, **27**(6), 954-66, (2016), [10.1091/mbc.E15-06-0417](https://doi.org/10.1091/mbc.E15-06-0417)
13. H. Katsuno, M. Toriyama, Y. Hosokawa, K. Mizuno, K. Ikeda, Y. Sakumura, *N. Inagaki, “Actin Migration Driven by Directional Assembly and Disassembly of Membrane Anchored Actin Filaments”, *Cell Reports*, **12**, 648-660, (2015), [10.1016/j.celrep.2015.06.048](https://doi.org/10.1016/j.celrep.2015.06.048)
14. H. Abiko, S. Fujiwara, *K. Ohashi, R. Hiatari, T. Mashiko, N. Sakamoto, M. Sato, *K. Mizuno, “Rho Guanine Nucleotide Exchange Factors Involved in Cyclic-stretch-induced Reorientation of Vascular Endothelial Cells.,” *J. Cell Sci.*, **128**, 1683-1695, (2015), [10.1242/jcs.157503](https://doi.org/10.1242/jcs.157503)
15. Y. Amagai, T. Itoh, M. Fukuda, *K. Mizuno, “Rabin8 Suppresses Autophagosome Formation Independently of its Guanine Nucleotide Exchange Activity Toward Rab8”, *J. Biochem.*, **158**, 139-153, (2015), [10.1093/jb/mvv032](https://doi.org/10.1093/jb/mvv032)
16. H. Katsuno, M. Toriyama, Y. Hosokawa, K. Mizuno, K. Ikeda, Y. Sakumura, *N. Inagaki, “Actin Migration Driven by Directional Assembly and Disassembly of Membrane Anchored Actin Filaments”, *Cell Reports*, **12**, 648-660, (2015), [10.1016/j.celrep.2015.06.048](https://doi.org/10.1016/j.celrep.2015.06.048)
17. K. Ohashi, K. Sampei, M. Nakagawa, N. Uchiumi, T. Amanuma, S. Aiba, M. Oikawa, *K. Mizuno, “Damnacanthal, an Effective Inhibitor of LIM-kinase, Inhibits Cell Migration and Invasion”, *Mol. Biol. Cell*, **25**, 828-840, (2014), [10.1091/mbc.E13-09-0540](https://doi.org/10.1091/mbc.E13-09-0540)
18. T. Nagai, *K. Mizuno, “Multifaceted Roles of Furry Proteins in Invertebrates and Vertebrates”, *J. Biochem.*, **155**, 137-146, (2014), [10.1093/jb/mvu001](https://doi.org/10.1093/jb/mvu001)
19. Y. Homma, S. I. Kanno, K. Sasaki, M. Nishita, A. Yasui, T. Asano, K. Ohashi, *K. Mizuno, “Insulin Receptor Substrate-4 Binds to Slingshot-1 and Promotes Cofilin Dephosphorylation”, *J. Biol. Chem.*, **289**, 26302-26313, (2014), [10.1074/jbc.M114.565945](https://doi.org/10.1074/jbc.M114.565945)
20. T. Oda, S. Chiba, T. Nagai, *K. Mizuno, “Binding to Cep164, but not EB1, is Essential for Centriolar Localization of TTBK2 and its Function in Ciliogenesis”, *Genes Cells*, **19**, 927-940, (2014), [10.1111/gtc.12191](https://doi.org/10.1111/gtc.12191)
21. S. Takahashi, S. Kanno, *K. Mizuno, “Activation of Cytosolic Slingshot-1 phosphatase by Gelsolin-generated Soluble Actin Filaments”, *Biochem. Biophys. Res. Commun.*, **454**, 471-477, (2014), [10.1016/j.bbrc.2014.10.108](https://doi.org/10.1016/j.bbrc.2014.10.108)

【村田和義】

1. Y. Takeichi, T. Uebi, N. Miyazaki, K. Murata, K. Yasuyama, K. Inoue, T. Suzuki, H. Kubo, N. Kajimura, J. Takano, T. Omori, R. Yoshimura, Y. Endo, M.K. Hojo, E. Takaya, S. Kurihara, K. Tatsuta, K. Ozaki, M. Ozaki, “Putative Neural Network within an Olfactory Sensory Unit for Nestmate and Non-nestmate Discrimination in the Japanese Carpenter Ant: The Ultrastructures and Mathematical Simulation”, *Front. Cell. Neurosci.*, in press,(2018)
2. T. Watanabe, C. Song, K. Murata, T. Kureha, D. Suzuki, “Seeded Emulsion Polymerization of Styrene in the Presence of Water-Swollen Hydrogel Microspheres”, *Langmuir*, **34**(29), 8571-8580,(2018), [10.1021/acs.langmuir.8b01047](https://doi.org/10.1021/acs.langmuir.8b01047)
3. 岡本健太, *村田和義「巨大ウイルスの構造解析：クライオ電子顕微鏡の新たな挑戦,」 特集「クライオ電子顕微鏡で見えた生命のかたちとしくみ」, 実験医学 Vol.36 No.8 P.1344-1348 (2018) 羊土社
4. T. Uchihashi, Y. Watanabe, Y. Nakazaki, T. Yamasaki, H. Watanabe, T. Maruno, K. Ishii, S. Uchiyama, C. Song, K. Murata, R. Iino, T. Ando “Dynamic Structural States of ClpB Involved in Its Disaggregation Function”, *Nature Commun.*, **9**, 2147, (2018), [10.1038/s41467-018-04587-w](https://doi.org/10.1038/s41467-018-04587-w)
5. *K. Murata, Y. Kaneko, “Visualization of DNA Compaction in Cyanobacteria by High-voltage Cryo-electron Tomography”, *J. Visual. Exper.*, **137**, e57197, (2018), DOI: 10.3791/57197

6. N. Kobayashi, K. Inano, K. Sasahara, T. Sato, K. Miyazawa, T. Fukuma, M. Hecht, C. Song, K. Murata, R. Arai, "Self-Assembling Supramolecular Nanostructures Constructed from de Novo Extender Protein Nanobuilding Blocks", *ACS Synth Biol.*, **7**, 1381-1394, (2018), [10.1021/acssynbio.8b00007](https://doi.org/10.1021/acssynbio.8b00007)
7. KK. Asare, M. Sakaguchi, AB. Lucky, M. Asada, S. Miyazaki, Y. Katakai, S. Kawai, S. Song, K. Murata, K. Yahata, O. Kaneko, "The Plasmodium Knowlesi MAHRP2 Ortholog Localizes to Structures Connecting Sinton Mulligan's Clefts in the Infected Erythrocyte", *Parasitol Int.*, **67**, 481-492, (2018), [10.1016/j.parint.2018.04.005](https://doi.org/10.1016/j.parint.2018.04.005)
8. *K. Okamoto, N. Miyazaki, HKN. Reddy, MF. Hantke, FRNC. Maia, DSD. Larsson, C. Abergel, JM. Claverie, J. Hajdu, *K. Murata, M. Svenda, "Cryo-EM Structure of a Marseilleviridae Virus Particle Reveals a Large Internal Microassembly", *Virology*, **516**, 239–245, (2018), [10.1016/j.virol.2018.01.021](https://doi.org/10.1016/j.virol.2018.01.021)
9. 宋 致宏, *村田 和義「クライオ電子顕微鏡によるタンパク質の動的構造解析」, *J. Comput. Chem. Japan*, **17**(1) : 38-45, (2018), [10.2477/jccj.2018-0007](https://doi.org/10.2477/jccj.2018-0007)
10. 片山和彦, 芳賀 慧, 藤本 陽, 戸田玲子, 村上耕介, 村田和義, 中西 章「ノロウイルス研究の最新知見」 感染制御と予防衛生 Vol.1 No.1, P.4-11, (2017) メディカルレビュー社
11. A. Nakao, N. Miyazaki, K. Ohira, H. Hagihara, T. Takagi, N. Usuda, S. Ishii, K. Murata, *T. Miyakawa, "Immature Morphological Properties in Subcellular-scale Structures in the Dentate Gyrus of Schnurri-2 Knockout Mice: a Model for Schizophrenia and Intellectual Disability", *Mol Brain*, **10**, 60, (2017), [10.1186/s13041-017-0339-2](https://doi.org/10.1186/s13041-017-0339-2)
12. A. Nakamura, T. Tasaki, Y. Okuni, C. Song, K. Murata, T. Kozai, M. Hara, H. Sugimoto, K. Suzuki, T. Watanabe, T. Uchihashi, H. Noji, *R. Iino, "Rate Constants, Processivity, and Productive Binding Ratio of Chitinase A Revealed by Single-molecule Analysis.", *Phys. Chem. Chem. Phys.*, **20**, 3010-3018, (2017), [10.1039/C7CP04606E](https://doi.org/10.1039/C7CP04606E)
13. *K. Okamoto, N. Miyazaki, C. Song, FRNC. Maia, HKN. Reddy, C. Abergel, J-M. Claverie, J. Hajdu, M. Svenda, *K. Murata, "Structural Variability and Complexity of the Giant Pithovirus Sibericum Particle Revealed by High-Voltage Electron Cryo-tomography and Energy-filtered Electron Cryo-microscopy.", *Sci. Rep.*, **7**, 13291, (2017), [10.1038/s41598-017-13390-4](https://doi.org/10.1038/s41598-017-13390-4)
14. *T. Satoh , C. Song, T. Zhu, T. Toshimori, K. Murata, Y. Hayashi, H. Kamikubo, T. Uchihashi, *K. Kato, "Visualisation of a Flexible Modular Structure of the ER Folding-sensor Enzyme UGGT", *Sci. Rep.*, **7**, 12142, (2017), [10.1038/s41598-017-12283-w](https://doi.org/10.1038/s41598-017-12283-w)
15. *K. Murata, *M. Wolf, "Cryo-electron Microscopy for Structural Analysis of Dynamic Biological Macromolecules.", *Biochim. Biophys. Acta, (BBA), Gen. Subj.*, **1862**, 324-334, (2017), [10.1016/j.bbagen.2017.07.020](https://doi.org/10.1016/j.bbagen.2017.07.020)
16. M. Conley, E. Emmott, R. Orton, D. Taylor, DG. Carneiro, K. Murata, IG. Goodfellow, GS. Hansman, *D. Bhella , "Vesivirus 2117 Capsids More Closely Resemble Sapovirus and Lagovirus Particles than Other Known Vesivirus Structures", *J. Gen. Virol.*, **98**, 68–76, (2017), [10.1099/jgv.0.000658](https://doi.org/10.1099/jgv.0.000658)
17. K. Sai, S. Wang, A. Kaito, T. Fujiwara, T. Maruo, Y. Itoh, M. Miyata, S. Sakakibara, N. Miyazaki, K. Murata, Y. Yamaguchi, T. Haruta, H. Nishioka, Y. Motojima, M. Komura, K. Kimura, *K. Mandai, *Y. Takai, *A. Mizoguchi*, "Multiple Roles of Afadin in the Ultrastructural Morphogenesis of Mouse Hippocampal Mossy Fiber Synapses.", *J. Comp. Neurol.*, **525**, 2719–2734, (2017), [10.1002/cne.24238](https://doi.org/10.1002/cne.24238)
18. C. Song, K. Murata, T. Suzuki, "Intracellular Symbiosis of Algae with Possible Involvement of Mitochondrial Dynamics", *Sci. Rep.*, **7**, 1221, (2017), [10.1038/s41598-017-01331-0](https://doi.org/10.1038/s41598-017-01331-0)
19. K. Murata, Q. Zhang, JG. Galaz-Montoya, C. Fu, ML. Coleman, MS. Osburne, MF. Schmid, MB. Sullivan, SW. Chisholm, W. Chiu, "Visualizing Adsorption of Cyanophage P-SSP7 onto Marine Prochlorococcus", *Sci. Rep.*, **7**, 44176, (2017), [10.1038/srep44176](https://doi.org/10.1038/srep44176)
20. K. Ichimura, S. Kakuta, Y. Kawasaki, T. Miyaki, T. Nonami, N. Miyazaki, T. Nakao, S. Enomoto, S. Arai, M. Koike, K. Murata, T. Sakai, "Morphological Process of Podocyte Development Revealed by Block-face Scanning Electron Microscopy", *J. Cell Sci.*, **130**, 132–142, (2016), [10.1242/jcs.187815](https://doi.org/10.1242/jcs.187815)
21. T. Watanabe, C. Kobayashi, C. Song, K. Murata, T. Kureha, D. Suzuki, "Impact of Spatial Distribution of Charged Groups in Core Poly(N - isopropylacrylamide)-Based Microgels on the Resultant Composite Structures Prepared by Seeded Emulsion Polymerization of Styrene", *Langmuir*, **32**, 12760–12773, (2016), [10.1021/acs.langmuir.6b03172](https://doi.org/10.1021/acs.langmuir.6b03172)
22. *K. Murata, S. Hagiwara, Y. Kimori, *Y. Kaneko, "Ultrastructure of Compacted DNA in Cyanobacteria by High-voltage Cryo-electron Tomography", *Sci. Rep.*, **6**, 34934, (2016), [10.1038/srep34934](https://doi.org/10.1038/srep34934)

23. K. Haga, A. Fujimoto, R. Takai-Todaka, M. Miki, YH. Doan, K. Murakami, M. Yokoyama, K. Murata, A. Nakanishi, K. Katayama, "Functional Receptor Molecules CD300lf and CD300ld Within the CD300 Family Enable Murine Noroviruses to Infect Cells", *Proc. Natl. Acad. Sci. USA*, **113**(41), E6248–E6255, (2016), [10.1073/pnas.1605575113](https://doi.org/10.1073/pnas.1605575113)
24. *K. Okamoto, N. Miyazaki, DS. Larsson, D. Kobayashi, M. Svenda, K. Mühlig, FR. Maia, LH. Gunn, H. Isawa, M. Kobayashi, K. Sawabe, *K. Murata, J. Hajdu, "The Infectious Particle of Insectborne Totivirus-like Omono River Virus has Raised Ridges and Lacks Fibre Complexes", *Sci. Rep.*, **6**, 33170, (2016), [10.1038/srep33170](https://doi.org/10.1038/srep33170)
25. T. Negishi, N. Miyazaki, K. Murata, H. Yasuo, N. Ueno, "Physical Association between a Novel Plasma-membrane Structure and Centrosome Orients Cell Division", *eLife*, **5**, e16550, (2016), [10.7554/eLife.16550](https://doi.org/10.7554/eLife.16550)
26. M. Yamaguchi, H. Yamada, K. Higuchi, Y. Yamamoto, S. Arai, K. Murata, Y. Mori, H. Furukawa, MS, Uddin, H. Chibana, "High-voltage Electron Microscopy Tomography and Structome Analysis of Unique Spiral Bacteria from the Deep Sea", *Microscopy*, **65**, 363–369, (2016), [10.1093/jmicro/dfw016](https://doi.org/10.1093/jmicro/dfw016)
27. M. Takeuchi, I. Karahara, N. Kajimura, A. Takaoka, K. Murata, K. Misaki, S. Yonemura, LA, Staehelin, Y. Mineyuki, "Single Microfilaments Mediate the Early Steps of Microtubule Bundling during Preprophase Band Formation in Onion Cotyledon Epidermal Cells", *Mol. Biol. Cell.*, **27**, 1809–1820, (2016), [10.1091/mbc.E15-12-0820](https://doi.org/10.1091/mbc.E15-12-0820)
28. T. Kaji, K. Kakui, N. Miyazaki, K. Murata, AR Palmer, "Mesoscale Morphology at Nanoscale Resolution: Serial Block-face Scanning Electron Microscopy Reveals Fine 3D Detail of a Novel Silk Spinneret System in a Tube-building Tanaid Crustacean", *Front. Zool.*, **13**, 14, (2016), [10.1186/s12983-016-0146-0](https://doi.org/10.1186/s12983-016-0146-0)
29. *A. Oshima, T. Matsuzawa, K. Murata, K. Tani, Y Fujiyoshi, "Hexadecameric Structure of an Invertebrate Gap Junction Channel", *J. Mol. Biol.*, **428**, 1227–36, (2016), [10.1016/j.jmb.2016.02.011](https://doi.org/10.1016/j.jmb.2016.02.011)
30. C. Kobayashi, T. Watanabe, K. Murata, T. Kureha, *D. Suzuki, "Localization of Polystyrene Particles on the Surface of Poly(N -isopropylacrylamide- co -methacrylic acid) Microgels Prepared by Seeded Emulsion Polymerization of Styrene" *Langmuir* **32**, 1429–1439, (2016), [10.1021/acs.langmuir.5b03698](https://doi.org/10.1021/acs.langmuir.5b03698)
31. *M. Sakaguchi, N. Miyazaki, H. Fujioka, O. Kaneko, *K. Murata, "Three-dimensional Analysis of Morphological Changes in the Malaria Parasite Infected Red Blood Cell by Serial Block-face Scanning Electron Microscopy", *J. Struct. Biol.*, **193**, 162–171, (2016), [10.1016/j.jsb.2016.01.003](https://doi.org/10.1016/j.jsb.2016.01.003)
32. N. Miyazaki, D.W. Taylor, *G.S. Hansman, *K. Murata, "Antigenic and Cryo-electron Microscopy Structure Analysis of a Chimeric Sapovirus Capsid", *J. Virol.*, **90**, 2664–2675, (2015), [10.1128/JVI.02916-15](https://doi.org/10.1128/JVI.02916-15)
33. M. Watanabe, Y. Suzuki, K. Uchida, N. Miyazaki, K. Murata, S. Matsumoto, H. Kakizaki, *M, Tominaga, "Trpm7 Contributes to Intercellular Junction Formation in Mouse Urothelium" *J. Biol. Chem.*, **290**, 29882–92, (2015), [10.1074/jbc.M115.667899](https://doi.org/10.1074/jbc.M115.667899).
34. K.Satoh, K.Takanami, K. Murata, M.Kawata, T.Sakamoto, H.Sakamoto, "Three-dimensional Visualization of Multiple Synapses in Thick Sections Using High-voltage Electron Microscopy in the Rat Spinal Cord", *Data Br.*, **4**, 566–570, (2015), [10.1016/j.dib.2015.07.005](https://doi.org/10.1016/j.dib.2015.07.005)
35. K.Satoh, K.Takanami, K. Murata, M. Kawata, T.Sakamoto, H. Sakamoto, "Effective Synaptome Analysis of Itch-mediating Neurons in the Spinal Cord : A novel Immunohistochemical Methodology Using High-voltage Electron Microscopy", *Neurosci Lett.*, **599**, 86–91, (2015), [10.1016/j.neulet.2015.05.031](https://doi.org/10.1016/j.neulet.2015.05.031)
36. *K. Ichimura, N. Miyazaki, S. Sadayama, K. Murata, M. Koike, K. Nakamura, K. Ohta, T. Sakai, "Three-dimensional Architecture of Podocytes Revealed by Block-face Scanning Electron Microscopy", *Sci. Rep.*, **5**, 8993, (2015), [10.1038/srep08993](https://doi.org/10.1038/srep08993)
37. 宮崎直幸, *村田和義, Serial Block-Face SEM (SBF-SEM) による細胞小器官の 3 次元形態観察 *Plant Morphology* **27**, 9-13, (2015)
38. *K. Murata, M. Esaki, T. Ogura, S. Arai, Y. Yamamoto, N. Tanaka, "Whole-Cell Imaging of the Budding Yeast *Saccharomyces cerevisiae* by High-Voltage Scanning Transmission Electron Tomography", *Ultramicroscopy*, **146**, 39–45, (2014), [10.1016/j.ultramic.2014.05.008](https://doi.org/10.1016/j.ultramic.2014.05.008)
39. N. Miyazaki, M. Esaki, T. Ogura, *K. Murata, "Serial Block-face Scanning Electron Microscopy for Three-dimensional Analysis of Morphological Changes in Mitochondria Regulated by Cdc48p/p97 ATPase", *J. Struct. Biol.*, **187**, 187–193, (2014), [10.1016/j.jsb.2014.05.010](https://doi.org/10.1016/j.jsb.2014.05.010)

40. *M. Yoshioka-Nishimura, D. Nanba, T. Takaki, C. Ohba, N. Tsumura, N. Morita, H. Sakamoto, K. Murata, Y. Yamamoto, "Quality Control of Photosystem II: Direct Imaging of the Changes in the Thylakoid Structure and Distribution of FtsH Proteases in Spinach Chloroplasts under Light Stress", *Plant and Cell Physiology*, **55**(7) 1255-1265, (2014), [10.1093/pcp/pcu079](https://doi.org/10.1093/pcp/pcu079)

【山本量一】

1. C. Shih, J. J. Molina, *R. Yamamoto, "Field-induced Dipolar Attraction between Like-charged Colloids", *Soft Matter*, **12**, 914-924, (2018), [10.1039/C8SM00395E](https://doi.org/10.1039/C8SM00395E)
2. M. Shakeel, A. Hamid, A. Ullah, J. J. Molina, *R. Yamamoto, "Direct Numerical Simulations of Correlated Settling particles", *J. Phys. Soc. Japan*, **87**, 064402, (2018), [10.7566/JPSJ.87.064402](https://doi.org/10.7566/JPSJ.87.064402)
3. M. Tarama, *R. Yamamoto, "Mechanics of cell crawling by means of force-free cyclic motion", *J. Phys. Soc. Japan*, **87**, 044803, (2018), [10.7566/JPSJ.87.044803](https://doi.org/10.7566/JPSJ.87.044803)
4. *R. Yamamoto, J.J. Molina, S.K. Schnyder, 「基板上で遊走・増殖する細胞集団のモデリング」 *J. Comput. Chem. Japan* 17, 14 (2018), [10.2477/jccj.2018-0003](https://doi.org/10.2477/jccj.2018-0003)
5. N. Oyama, J.J. Molina, *R. Yamamoto, "Simulations of model micro-swimmers with fully resolved hydrodynamics", *J. Phys. Soc. Japan*, **86**, 101008, (2017), [10.7566/JPSJ.86.101008](https://doi.org/10.7566/JPSJ.86.101008)
6. N. Oyama, J. J. Molina, *R. Yamamoto, "Do Hydrodynamically Assisted Binary Collisions Lead to Orientational Ordering of Microswimmers?", *Eur. Phys. J. E* **40**, 95 (2017), [10.1140/epje/i2017-11586-4](https://doi.org/10.1140/epje/i2017-11586-4)
7. S. K. Schnyder, Y. Tanaka, J. J. Molina, *R. Yamamoto, "Collective Motion of Cells Crawling on a Substrate: Roles of Cell Shape and Contact Inhibition", *Sci. Rep.*, **7**, 5163, (2017), [10.1038/s41598-017-05321-0](https://doi.org/10.1038/s41598-017-05321-0)
8. N. Oyama, J. J. Molina, *R. Yamamoto, "Simulations of Model Micro-swimmers with Fully Resolved Hydrodynamics," *J. Phys. Soc. Jpn.*, **86**, 101008, (2017), [10.7566/JPSJ.86.101008](https://doi.org/10.7566/JPSJ.86.101008)
9. S. Mehdi, A. Hamid, A. Ullah, *R. Yamamoto, "Microstructure of Rod like Sedimenting Particles: Direct Numerical Simulations," *IBCAST*, 622-626 (2017), [10.1109/IBCAST.2017.7868115](https://doi.org/10.1109/IBCAST.2017.7868115)
10. G. Lericain, R. Yamamoto, U. Hampel, *T. Taniguchi, "Direct Numerical Simulation of a Single Particle Attachment on a Stationary Immersed Bubble," *Phys. Fluids*, **28**, 083301, (2016), [10.1063/1.4960627](https://doi.org/10.1063/1.4960627)

班友

【澤田知久】

1. *T. Sawada, Y. Inomata, M. Yamagami, *M. Fujita, 'Self-Assembly of a Peptide [2]Catenane through Ω -loop Folding,' *Chem. Lett.*, **46**, 1119–1121, (2017), [10.1246/cl.170438](https://doi.org/10.1246/cl.170438)
2. *T. Sawada, M. Yamagami, S. Akinaga, T. Miyaji, *M. Fujita, "Porous Peptide Complexes by a Folding-and-Assembly Strategy", *Chem. Asian J.*, **12**(14), 1715-1718, (2017), [10.1002/asia.201700458](https://doi.org/10.1002/asia.201700458)
3. S. Wang, T. Sawada, *M. Fujita, "Capsule–bowl Conversion Triggered by a Guest Reaction", *Chem. Commun.*, **52**, 11653, (2016), [10.1039/c6cc06551a](https://doi.org/10.1039/c6cc06551a)
4. B. M. Schmidt, T. Osuga, T. Sawada, M. Hoshino, *M. Fujita, "Compressed Corannulene in a Molecular Cage", *Angew. Chem. Int. Ed.*, **55**, 1561, (2016), [10.1002/anie.201509963](https://doi.org/10.1002/anie.201509963)
5. S. Wang, T. Sawada, K. Ohara, K. Yamaguchi, *M. Fujita, "Capsule–Capsule Conversion by Guest Encapsulation," *Angew. Chem. Int. Ed.*, **55**, 2063, (2016), [10.1002/anie.201509278](https://doi.org/10.1002/anie.201509278)
6. *T. Sawada, M. Yamagami, K. Ohara, K. Yamaguchi, *M. Fujita, 'Peptide [4]Catenane via Folding and Assembly,' *Angew. Chem. Int. Ed.*, **55**, 4519–4522, (2016), [10.1002/anie.201600480](https://doi.org/10.1002/anie.201600480)
7. *T. Sawada, A. Matsumoto, *M. Fujita, "Coordination-Driven Folding and Assembly of a Short Peptide into a Protein-like Two-Nanometer-Sized Channel" *Angew. Chem. Int. Ed.*, **53** (28), 7228-7232, (2014), [10.1002/anie.201403506](https://doi.org/10.1002/anie.201403506)
8. T. Sawada, H. Hisada, *M. Fujita, "Mutual Induced Fit in a Synthetic Host–Guest System", *J. Am. Chem. Soc.*, **136** (12), 4449-4451, (2014), [10.1021/ja500376x](https://doi.org/10.1021/ja500376x)

【重田育照】

1. *R. Harada, H. Aida, *Y. Shigeta, "The Formation of Hydrophobic Core Regulates the Protein Folding of Villin Elucidated with Parallel Cascade Selection Molecular Dynamics", *Chem. Lett.*, in press, (2018)
2. *R. Harada, *Y. Shigeta, "How do Rules for Selecting Initial Structures in Parallel Cascade Selection Molecular Dynamics Affect its Conformational Sampling Efficiency?", *Chem. Lett.*, in press, (2018), [10.1246/cl.180464](https://doi.org/10.1246/cl.180464)

3. *R. Harada, *Y. Shigeta, "Temperature Shuffled Structural Dissimilarity Sampling Based on a Root-Mean Square Deviation", *J. Chem. Inf. Model.*, **58**(7), 1397-1405, (2018), 10.1021/acs.jcim.8b00095
4. *R. Harada, *Y. Shigeta, "How Low-resolution Data Can Predict Conformational Changes of a Protein: a Molecular Dynamics Study", *Phys. Chem. Chem. Phys.*, **20**, 17790-17798, (2018), 10.1039/C8CP02246A
5. *R. Harada, *Y. Shigeta, "On-the-Fly Specifications of Reaction Coordinates in Parallel Cascade Selection Molecular Dynamics Accelerate Conformational Transitions of Proteins", *J. Chem. Theor. Comput.*, **14**, 3332-3341, (2018), 10.1021/acs.jctc.8b00264
6. *R. Harada, T. Mashiko, M. Tachikawa, S. Hiraoka, *Y. Shigeta, "Programmed Dynamical Ordering in Self-organization Processes of Nano-cube: A Molecular Dynamics Study", *Phys. Chem. Chem. Phys.*, **20**, 9115-9122, (2018), 10.1039/C8CP00284C
7. R. Sato*, R. Harada, Y. Shigeta*, "The binding structure and affinity of photodamaged duplex DNA with members of the photolyase/cryptochrome family: A computational study", *Biophysics and Physicobiology* **15**, 18-27 (2018), 10.2142/biophysico.15.0_18
8. R. Harada*, Y. Shigeta*, "Self-Avoiding Conformational Sampling (SACS) Based on Histories of Past Conformational Searches", *J. Chem. Inf. Model.*, **57**, 3070-3078, (2017), 10.1021/acs.jcim.7b00573
9. Y. Sasano, R. Sato, Y. Shigeta, N. Yasuda, H. Maeda*, "H-aggregated π-systems based on disulfide-linked dimers of dipyrrolyldiketone boron complexes", *J. Org. Chem.*, **82**, 11166-11172, (2017), 10.1021/acs.joc.7b02185
10. R. Harada*, Y. Shigeta, "An Assessment of Optimal Time Scale of Conformational Resampling in Parallel Cascade Molecular Dynamics", *Molecular Simulation*, in press, (2017), [10.1080/08927022.2017.1362696](https://doi.org/10.1080/08927022.2017.1362696)
11. R. Harada*, Y. Shigeta, "Dynamic Specification of Initial Structures in Parallel Cascade Selection Molecular Dynamics (PaCS-MD) Efficiently Promotes Biologically Relevant Rare Events", *Bull. Chem. Soc. Jap.*, **90**, 1236-1243, (2017), [10.1246/bcsj.20170177](https://doi.org/10.1246/bcsj.20170177)
12. R. Harada*, Y. Shigeta, "Temperature-shuffled Parallel Cascade Selection Molecular Dynamics Accelerates the Structural Transitions of Proteins", *J. Comput. Chem., (Rapid Communication)*, **38**, 2671-2674, (2017), [10.1002/jcc.25060](https://doi.org/10.1002/jcc.25060)
13. R. Harada*, Y. Shigeta, "Structural Dissimilarity Sampling with Dynamically Self-Guiding Selection", *J. Comput. Chem.*, **38**, 1921-1929, (2017), [10.1002/jcc.24837](https://doi.org/10.1002/jcc.24837)
14. Y. Shigeta*, R. Harada, R. Sato, H. Kitoh-Nishioka, T. K. M. Bui, A. Sato, A. Kyan, Y. Ishii, M. Kimatsuka, S. Yamasaki, M. Kayanuma, M. Shoji, "Classical Cumulant Dynamics for Statistical Chemical Physics", *Molecular Simulation*, **43**(13-16), 1260-1268, (2017), [10.1080/08927022.2017.1315770](https://doi.org/10.1080/08927022.2017.1315770)
15. J. Fujita, R. Harada, Y. Maeda, Y. Saito, E. Mizohata, T. Inoue, Y. Shigeta*, H. Matsumura*, "Identification of the Key Interactions in Structural Transition Pathway of FtsZ from Staphylococcus Aureus", *J. Struct. Biol.*, **198**, 65-73, (2017), [10.1016/j.jsb.2017.04.008](https://doi.org/10.1016/j.jsb.2017.04.008)
16. R. Harada*, Y. Shigeta, "How does the Number of Initial Structures Affect the Conformational Sampling Efficiency and Quality in Parallel Cascade Selection Molecular Dynamics (PaCS-MD)?", *Chem. Lett.*, **46**, 862-865, (2017), [10.1246/cl.170207](https://doi.org/10.1246/cl.170207)
17. R. Harada*, Y. Shigeta, "Efficient Conformational Search Based on Structural Dissimilarity Sampling: Applications to Reproductions of Structural Transitions on Maltodextrin Binding Protein", *J. Chem. Theory Comput.*, **13**, 1411-1423, (2017), [10.1021/acs.jctc.6b01112](https://doi.org/10.1021/acs.jctc.6b01112)
18. R. Yamakado, Y. Ashida, R. Sato, Y. Shigeta, N. Yasuda, H. Maeda*, "Cooperatively Interlocked [2+1]-Type π-System-Anion Complexes", *Chemistry A European Journal*, **23**, 4160-4168, (2017), [10.1002/chem.201605765](https://doi.org/10.1002/chem.201605765)
19. R. Harada*, Y. Takano, Y. Shigeta, "Common Folding Processes of Mini Proteins: Partial Formations of Secondary Structures Initiate the Immediate Protein Folding", *J. Comput. Chem.*, **38**, 790-797, (2017), [10.1002/jcc.24748](https://doi.org/10.1002/jcc.24748)
20. *R. Sato, R. Harada, Y. Shigeta, "Theoretical Analyses on a Flipping Mechanism of UV-Induced DNA Damage", *Biophysics and Physicobiology*, **13**, 311-319, (2016), [10.2142/biophysico.13.0_311](https://doi.org/10.2142/biophysico.13.0_311)
21. W. Tanaka, *M. Shoji, F. Tomoike, Y. Ujiie, K. Hanaoka, R. Harada, M. Kayanuma, K. Kamiya, T. Ishida, R. Masui, S. Kuramitsu, Y. Shigeta, "Molecular Mechanisms of Substrate Specificities of Uridine-Cytidine Kinase", *Biophysics and Physicobiology*, **13**, 77-84, (2016), [10.2142/biophysico.13.0_77](https://doi.org/10.2142/biophysico.13.0_77)

22. *R. Harada, T. Nakamura, Y. Shigeta, “A Fast Convergent Simulated Annealing Algorithm: Simulated Annealing Outlier FLOODing (SA-OFLOOD) Method”, *Bull. Chem. Soc. Jap.*, **89**, 1361-1367, (2016), [10.1246/bcsj.20160244](https://doi.org/10.1246/bcsj.20160244)
23. R. Yamakado, R. Sato, Y. Shigeta, *H. Maeda, “Ion-Pairing Crystal Polymorphs of Interlocked [2+1]-type Receptor–Anion Complexes”, *J. Org. Chem.*, **81**, 8530–8536, (2016), [10.1021/acs.joc.6b01688](https://doi.org/10.1021/acs.joc.6b01688)
24. *S. Negoro, Y. Kawashima, N. Shibata, T. Kobayashi, T. Baba, Y.-H. Lee, K. Kamiya, Y. Shigeta, K. Nagai, I. Takehara, D.-I. Kato, M. Takeo, Y. Higuchi, “Mutations Affecting the Internal Equilibrium of the 6-aminohexanoate-dimer Hydrolase Reaction”, *FEBS Letter*, **590**, 3133-3143, (2016), [10.1002/1873-3468.12354](https://doi.org/10.1002/1873-3468.12354)
25. W. Naito, N. Yasuda, T. Morimoto, Y. Shigeta, H. Takaya, I. Hisaki, *H. Maeda, “Doubly N-Methylated Porphyrinoids”, *Org. Lett.*, **18**, 3006-3009, (2016), [10.1021/acs.orglett.6b01377](https://doi.org/10.1021/acs.orglett.6b01377)
26. *R. Harada, Y. Takano, Y. Shigeta, “TaBoo SeArch (TBSA) Algorithm with a Modified Inverse Histogram for Reproducing Biologically Relevant Rare-events of Proteins”, *J. Chem. Theory Comput.*, **12**(5), 2436-2445, (2016), [10.1021/acs.jctc.6b00082](https://doi.org/10.1021/acs.jctc.6b00082)
27. M. Kayanuma, M. Shoji, M. Yoda, M. Odaka, Y. Shigeta, “Catalytic Mechanism of Nitrile Hydratase Subsequent to Cyclic Intermediate Formation: A QM/MM Study”, *J. Phys. Chem. B*, **120**, (13), 3259–3266, (2016), [10.1021/acs.jpcb.5b11363](https://doi.org/10.1021/acs.jpcb.5b11363)
28. R. Harada, T. Nakamura, Y. Shigeta, “Sparsity-weighted Outlier FLOODing (OFLOOD) Method: Efficient Rare Event Sampling Method Using Sparsity of Distribution”, *J. Comput. Chem.*, **37**, 724–738, (2016), [10.1002/jcc.24255](https://doi.org/10.1002/jcc.24255)
29. R. Harada, Y. Inagaki, Y. Shigeta, “Protein Folding and Evolution” (**invited review**), *Reference modules in Materials Science and Engineering 2016, Article ID: Protein Folding and Evolution Elsevier*, (2016), [10.1016/B978-0-12-803581-8.09800-3](https://doi.org/10.1016/B978-0-12-803581-8.09800-3)
30. R. Harada, *T. Nakamura, *Y. Shigeta, “Automatic Detection of Hidden Dimension in Outlier FLOODing (OFLOOD) Method”, *Chem. Phys. Lett.*, **639**, 269-274, (2015), [10.1016/j.cplett.2015.09.031](https://doi.org/10.1016/j.cplett.2015.09.031)
31. *M. Shoji, M. Kayanuma, H. Umeda, Y. Shigeta, “Performance of the Divide-and-conquer Approach Used as an Initial Guess”, *Chem. Phys. Lett.*, **634**, 181–187, (2015), [10.1016/j.cplett.2015.06.011](https://doi.org/10.1016/j.cplett.2015.06.011)
32. *R. Harada, *Y. Takano, *Y. Shigeta, “Efficient Conformational Sampling of Proteins Based on a Multi-dimensional Inverse Histogram: An Application to Folding of Chignolin in Explicit Solvent”, *Chem. Phys. Lett.*, **630**, 68-75, (2015), [10.1016/j.cplett.2015.04.039](https://doi.org/10.1016/j.cplett.2015.04.039)
33. *R. Harada, T. Nakamura, Y.Takano, Y. Shigeta, “Protein Folding Pathways Extracted by OFLOOD: Outlier FLOODing Method”, *J. Comput. Chem.*, **36**, 97-102, (2015), [10.1002/jcc.23773](https://doi.org/10.1002/jcc.23773)
34. S. Maekawa, T. Matsui, K. Hirao, *Y. Shigeta, “A Theoretical Study on Reaction Mechanisms of Nitrite Reduction in Copper Nitrite Complexes as Models for the Copper Nitrite Reductase”, *J. Phys. Chem. B*, **119**, 5392–5403, (2015), [10.1021/acs.jpcb.5b01356](https://doi.org/10.1021/acs.jpcb.5b01356)
35. K. Okuno, Y. Shigeta, R. Kishi, *M. Nakano, “Theoretical Design of Solvatochromism Switching by Photochromic Reactions Using Donor-acceptor Disubstituted Diarylethene Derivatives with Oxidized Thiophene Rings”, *Phys. Chem. Chem. Phys.*, **17**, 6484-6494, (2015), [10.1039/C4CP05946H](https://doi.org/10.1039/C4CP05946H)
36. R. Nakamura, *Y. Shigeta, K. Okuno, M. Fukushima, M. Hasegawa, S. Suzuki, M. Kozaki, K. Okada, M. Nakano, “Substitution Effects on Optical Properties of Iminonitroxide-substituted Iminonitroxide Diradical”, *Mol. Phys.*, **113**, 267-273, (2015), [10.1080/00268976.2014.937777](https://doi.org/10.1080/00268976.2014.937777)
37. *R. Harada, *Y. Takano, *Y. Shigeta, “Enhanced Conformational Sampling Method for Proteins Based on the TaBoo SeArch (TBSA) Algorithm: Application to the Folding of a Mini-protein, Chignolin”, *J. Comput. Chem.*, **36**, 763-772, (2015), [10.1002/jcc.23854](https://doi.org/10.1002/jcc.23854)
38. *T. Baba, M. Boero, K. Kamiya, H. Ando, S. Negoro, M. Nakano, *Y. Shigeta, “Unraveling the Degradation of Artificial Amide Bonds in Nylon Oligomer Hydrolase: from Induced-fit to Acylation Processes”, *Phys. Chem. Chem. Phys.*, **17**, 4492-4504, (2015), [10.1039/C4CP04419C](https://doi.org/10.1039/C4CP04419C)
39. *T. Matsui, Y. Kitagawa, M. Okumura, *Y. Shigeta, “Accurate Standard Hydrogen Electrode Potential and Applications to the Redox Potentials of Vitamin C and NAD/NADH”, *J. Phys. Chem. A*, **119**, 369-376, (2015), [10.1021/jp508308y](https://doi.org/10.1021/jp508308y)
40. H. Harada, Y. Takano, T. Baba, Y. Shigeta, “Simple, Yet Powerful Methodologies for Conformational Sampling of Proteins”, *Phys. Chem. Chem. Phys.*, **17**, 6155-6173, (2015), [10.1039/C4CP05262E](https://doi.org/10.1039/C4CP05262E)

41. Y. Shigeta, H. Harada, M. Kayanuma, M. Shoji, “Quantal Cumulant Dynamics for Real-time Simulations of Quantum Many-body Systems”, *Int. J. Quant. Chem.*, **115**, 300-308, (2015), [10.1002/qua.24820](https://doi.org/10.1002/qua.24820)
42. T. Baba, T. Matsui, K. Kamiya, M. Nakano, *Y. Shigeta, “A Density Functional Study on pKa of Small Polyprotic Molecules”, *Int. J. Quant. Chem.*, **114**, 1128–1134, (2014), [10.1002/qua.24631](https://doi.org/10.1002/qua.24631)
43. *K. Kamiya, T. Baba, M. Boero, T. Matsui, S. Negoro, Y. Shigeta, “A Nylon-oligomer Hydrolase Promoting Cleavage Reactions in Unnatural Amide Compounds”, *J. Phys. Chem. Lett.*, **5**, 1210-1216, (2014), [10.1021/jz500323y](https://doi.org/10.1021/jz500323y)
44. *R. Harada, Y. Takano, Y. Shigeta, “Fluctuation Flooding Method (FFM) for Accelerating Conformational Transitions of Proteins”, *J. Chem. Phys.*, **140**, 125103, (2014), [10.1063/1.4869594](https://doi.org/10.1063/1.4869594)
45. T. Baba, R. Harada, M. Nakano, *Y. Shigeta, “On the Induced-fitmechanism of Substrate-enzyme Binding Structures of Nylon-oligomer Hydrolase”, *J. J. Comput. Chem.*, **35**, 1240-1247, (2014), [10.1002/jcc.23614](https://doi.org/10.1002/jcc.23614)
46. H. Ando, *Y. Shigeta, T. Baba, C. Watanabe, Y. Okiyama, Y. Mochizuki, M. Nakano, “Hydration Effects on Enzyme-Substrate Complex of Nylon Oligomer Hydrolase: Inter-Fragment Interaction Energy Study by the Fragment Molecular Orbital Method”, *Mol. Phys.*, **113**, 319-326, (2014), [10.1080/00268976.2014.941311](https://doi.org/10.1080/00268976.2014.941311)

【塚崎智也】

1. A. Furukawa, S. Nakayama, K. Yoshikai, Y. Tanaka, *T. Tsukazaki. “Remote Coupled Drastic β -Barrel to β -Sheet Transition of the Protein Translocation Motor,” *Structure* **26**, 485–489, (2018), [10.1016/j.str.2018.01.002](https://doi.org/10.1016/j.str.2018.01.002)
2. *T. Tsukazaki, “Structure-based Working Model of SecDF, a Proton-driven Bacterial Protein Translocation Factor,” *FEMS Microbiology Letters*, **365**, fny112, (2018), [10.1093/femsle/fny112](https://doi.org/10.1093/femsle/fny112)
3. Y. Sugano, A. Furukawa, O. Nureki, Y. Tanaka, *T. Tsukazaki, ‘SecY-SecA Fusion Protein Retains the Ability to Mediate Protein Transport,’ *PLoS One*, **12**, e0183434, (2017), [10.1371/journal.pone.0183434](https://doi.org/10.1371/journal.pone.0183434)
4. Y. Tanaka, S. Iwaki, *T. Tsukazaki, ‘Crystal Structure of a Plant Multidrug and Toxic Compound Extrusion Family Protein,’ *Structure*, **25**, 1455-1460, (2017), [10.1016/j.str.2017.07.009](https://doi.org/10.1016/j.str.2017.07.009)
5. Y. Daimon, C. Iwama-Masui, Y. Tanaka, T. Shiota, T. Suzuki, R. Miyazaki, H. Sakurada, T. Lithgow, N. Dohmae, H. Mori, *T. Tsukazaki, *S. Narita, *Y Akiyama, ‘The TPR Domain of BepA is Required for Productive Interaction with Substrate Proteins and the β -barrel Assembly Machinery Complex,’ *Mol. Microbiol.*, **106**, 760-776, (2017), [10.1111/mmi.13844](https://doi.org/10.1111/mmi.13844)
6. A. Furukawa, K. Yoshikai, T. Mori, H. Mori, V.Y. Morimoto, Y. Sugano, S. Iwaki, T. Minamino, Y. Sugita, Y. Tanaka, T. Tsukazaki, ‘Tunnel Formation Inferred from the I-Form Structures of the Proton-Driven Protein Secretion Motor SecDF,’ *Cell Rep.*, **9**(5), 895-901, (2017), [10.1016/j.celrep.2017.04.030](https://doi.org/10.1016/j.celrep.2017.04.030)
7. Y. Tanaka, Y. Sugano, M. Takemoto, T. Mori, A. Furukawa, T. Kusakizako, K. Kumazaki, A. Kashima, R. Ishitani, Y. Sugita, *O. Nureki, *T. Tsukazaki, ‘Crystal Structures of SecYEG in Lipidic Cubic Phase Elucidate a Precise Resting and a Peptide-Bound State’, *Cell Rep.*, **13**, 1561-1568, (2015), [10.1016/j.celrep.2015.10.025](https://doi.org/10.1016/j.celrep.2015.10.025)
8. N. Shimokawa-Chiba, K. Kumazaki, T. Tsukazaki, O. Nureki, K. Ito, *S. Chiba, ‘Hydrophilic Microenvironment Required for the Channel-independent Insertase Function of YidC Protein’, *Proc. Natl. Acad. Sci. USA*, **112**, 5063-8, (2015), [10.1073/pnas.1423817112](https://doi.org/10.1073/pnas.1423817112)
9. K. Kumazaki, S. Chiba, M. Takemoto, A. Furukawa, K. Nishiyama, Y. Sugano, T. Mori, N. Dohmae, K. Hirata, Y. Nakada-Nakura, AD. Maturana, Y. Tanaka, H. Mori, Y. Sugita, F. Arisaka, K. Ito, R. Ishitani, *T. Tsukazaki, *O. Nureki, ‘Structural Basis of Sec-independent Membrane Protein Insertion by YidC.’, *Nature*, **509**, 516-520, (2014), [10.1038/nature13167](https://doi.org/10.1038/nature13167)
10. K. Kumazaki, *T. Tsukazaki, T. Nishizawa, Y. Tanaka, HE. Kato, K. Hirata, Y. Nakada-Nakura, Y. Mori , H. Suga, N. Dohmae, R. Ishitani, *O. Nureki, ‘Crystallization and Preliminary X-ray Diffraction Analysis of YidC, a Membrane Chaperone/insertase from *Bacillus halodurans*’, *Acta Crystallogr. F*, **70**, 1056-1060, (2014), [10.1107/S2053230X14012540](https://doi.org/10.1107/S2053230X14012540)
11. K. Kumazaki, T. Kishimoto, A. Furukawa, H. Mori, Y. Tanaka, N. Dohmae, R. Ishitani, *T. Tsukazaki, O. Nureki, ‘Crystal Structure of Escherichia coli YidC, a Membrane Protein Chaperone and Insertase’, *Sci Rep.*, **4**, 7299, (2014), [10.1038/srep07299](https://doi.org/10.1038/srep07299)

【長田裕也】

1. Y. Nagata, * T. Nishikawa, * M. Suginome, ‘Abnormal Sergeants-and-Soldiers Effect of

- Poly(quinoxaline-2,3-diyl)s Enabling Discrimination of One-Carbon Homologous *n*-Alkanes through a Highly Sensitive Solvent-dependent Helix Inversion” *Chem. Comm.* **54**, 6867-6870, (2018), 10.1039/C8CC02836B
2. *Y. Nagata, Y. Shimada, T. Nishikawa, R. Takeda, M. Uno, *T. Ogoshi, *M. Sugino, “A Planar-Chiral Pillar[5]arene-Based Monophosphine Ligand with Induced Chirality at the Biaryl Axis”, *Synlett*, in press, (2018), 10.1055/s-0037-1610635
 3. Y. Nagata, * T. Nishikawa, M. Sugino, * S. Sato, M. Sugiyama, * L. Porcar, A. Martel, R. Inoue, N. Sato, “Elucidating the Solvent Effect on the Switch of the Heliicity of Poly(quinoxaline-2,3-diyl)s: A Conformational Analysis by Small-Angle Neutron Scattering”, *J. Am. Chem. Soc.* **140**, 2722, (2018), 10.1021/jacs.7b11626
 4. *T. Miura, T. Nakamuro, S. G. Stewart, Y. Nagata, *M. Murakami, “Synthesis of Enantiopure C3-Symmetric Triangular Molecules”, *Angew. Chem. Int. Ed.* **56**, 3334, (2017), 10.1002/anie.201612585
 5. F. K. C. Leung, F. Ishiwari, Y. Shoji, T. Nishikawa, R. Takeda, Y. Nagata, M. Sugino, Y. Uozumi, *Y. M. A. Yamada, *T. Fukushima, “Synthesis and Catalytic Applications of a Triptycene-Based Monophosphine Ligand for Palladium-Mediated Organic Transformations”, *ACS Omega* **2**, 1930, (2017), 10.1021/acsomega.7b00200
 6. H. Hasegawa, *Y. Nagata, *K. Terao, M. Sugino, “Synthesis and Solution Properties of a Rigid Helical Star Polymer: Three-Arm Star Poly(quinoxaline-2,3-diyl)”, *Macromolecules* **50**, 7491, (2017), 10.1021/acs.macromol.7b01797
 7. T. Nishikawa, Y. Nagata, M. Sugino, “Poly(quinoxaline-2,3-diyl) as a Multifunctional Chiral Scaffold for Circularly Polarized Luminescent Materials: Color Tuning, Energy Transfer, and Switching of the CPL Handedness”, *ACS Macro Lett.* **6**, 431, (2017), 10.1021/acsmacrolett.7b00131
 8. Y. Nagata, R. Takeda, M. Sugino, “High-Pressure Circular Dichroism Spectroscopy up to 400 MPa Using Polycrystalline Yttrium Aluminum Garnet (YAG) as Pressure-Resistant Optical Windows”, *RSC Adv.* **6**, 109726, (2016), 10.1039/C6RA23736C
 9. Y. Nagata, M. Uno, M. Sugino, “Three-Way-Switchable (Right/Left/OFF) Selective Reflection of Circularly Polarized Light on Solid Thin Films of Helical Polymer Blends”, *Angew. Chem., Int. Ed.* **55**, 7126, (2016), 10.1002/anie.201602035
 10. Y. Nagata, T. Nishikawa, M. Sugino, “Solvent Effect on the Sergeants-and-Soldiers Effect Leading to Bidirectional Induction of Single-Handed Helical Sense of Poly(quinoxaline-2,3-diyl)s Copolymers in Aromatic Solvents”, *ACS Macro Lett.* **5**, 519, (2016), 10.1021/acsmacrolett.6b00191

【濱田大三】

1. Y. Matsushita, H. Sekiguchi, J.W. Chang, M. Nishijima, K. Ikezaki, D. Hamada, Y. Goto, *Y.C. Sasaki, “Nanoscale Dynamics of Protein Assembly Networks in Supersaturated Solutions..”, *Sci. Rep.* **7**, 13883, (2017), 10.1038/s41598-017-14022-7
2. M. Nawata, H. Tsutsumi, Y. Kobayashi, S. Unzai, S. Mine, T. Nakamura, K. Uegaki, H. Kamikubo, M. Kataoka, *D. Hamada, “Heat-induced Native Dimerization Prevents Amyloid Formation by Variable Domain from Immunoglobulin Light-chain REI”, *FEBS J.* **284**, 3114-3127, (2017), 10.1111/febs.14181
3. K. Kuroki, K. Mio, A. Takahashi, H. Matsubara, Y. Kasai, S. Manaka, M. Kikkawa, D. Hamada, C. Sato, *K. Maenaka. “Cutting Edge: Class II-like Structural Features and Strong Receptor Binding of the Nonclassical HLA-G2 Isoform Homodimer”, *J Immunol.* **198**, 3399-3403, (2017), 10.4049/jimmunol.1601296
4. B. Wang, M. Nishimura, H. Tang, A. Kawabata, N. F. Mahmoud, Z. Khanlari, D. Hamada, H. Tsuruta, *Y. Mori, “Crystal Structure of Human Herpesvirus 6B Tegument Protein U14”, *PLoS Pathog.* **12**, e1005594, (2016), 10.1371/journal.ppat.1005594

【原野幸治】

1. *Y. Zhen, K. Inoue, Z. Wang, T. Kusamoto, K. Nakabayashi, S. Ohkoshi, W. Hu, *Y. Guo, *K. Harano, *E. Nakamura, “Acid-Responsive Conductive Nanofiber of Tetrabenzoporphyrin Made by Solution Processing” *J. Am. Chem. Soc.* **140**, 62-65, (2018), 10.1021/jacs.7b10575
2. S. Kai, S. P. Maddala, T. Kojima, S. Akagi, K. Harano, E. Nakamura, *S. Hiraoka, “Flexibility of Components Alters the Self-assembly Pathway of Pd₂L₄ Coordination Cages” *Dalton Trans.* **47**, 3258-3263, (2018) 10.1039/C8DT00112J
3. S. Okada, S. Kowashi, L. Schweighauser, K. Yamanouchi, *K. Harano, *E. Nakamura, “Direct

- Microscopic Analysis of Individual C₆₀ Dimerization Events: Kinetics and Mechanisms” *J. Am. Chem. Soc.* **139**, 18281–18287 (2017), [10.1021/jacs.7b09776](https://doi.org/10.1021/jacs.7b09776)
4. R. Aoki, R. Toyoda, J. F. Kögel, *R. Sakamoto, J. Kumar, Y. Kitagawa, K. Harano, T. Kawai, *H. Nishihara, “Bis(dipyrrinato)zinc(II) Complex Chiroptical Wires: Exfoliation into Single Strands and Intensification of Circularly Polarized Luminescence,” *J. Am. Chem. Soc.*, **139**, 16024–16027, (2017), [10.1021/jacs.7b07077](https://doi.org/10.1021/jacs.7b07077)
 5. *原野幸治, “お茶のなかの分子世界”, 現代化学, **556**, 42–45, (2017)
 6. L. Schweighauser, *K. Harano, *E. Nakamura, “Experimental Study on Interconversion between Cubic MOF-5 and Square MOF-2 Arrays,” *Inorg. Chem. Commun.*, **84**, 1–4, (2017), [10.1016/j.inoche.2017.07.009](https://doi.org/10.1016/j.inoche.2017.07.009)
 7. H. Nitta, *K. Harano, M. Isomura, E. H. G. Backus, M. Bonn, *E. Nakamura, “Conical Ionic Amphiphiles Endowed with Micellization Ability but Lacking Air- and Oil-Water Interfacial Activity,” *J. Am. Chem. Soc.*, **139**, 7677–7680, (2017), [10.1021/jacs.7b01596](https://doi.org/10.1021/jacs.7b01596)
 8. M. Yamada, N. Yoshinari, N. Kuwamura, T. Saito, S. Okada, S. P. Maddala, K. Harano, E. Nakamura, K. Yamagami, K. Yamanaka, A. Sekiyama, T. Suenobu, Y. Yamada, *T. Konno, “Heterogeneous Catalase-like Activity of Gold(I)-Cobalt(III) Metallocsupramolecular Ionic Crystals”, *Chem. Sci.*, **8**, 2671–2676, (2017), [10.1039/C6SC04993A](https://doi.org/10.1039/C6SC04993A)
 9. P. Bairi, *K. Minami, J. P. Hill, W. Nakanishi, *L. K. Shrestha, C. Liu, K. Harano, E. Nakamura, *K. Ariga, “Supramolecular Differentiation for Constructing Anisotropic Fullerene Nanostructures by Time-Programmed Control of Interfacial Growth,” *ACS Nano*, **10**, 8796–8802, (2016), [10.1021/acsnano.6b04535](https://doi.org/10.1021/acsnano.6b04535)
 10. R. M. Gorgoll, *K. Harano, *E. Nakamura, “Nanoscale Control of Polymer Assembly on a Synthetic Catalyst-Bilayer System,” *J. Am. Chem. Soc.*, **138**, 9675–9681, (2016), [10.1021/jacs.6b05414](https://doi.org/10.1021/jacs.6b05414)
- 【前田大光】
1. A. Kuno, M. Fujiwara, Y. Haketa, *H. Maeda, “Arylpyrrolyldiketone Boron Complexes Exhibiting Various Anion-Binding Modes Based on Dynamic Conformation Changes” *Chem. Asian J.* **14**, in press (2018), [10.1002/asia.201801040](https://doi.org/10.1002/asia.201801040)
 2. *H. Maeda, T. Okubo, Y. Haketa, N. Yasuda, ”Pyrrole-Based Zwitterionic π-Electronic Systems That Form Self-Assembled Dimers” *Chem. Eur. J.*, **24**, in press, (2018), [10.1002/chem.201803796](https://doi.org/10.1002/chem.201803796)
 3. K. Kaneko, M. Goto, Y. Haketa, *H. Maeda, T. Hanasaki, “Induced Homeotropic Alignment of Nematic Liquid Crystals by Doping Side-on Carbosilane-Based Oligomers” *Chem. Lett.* **47**, in press, (2018), [10.1246/cl.180509](https://doi.org/10.1246/cl.180509)
 4. S. Kaname, Y. Haketa, N. Yasuda, *H. Maeda, “Cyclic Anion-Responsive π-Electronic Molecules That Overcome Energy Losses Induced by Conformation Changes,” *Org. Lett.* **20** (11), 3268–3272 (2018), [10.1021/acs.orglett.8b01138](https://doi.org/10.1021/acs.orglett.8b01138)
 5. G. Hirata, *H. Maeda, Pyrrole-Based Anion-Responsive π-Electronic Molecules as Hydrogen-Bonding Catalysts,” *Org. Lett.* **20** (10), 2853–2856, (2018), [10.1021/acs.orglett.8b00855](https://doi.org/10.1021/acs.orglett.8b00855)
 6. *H. Maeda, Y. Takeda, Y. Haketa, Y. Morimoto, N. Yasuda, “Ion-Pairing Assemblies of π-Electronic Anions Formed by Intramolecular Hydrogen Bonding,” *Chem. Eur. J.* **24**(35), 8910–8916, (2018), [10.1002/chem.201801375](https://doi.org/10.1002/chem.201801375)
 7. N. Oka, *F. Ito, Y. Haketa, *H. Maeda, T. Miyano, N. Tohnai, S. Ito, H. Miyasaka, S. Ozeki, “Dynamic Polymorph Formation During Evaporative Crystallization from Solution: The Key Role of Liquid-like Clusters as "Crucible" at Ambient Temperature,” *Chem. Eur. J.* **24**(17), 4343–4349, (2018), [10.1002/chem.201705356](https://doi.org/10.1002/chem.201705356)
 8. Y. Haketa, *H. Maeda, “Dimension-Controlled π-Electronic Ion-Pairing Assemblies,” *Bull. Chem. Soc. Jpn.* **91**(3), 420–436, (2018), [10.1246/bcsj.20170434](https://doi.org/10.1246/bcsj.20170434)
 9. R. Yamakado, M. Hara, S. Nagano, T. Seki, *H. Maeda, “Complexation of Anion-Responsive π-Electronic System with Alkyl-Substituted Azobenzene Carboxylate Providing Ion-Pairing Assemblies,” *Chem. Lett.* **47**(4), 404–407, (2018), [10.1246/cl.171128](https://doi.org/10.1246/cl.171128)
 10. Y. Sasano, R. Sato, Y. Shigeta, N. Yasuda, *H. Maeda, “H-Aggregated π-Systems Based on Disulfide-Linked Dimers of Dipyrrrolyldiketone Boron Complexes,” *J. Org. Chem.* **82**, 11166–11172, (2017), [10.1021/acs.joc.7b02185](https://doi.org/10.1021/acs.joc.7b02185)
 11. A. Kuno, N. Tohnai, N. Yasuda, *H. Maeda, “Conjunction of Pyrrole and Amide Moieties: Highly Anion-Responsive π-Electronic Molecules Forming Ion-Free and Ion-Pairing Assemblies,” *Chem. Eur. J.*, **23**,

- 11357-11365, (2017), [10.1002/chem.201701921](https://doi.org/10.1002/chem.201701921)
12. *T. Fujita, Y. Haketa, H. Maeda, T. Yamamoto, "Relating Stacking Structures and Charge Transport in Crystal Polymorphs of the Pyrrole-Based π -Conjugated Molecule," *Org. Electron.*, **49**, 53-63, (2017), [10.1016/j.orgel.2017.06.028](https://doi.org/10.1016/j.orgel.2017.06.028)
 13. K. Nakamura, *H. Maeda, "Pyrrole-Based Hydrogen-Bonding Dimers Providing Discotic Columnar Structures," *Chem. Lett.*, **46**, 1269-1271, (2017), [10.1246/cl.170487](https://doi.org/10.1246/cl.170487)
 14. R. Yamakado, M. Hara, S. Nagano, T. Seki, *H. Maeda, "Photo-Responsive Soft Ionic Crystals: Ion-Pairing Assemblies of Azobenzene Carboxylates," *Chem. Eur. J.*, **23**, 9244-9248, (2017), [10.1002/chem.201701925](https://doi.org/10.1002/chem.201701925)
 15. Y. Sasano, N. Yasuda, *H. Maeda, "Deprotonated Meso-hydroxyporphyrin as a Stable π -electronic Anion: the Building Unit of Ion-pairing Assembly," *Dalton Trans.* **46**, 8924-8928, (2017), [10.1039/c7dt01635b](https://doi.org/10.1039/c7dt01635b)
 16. Y. Haketa, *H. Maeda, "Dimension-controlled Ion-pairing Assemblies Based on π -electronic Charged Species", *Chem. Commun.*, **53**(20), 2894-2909, (2017), [10.1039/c6cc10255g](https://doi.org/10.1039/c6cc10255g)
 17. V. Lakshmi, Y. Haketa, R. Yamakado, N. Yasuda, *H. Maeda, "Dimension-controlled Assemblies of Anion-responsive π -electronic Systems Bearing Aryl Substituents with Fan-Shaped Geometries", *Chem. Commun.*, **53**(27), 3834-3837, (2017), [10.1039/c7cc01551h](https://doi.org/10.1039/c7cc01551h)
 18. R. Yamakado, Y. Ashida, R. Sato, Y. Shigeta, N. Yasuda, *H. Maeda, "Cooperatively Interlocked [2+1]-Type π -System-Anion Complexes", *Chem. Eur. J.*, **23**(17), 4160-4168, (2017), [10.1002/chem.201605765](https://doi.org/10.1002/chem.201605765)
 19. Y. Sasano, N. Yasuda, *H. Maeda, "Negatively Charged π -Electronic Systems by Deprotonation of Hydroxy-Substituted Dipyrrolyldiketone Boron Complexes", *Chem. Asian J.*, **11**(23), 3423-3429, (2016), [10.1002/asia.201601205](https://doi.org/10.1002/asia.201601205)
 20. R. Yamakado, R. Sato, Y. Shigeta, *H. Maeda, "Ion-Pairing Crystal Polymorphs of Interlocked [2+1]-Type Receptor-Anion Complexes", *J. Org. Chem.*, **81**(18), 8530-8536, (2016), [10.1021/acs.joc.6b01688](https://doi.org/10.1021/acs.joc.6b01688)
 21. Y. Haketa, Y. Tamura, N. Yasuda, *H. Maeda, "Dipyrrolylpyrimidines as Anion-responsive π -Electronic Systems", *Org. Biomol. Chem.*, **14**(34), 8035-8038, (2016), [10.1039/c6ob01466f](https://doi.org/10.1039/c6ob01466f)
 22. Y. Haketa, D. Katayama, S. Fukunaga, Y. Bando, T. Sakurai, W. Matsuda, S. Seki, *H. Maeda, "Ion-Free and Ion-Pairing Assemblies of Anion-Responsive π -Electronic Systems Possessing Directly Linked Alkyl Chains", *Chem. Asian J.*, **11**(14), 2025-2029, (2016), [10.1002/asia.201600712](https://doi.org/10.1002/asia.201600712)
 23. W. Naito, N. Yasuda, T. Morimoto, Y. Shigeta, H. Takaya, I. Hisaki, *H. Maeda, "Doubly N-Methylated Porphyrinoids", *Org. Lett.*, **18**(12), 3006-3009, (2016), [10.1021/acs.orglett.6b01377](https://doi.org/10.1021/acs.orglett.6b01377)
 24. Y. Haketa, R. Takasago, *H. Maeda, " β -Perfluoroalkyl-substituted Pyrrole as an Anion-Responsive π -electronic System Through a Single NH Moiety", *Chem. Commun.*, **52**(46), 7364-7367, (2016), [10.1039/c6cc03619h](https://doi.org/10.1039/c6cc03619h)
 25. K. Nakamura, N. Yasuda, *H. Maeda, "Dimension-controlled Assemblies of Modified Bipyrroles Stabilized by Electron-withdrawing Moieties", *Chem. Commun.*, **52**(44), 7157-7160, (2016), [10.1039/c6cc03423c](https://doi.org/10.1039/c6cc03423c)
 26. Y. Bando, Y. Haketa, T. Sakurai, W. Matsuda, S. Seki, H. Takaya, *H. Maeda, "Ion-Pairing Assemblies Based on Pentacyano-Substituted Cyclopentadienide as a π -Electronic Anion", *Chem. Eur. J.* **22**(23), 7843-7850, (2016), [10.1002/chem.201600686](https://doi.org/10.1002/chem.201600686)
 27. R. Yamakado, T. Sakurai, W. Matsuda, S. Seki, N. Yasuda, S. Akine, *H. Maeda, " π -Electronic Systems That Form Planar and Interlocked Anion Complexes and Their Ion-Pairing Assemblies", *Chem. Eur. J.*, **22**, 626-638, (2016), [10.1002/chem.201503654](https://doi.org/10.1002/chem.201503654)
 28. R. Yamakado, *H. Maeda, "Ion-pairing Assemblies of Photoresponsive Cations and an Interlocked [2+1]-type π -system-anion Complex" *J. Photochem. Photobiol. A*, **331**, 215-223, (2016), [10.1016/j.jphotochem.2015.10.013](https://doi.org/10.1016/j.jphotochem.2015.10.013)
 29. 羽毛田洋平・山門陵平・*前田大光, 「イオンペア集合体を形成するアニオン応答性 π 電子系の合成」有機合成化学協会誌, **74** (3), 243-253, (2016)
 30. H. Maeda, A. Fukui, R. Yamakado, N. Yasuda, "Dipyrrolyphenol as a Precursor of π -electronic Anion that Forms Ion Pairs with Cations" *Chem. Commun.*, **51**, 17572-17575, (2015), [10.1039/c5cc07493b](https://doi.org/10.1039/c5cc07493b)

31. *H. Maeda, K. Chigusa, R. Yamakado, T. Sakurai, S. Seki, "Carboxylate-Driven Supramolecular Assemblies of Protonated meso-Aryl-Substituted Dipyrrolypyrazoles," *Chem. Eur. J.*, **21**, 9520-9527, (2015), 10.1002/chem.201500681
32. *H. Maeda, "Dimension-Controlled Assemblies Comprising π -Electronic Systems" *Chem. Rec.*, **15**, 1151-1152, (2015), 10.1002/tcr.201510007
33. *H. Maeda, T. Nishimura, A. Tsujii, K. Takaishi, M. Uchiyama, A. Muranaka, "Helical π -Systems of Bidipyrrin-Metal Complexes", *Chem. Lett.*, **43**, 1078-1080, (2014), 10.1246/cl.140260
34. R. Sekiya, Y. Tsutsui, W. Choi, T. Sakurai, *S. Seki, Y. Bando, *H. Maeda, "Ion-based assemblies of planar anion complexes and cationic PtII complexes", *Chem. Commun.*, **50**, 10615-10618, (2014), 10.1039/c4cc04565c

(2) 学会発表

A01 : 動的秩序の探査

【秋山修志】

1. Shuji Akiyama, Slow and Temperature-compensated ATP Hydrolysis Reaction Catalyzed by Circadian Clock Protein KaiC, The 3rd Solar FuelMaterial Workshop, 2018.3.18, Osaka, Japan
2. 秋山修志*, 地球の自転周期がエンコードされたタンパク質,KaiC 講演会, 京都大学, 2018.2.6, 京都
3. Shuji Akiyama, Cyanobacterial circadian clock system: How and why can it be so slow and stable? Sokendai Asian Winter School, 分子科学研究所, 2017.12.15, Okazaki
4. Shuji Akiyama, Frequency characteristics as a basis for meta-tuning of the Cyanobacterial Circadian Clock, ConBio2017, 神戸ポートアイランド, 2017.12.9, Kobe
5. 秋山修志*, Frequency Characteristics and Temperature Compensation in the Cyanobacterial Circadian Clock, 第 24 回日本時間生物学会学術大会, 京都大学, 2017.10.29, Kyoto
6. 秋山修志*, Lessons from Cyanobacterial Circadian Clock System, 第 55 回日本生物物理学会年会, 熊本大学, 2017.9.20, 熊本
7. 秋山修志*, 生命の時間, 第 4 回 大型実験施設とスーパーコンピュータとの連携利用シンポジウム, 東京・秋葉原 UDX, 2017.9.8, 東京
8. Shuji Akiyama, The frontier in cyanobacterial circadian clock system, OIIB Summer School 2017, 2017.8.17, Okazaki, Japan
9. 秋山修志*, 藍藻生物時計システムにおける概日周期の根源と貫階層性, 第 69 回日本細胞生物学会大会, 仙台国際センター, 2017.6.13, 仙台市
10. Shuji Akiyama, Atomic-scale origins of slowness and temperature compensation in KaiC Circadian Clock of Cyanobacteria during 1991-2017, 2017.3.12, Nagoya, Japan
11. Shuji Akiyama, Biological X-ray Solution Scattering Activities at the Institute for Molecular Science Okazaki Conference 2017 on Grand Challenges in Small-angle Scattering, 2017.3.19, Okazaki, Japan
12. 秋山修志*, 生物の不思議～体内時計～, 日本放射光学会市民公開講座, 神戸芸術センター2017.1.8, 神戸市
13. Shuji Akiyama, Cyanobacterial circadian clock system. How and why can it be so slow and stable? Sapporo Symposium on Biological Rhythm, 2016.11.10, Sapporo, Japan
14. 秋山修志*, 生体分子システム研究に必要なツールとは?中性子生命科学研究会 「生命科学における中性子利用の現状と未来」, IQBRC 1 階大会議室, 2016.5.17
15. Mukaiyama A*, Abe J, and Akiyama S, Circadian periodicity encoded in KaiC ATPase Winter School of ASIA CORE Program, 2016.2.26, Beijing, China (Invited talk)
16. Shuji Akiyama, *X-ray Solution Scattering as Research Tools for Bio-molecular Systems, Winter School of ASIA CORE Program, 2016.2.26, Beijing, China (Invited talk)
17. Shuji Akiyama, Atomic-scale origins of slowness in the cyanobacterial circadian clock, 8th Japan-Korea Seminars on Biomolecular Science: Experiments and Simulation, 2016.2.16, Okazaki, Japan (Invited talk)
18. 秋山修志, タンパク質の奏でる生体リズム, 城北科学の日, 岡崎市立城北中学校, 2016.2.12, 岡崎市 (基調講演)
19. 秋山修志*, 階層横断的に機能する分子システムの構造アンサンブルとダイナミクス研究 第 29 回日本放射光学会年会, 東京大学柏の葉キャンパス駅前サテライト 2016.1.11, 東京 (招待講演)

20. 秋山修志*, 藍藻生物時計システムに見られる貫階層性, 第6回神経科学と構造生物学の融合研究会, 岡崎シンファレンスセンター2015.11.26. 岡崎市 (招待講演)
21. 秋山修志*, 藍藻の時計タンパク質に内包された概日周期と遅さの根源
藍藻の分子生物学 2015, かずさアカデミアホール 2015.11.17 (招待講演)
22. Shuji Akiyama, KaiC as Circadian Pacemaker of Cyanobacterial Circadian Clock, Grand Design of Molecular Systems, 分子科学研究所 2015.10.8 (招待講演)
23. 秋山修志, 生物時計システムに見られる階層性～原子から細胞スケールまでを貫くロジック～, 2015年度第1回バイオ单分子研究会, 新世代研究所, 2015.8.26 (招待講演)
24. Shuji Akiyama, KaiC as Circadian Pacemaker of Cyanobacterial Circadian Clock, European Biological Rhythms Society (EBRS)/World Congress of Chronobiology (WCC) meeting, 2015.8.6, Manchester, UK (Invited talk)
25. 秋山修志, 概日時計システム研究における bioSANS への期待と展望, 第3回 Neutrons in Biology 研究会, 日本原子力研究開発機構・原子力科学研究所, 2015.3.31 (招待講演)
26. 秋山修志, 会議報告, ガイドラインに関する査読者とのやり取り, 分子システム研究における溶液散乱, 分子科学研究所, 2014.12.20 (招待講演)
27. Shuji Akiyama, Watching slow but ordered dynamics of clock proteins, 第21回日本時間生物学会学術大会, 九州大学, 2014.11.8, 福岡 (招待講演)
28. 秋山修志, 概日時計システム研究における bioSANS への期待と展望, 平成26年度第1回生物構造学研究会, エッサム神田ホール 401 会議室, 2014.10.3, 東京 (招待講演)
29. Shuji Akiyama, KaiC as Circadian Pacemaker of Cyanobacterial Circadian Clock, Society for Research on Biological Rhythms, 2014.6.17, Montana, USA (Invited talk)
30. Shuji Akiyama, Small-angle x-ray Scattering Study on Cyanobacterial Clock Protein KaiC, The 6th Taiwan-Japan Joint Meeting on X-ray and Neutron Scattering, 2014.3.11, Taipei, Taiwan (Invited talk)
31. Shuji Akiyama, KaiC as A Circadian Pacemaker of Cyanobacterial Circadian Clock, Asian CORE Winter School on Frontiers of Molecular, Photo-, and Material Sciences, 2014.2.25, Taipei, Taiwan (Invited talk)
32. Shuji Akiyama, KaiC as A Circadian Pacemaker of Cyanobacterial Circadian Clock
6th Japan-Korea Seminars on Biomolecular Science, 2013.11.27, Okazaki, Japan (Invited talk)
33. Shuji Akiyama, KaiC as a Circadian Pacemaker of Cyanobacterial Circadian Clock, 15th Japan-Korea Symposium on Molecular Science, Hierarchical Structure from Quantum to Functions of Biological Systems, 2013.7.5, Sannomiya, Japan (Invited talk)
34. Shuji Akiyama, Circadian Pacemaker of Cyanobacteria by Intra-molecular Feedback Regulation of KaiC ATPase, IMS Workshop on "Hierarchical Molecular Dynamics: From Ultrafast Spectroscopy to Single Molecule Measurements", 2013.6.25, Okazaki, Japan (Invited talk)

【秋山良】

1. Ryo Akiyama, Statistical Mechanics Study of Reentrant Phase Behavior of Proteins in an Electrolyte Solution, The 17th KIAS Conference on Protein Structure and Function, 2017.9.21-23, Seoul, Korea (Invited talk)
2. 秋山良, 積分方程式理論で計算した電解質溶液中の同符号荷電粒子間実効相互作用, 第6回ソフトマター研究会, 北海道大学, 2016.10.25, 札幌 (招待講演)
3. 秋山良, 電解質溶液中で働く強い同符号荷電間実効引力の理論的研究とタンパク質の凝集挙動, 福岡大学化学生物学系談話会, 福岡大学, 2016.8.9, 福岡 (講演会)
4. 秋山良, 巨大分子を使った寄木細工の為の方針の2つ『分子を使った寄せ木細工』:自己組織化したソフトマテリアルが織り成す『かたち』と機能, 島根大学教育学部, 2016.5.28, 松江
5. 秋山良, 電解質溶液中の同符号荷電コロイド粒子間実効引力と蛋白質溶液の相図, 慶應義塾大学理工学部物理学科談話会, 2015.11.24, 横浜 (講演)
6. Ryo Akiyama, Effective attraction between negatively charged sites on proteins and ordering of proteins in an electrolyte solution The 4th International Symposium "Dynamical Ordering of Biomolecular Systems for creation of integrated functions" (新学術領域研究『動的秩序と機能』第4回国際シンポジウム), 2015.11.22-23, (招待講演)
7. Ryo Akiyama, Spatiotemporal dynamic ordering regulated by ATP hydrolysis and effective attraction between negatively charged sites in a biofluid, International, The 53rd Annual Meeting of the Biophysical Society of Japan, Formation of spatiotemporal dynamic ordering mediated by ATP

- hydrolysis (新学術 領域研究『動的秩序と機能』共催シンポジウム), 2015.9.14, Kanazawa, (招待講演)
8. 秋山良, 静電相互作用による巨大分子構造形成とイオン交換による制御, 日本化学会第95回春季年会, 日本大学理工学部／薬学部 船橋キャンパス, 2015.3.26, 船橋 (特別企画『水溶液における静電的相互作用の本質を探る』における講演)
 9. 秋山良, 同符号電荷間の引力とアクチンの会合挙動, 茨城大学理学部物理学科セミナー, 茨城大学, 2014.11.18, 水戸 (学科講演)
 10. Ryo Akiyama, Association of Actin Monomers and Effective Attraction between Like-Charged Colloidal Particles Mediated by Multivalent Cations, 第52回日本生物物理学会シンポジウム『生体分子機械の動作機構を水から眺めてみる』, 札幌コンベンションセンター, 2014.9.26, 札幌 (シンポジウム講演)
 11. Shingo Fujihara, Akira Yoshimori, Takuto Sawayama, and Ryo Akiyama, Interaction between like-charged particles and attractive patches on like-charged proteins International of Statistical Mechanics of Molecular Liquid on Soft Matter, Kasetsart University, 2014.9.15, Bangkok, Thailand (Invited talk)
 12. 秋山良, 多価カチオンによって媒介される同符号アニオン間実効引力と秩序構造 (招待講演)
アニオンの科学研究会, 東京工業大学大岡山キャンパス, 2014.9.6, 東京 (招待講演)

【池谷鉄兵】

1. 池谷鉄兵, NMR データの自動解析および蛋白質立体構造計算の現状と創薬研究への応用の可能性, 横浜 NMR 研究会 第58回ワークショップ, 2017.9.26, 横浜 (招待講演)
2. 池谷鉄兵, 池田思朗, 木川隆則, 伊藤隆, Güntert Peter, スペース NMR データを用いた細胞内蛋白質立体構造決定, 第54回日本生物物理学会年会, つくば国際会議場, 2016.11.25-27, つくば (招待講演)
3. Teppei Ikeya, "Improved in-cell structure determination of proteins at near-physiological concentration", UK-Japan Symposium "From single molecules to cells and tissues", University of Leicester, 2016.7.5, Leicester, UK (Invited talk)
4. Teppei Ikeya, "Protein NMR Structure Refinement based on Bayesian Inference", High-Dimensional Data Driven Science, 2015.12.14-17, Kyoto (Invited talk)
5. 池谷鉄兵, ベイズ推定を用いた立体構造計算と in-cell NMR データへのアプローチ In-cell NMR シンポジウム "Computational aspects in biomolecular & in-cell NMR", 2015.4.24, 東京, (招待講演)
6. Teppei Ikeya, Jin Inoue, Peter Güntert, Yutaka Ito, three-Dimensional Protein Structure and Dynamics in Living Cells, 7th Korea-Japan Seminars on Biomolecular Sciences, 2014.11.27 韓国ソウル, 熊本, (招待講演)
7. 池谷鉄兵, 伊藤隆, 生きた細胞内の蛋白質の立体構造とダイナミクス, 構造活性関連シンポジウム, 2014.11.13, 熊本 (招待講演)
8. 池谷鉄兵, 井上仁, 伊藤隆, 生きた細胞中の天然変性蛋白質の動態, 第52回日本生物物理学会年会, 2014.9.25, 札幌 (招待講演),
9. 池谷鉄兵, 伊藤隆, タンパク質の立体構造計算における NMR データの可視化, 日本分光学会 NMR 分科会 2014, 2014.9.11, 東京 (招待講演)
10. 池谷鉄兵, 伊藤隆, A NMR structure calculation algorithm based on Bayesian inference, 大阪大学蛋白質研究所セミナー IPR Seminar, 2014.6.19, 大阪 (招待講演)

【岩田耕一】

1. 岩田耕一, 化学反応を追跡するための分光実験, 神奈川大学理学部化学科シンポジウム「化学のための最先端光科学」, 2018.3.10, 平塚市 (招待講演)
2. Koichi Iwata, Spectroscopy of Molecules Favoured by Raman Followers, 90 Years of Raman Effect: Current Status and Future Directions, 2018.3.1, Bangalore, India (Keynote lecture)
3. Koichi Iwata, Characterization of Lipid Bilayer Membranes with Time-Resolved Spectroscopy The 10th Asian Conference on Ultrafast Phenomena (ACUP2018), 2018.1.10, Hong Kong, China (Invited talk)
4. Koichi Iwata, Characterizing Lipid Bilayer Membranes with time-resolved Spectroscopies – Viscosity and Polarity, The 14th Biennial DAE-BRNS Trombay Symposium on Radiation and Photochemistry (TSRP-2018), 2018.1.3, Mumbai, India (Invited talk)
5. 岩田耕一, 時間分解分光法でみる化学反応, 公開講演会 最新化学談話シリーズ, 九州大学理学部化学教室談話会, 2017.11.20, 福岡 (招待講演)
6. Koichi Iwata, Loose electrons traced with time-resolved near-infrared spectroscopy - absorption and non-linear Raman, International Conference on Spectroscopy of Biomolecules and Advanced Materials (ICSBAM 2017), 2017.10.4, Alappuzha (India), Kerala, India (Invited talk)

7. 岩田耕一, 特異な化学反応場としての脂質二重膜-時間分解分光法による特性評価, 第11回分子科学討論会, 2017.9.15, 仙台 (招待講演)
8. Koichi Iwata, Examining Lipid Bilayer Membranes with Time-resolved Spectroscopies 6th Asian Spectroscopy Conference (ASC6), 2017.9.5, Hsinchu, Taiwan (Keynote lecture)
9. Koichi Iwata, Basic Theory for Non-linear Raman Processes, Taiwan Association of Raman Spectroscopy Summer Camp (TARS Summer Camp 2017), 2017.6.30, Chiayi City, Taiwan, (Invited talk)
10. Koichi Iwata, Electronic and Vibrational Relaxation Examined with Femtosecond Time-resolved Absorption and Stimulated Raman Spectroscopy in Near-infrared, 5th Taiwan International Symposium on Raman Spectroscopy (TISRS2017), 2017.6.28, Chiayi City, Taiwan (Keynote lecture)
11. Koichi Iwata, Resonance Raman Spectroscopy of “Loose Electrons” with Stimulated Raman Scattering in Near-infrared, 9th International Conference on Advanced Vibrational Spectroscopy (ICAVS2017), 2017.6.12, Victoria, Canada (Invited talk)
12. Koichi Iwata, Properties of liposome lipid bilayer membranes examined by time-resolved Raman and fluorescence spectroscopies, 3rd Symposium on Weak Molecular Interactions, 2017.3.29, Opole-Groszowice, Poland (Invited talk)
13. Koichi Iwata, How we examine lipid bilayer membranes as field of chemical reaction with time-resolved spectroscopy, International Workshop Present and Future of Ultrafast Spectroscopy, 2017.3.14, Wako, Japan (Invited talk)
14. Koichi Iwata, Local environment in ionic liquids and lipid bilayer membranes examined with Raman spectroscopy, 6th International Conference on Perspectives in Vibrational Spectroscopy (ICOPVS-2016), 2016.11.5, Lucknow, India (Keynote lecture)
15. Koichi Iwata, What we see by time-resolving in near-IR, SciX2016, 2016.9.22, Minneapolis, USA
16. Koichi Iwata, Femtosecond Time-resolved Near-infrared Absorption Spectroscopy of “Loose Electrons” as Photochemical Intermediates, Recent Advances in Molecular Spectroscopy: Fundamentals and Applications in Materials and Biology (RAMS2016), 2016.3.3, Hyderabad, India (Invited talk)
17. Yuki Nojima, Sho Kitamura, Tomohisa Takaya, Koichi Iwata, Characterizing lipid bilayer membranes with time-resolved spectroscopies, 2015 International Chemical Congress of the Pacific Basin Societies (Pacificchem 2015), 2015.12.16, Honolulu, Hawaii, USA (Invited talk)
18. Koichi Iwata, Lipid Bilayer Membranes as Field of Chemical Reactions – Characterization by Picosecond and Femtosecond Time - resolved Spectroscopy, First International Symposium of Institute for Catalysis – Global Collaboration in Catalysis Science toward Sustainable Society, 2015.10.15, Sapporo, Japan (Invited talk)
19. Koichi Iwata, Weak Molecular Interaction in Condensed Phases Examined with Time-resolved spectroscopies - Raman and Near-Infrared Absorption, 5th Asian Spectroscopy Conference (ASC5), 2015.10.2, Sydney, Australia (Invited talk)
20. Koichi Iwata, Ashok Zachariah Samuel, Sohshi Yabumoto, Kenichi Kawamura (招待講演)
Structure of porous PMMA thin film examined with multifocus Raman microspectroscopy
SciX2015, Providence, 2015.9.28, Rhode Island, USA (Invited talk)
21. Y. Nojima, T. Takaya, K. Iwata, Thermal Diffusivity of Lipid Bilayer Membranes Estimated with Picosecond Time-resolved Raman Spectroscopy, 3rd Taiwan International Symposium on Raman Spectroscopy (TISRS 2015), 2015.7.1, Sun-Moon Lake (Taiwan) (Invited talk)
22. Goh Mohri, Tomohisa Takaya, and Koichi Iwata, Photoinduced Electron-Transfer Reaction of 9,9'-Bianthryl in Liposome Lipid Bilayers Observed with Femtosecond Time-Resolved Near-Infrared Spectroscopy, 2015 Korea-Japan Symposium on Frontier Photoscience (KJFP-2015), 2015.6.27, Jeju (Korea) (Invited talk)
23. Koichi Iwata, Characteristics of Amphiphile Liquids Examined with Spectroscopic Methods International Workshop on Ionic Liquids and Related Materials, 2014.11.12, 東京 (Invited talk)
24. Manping Ye, Setsuka Arii, Takanori Uwabo, Tomohisa Takaya, Koichi Iwata, Structure of ionic liquids examined with smaller probes, 248th National ACS Meeting, 2014.8.10, San Francisco (USA), (Invited talk)
25. Koichi Iwata, Characterizing molecular assemblies with time-resolved Raman spectroscopy - lipid bilayer membranes, ionic liquids and loose electrons, Friedrich Schiller University Jena, 2014.8, Jena, Germany (Keynote lecture)
26. Setsuka Arii, Tomohisa Takaya, Koichi Iwata, Formation of Solvated Electrons in Alcohols Observed

- with Femtosecond Time-resolved Near-infrared Spectroscopy, – Japan Symposium on Frontier Photoscience (KJFP-2014), 2014.6.21, Seoul , Korea (Invited talk)
27. 岩田耕一, 分光で探るイオン液体の相互作用と構造, 平成 26 年度イオン液体研究会, 2014.6.17, 東京 (Invited talk)
- 【内橋貴之】
1. 内橋貴之, 高速原子間力顕微鏡で可視化する生体膜反応ダイナミクス, 2017 年度 生命科学系合同年次大会 ワークショップ「最先端の表面科学手法による生体膜反応の実動作下計測」2017.12.8, 神戸 (招待講演)
 2. Takayuki Uchihashi, High-speed atomic force microscopy: A tool for visualizing dynamic behavior from proteins to cells, The 28th 2017 International Symposium on Micro-NanoMechanical and Human Science, 2017.12.6, Japan (Keynote lecture)
 3. Takayuki Uchihashi, Visualization of Single-Molecule Dynamics Using High-Speed Atomic Force Microscopy, The 2nd Korea-Japan Joint Symposium on Single-Molecule Biophysics 2017, 2017.11.9, Korea (Invited talk)
 4. 内橋貴之, 高速原子間力顕微鏡で可視化する Kai タンパク質間相互作用のダイナミクス
第 24 回日本時間生物学会学術大会 シンポジウム「24 時間の創出原理」, 2017.10.29, 京都 (招待講演)
 5. Takayuki Uchihashi, Direct observation of single molecule dynamics at work with high-speed atomic force microscopy, Frontier in Single Molecule Biophysics 2017, 2017.10.16, Israel (Invited talk)
 6. Takayuki Uchihashi, High speed atomic force microscopy for a tool to visualise dynamic events on biological systems from single molecules to living cells, Workshop on “Nanofluidics in Biological Systems”, 2017.9.13, UK (Invited talk)
 7. 内橋貴之, 高速原子間力顕微鏡による生体分子のダイナミクス計測, 新世代研究所 バイオ单分子研究会「タンパク質の作動原理の理解へ向けて - 機能する姿を活写する -」, 2017.9.11, 富山 (招待講演)
 8. 内橋貴之, 高速原子間力顕微鏡で可視化する生体・人工高分子の動態, 2017 年真空・表面科学合同講演会, 表面: プローブ顕微鏡研究部会「走査プローブ顕微鏡によるナノ表面科学の最前線」, 2017.8.19, 横浜 (招待講演)
 9. 内橋貴之, 高速 AFM を用いた生体分子のその場観察, 2017 年真空・表面科学合同講演会, 合同シンポジウム 「バイオ表面・界面, 細胞, 生体組織のオペランド計測」, 2017.8.17, 横浜 (招待講演)
 10. Takayuki Uchihashi, Direct visualization of single molecule dynamics by high-speed atomic force microscopy" Telluride Science Research Center Workshop on Protein Dynamics, 2017.7.5, USA (Invited talk)
 11. 内橋貴之, 高速原子間力顕微鏡で可視化するタンパク質の動的秩序, 第 17 回 日本蛋白質科学会年会, ワークショップ「蛋白質動的秩序のマルチプローブを用いた統合的解析」, 2017.6.22, 仙台 (招待講演)
 12. 内橋貴之, 高速 AFM で明らかにする Kai タンパク質間の動的相互作用, 第 69 回 日本細胞生物学会大会, シンポジウム「分子の集合・離脱がつかさどる動的な細胞機能」, 2017.6.13, 仙台 (招待講演)
 13. Takayuki Uchihashi, Yo-hei Watanabe, Ryota Iino and Toshio Ando, Structural Flexibility and Chaperone Activity of TCpB revealed by High-Speed AFM, XIX. Annual Linz Winter Workshop, Sommerhotel Julius-Raab-Heim, 2017.2.3-6, Linz, Austria (Invited talk)
 14. Takayuki Uchihashi, Direct Visualization of Single Molecule Dynamics at Work with High-Speed Atomic Force Microscopy, BioNANO 2016, 22-23 November 2016, Malopolska Centre of Biotechnology, Jagiellonian University, 2016.11.22, Krakow, Poland (Invited talk)
 15. 内橋貴之, タンパク質のダイナミクスを可視化する高速原子間力顕微鏡
生体ボリュームイメージング研究部会 & 生理研研究会合同ワークショップ「電子顕微鏡ビッグデータが拓くバイオメディカルサイエンス」～限界を超えるための顕微鏡技術～, 2016.11.17, 岡崎 (招待講演)
 16. 内橋貴之, 高速原子間力顕微鏡で可視化する生体分子のダイナミクス, 理研シンポジウム: 第 4 回「光量子工学研究 -若手・中堅研究者から見た光量子工学の展開-」, 2016.11.1, 和光, 埼玉県 (招待講演)
 17. 内橋貴之, 高速 AFM による生体分子ダイナミクスのその場観察, 日本物理学会 2016 年秋季大会 領域 9 シンポジウム「表面界面ナノ構造のその場観察」, 2016.9.15, 金沢 (招待講演)
 18. 内橋貴之, 高速 AFM による生体試料の動態イメージング, 第 25 回日本バイオイメージング学会学術集会 シンポジウム 3「ナノバイオイメージング: 1 分子から細胞までの先端手法」, 2016.9.6, 名古屋 (招待講演)
 19. Takayuki Uchihashi, Noriyuki Kodera and Toshio Ando, Direct Visualization of Single Molecule Dynamics by High-Speed Atomic Force Microscopy, Gordon Research Conference, Single Molecule Approach to Biology, Chinese University of Hong Kong, 2016.7.3-8, China (Invited talk)
 20. 内橋貴之, 高速原子間力顕微鏡で調べる回転分子モーターの構造ダイナミクス

- 第 16 回 日本蛋白質科学会年会 ワークショップ”回転分子モーターを調べてつくって理解する”, 2016.6.9, 福岡 (招待講演)
21. 内橋貴之, Visualization of Functional Dynamics of Biological Molecules by High-Speed AFM
分子研研究会 “超機能分子の創成 合成, 計測, 数理が織りなす社会実装分子の戦略的設計と開発”, 2016.6.27, 岡崎 (招待講演)
 22. 内橋貴之, 高速 AFM による膜タンパク質のダイナミクス観察, 蛋白研セミナー ”膜タンパク質の構造ダイナミクス”, 2016.5.12-13, 大阪 (招待講演)
 23. Takayuki Uchihashi, Direct Observation of Single Molecule Dynamics at Work with High-Speed Atomic Force Microscopy, Les Houches-TSRC Workshop on Protein Dynamics, Ecole de Physique des Houches, 2016.4.3-8, France (Invited talk)
 24. 内橋貴之, 高速原子間力顕微鏡で探る生体分子の機能ダイナミクス, 表面科学技術研究会 2016 生きている固体と界面 ～走査型プローブ顕微鏡で探る～, 神戸大学, 2016.1.21, 神戸市 (招待講演)
 25. 内橋貴之, 高速原子間力顕微鏡の開発とバイオ応用, 日本顕微鏡学会 様々な極微イメージング技術若手研究部会 第 3 回研究会, 2015.11.23, 龍野, 兵庫県 (招待講演)
 26. Takayuki Uchihashi, Direct Visualization of Single Molecule Dynamics at Work with High-Speed Atomic Force Microscopy The 15th KIAS Conference on Protein Structure and Function, September 2015.9.17-19, Seoul, Korea (Invited talk)
 27. Takayuki Uchihashi, High-Speed Atomic Force Microscopy for Observation of Single-Molecule Dynamics The 6th RSC-CSJ Symposium (held in conjunction with the 95th CSJ Annual Meeting), 日本大学船橋, 2015.3.27, 2, 千葉 (招待講演)
 28. Takayuki Uchihashi, High-speed atomic force microscope for imaging of biomolecular dynamics at solid surface 10th Annual International Electromaterials Science Symposium, 2015.2.11-13, Wollongong, Australia (Invited talk)
 29. Takayuki Uchihashi, Visualization of single molecule dynamics at work with high-speed atomic force microscopy, The 7th Biennial Australian Colloid & Interface Symposium, 2015.2.1-5, Hobart, Tasmania, Ausutralia (Keynote lecture)
 30. 内橋貴之, 高速原子間力顕微鏡によるタンパク質のダイナミクスと物性計測第 9 回 NIBB バイオイメージングフォーラム「物理特性のイメージング」, 基礎生物学研究所, 2015.1.27, 愛知, 2015 年 1 月 27 日 (招待講演)
 31. 内橋貴之, 高速原子間力顕微鏡の開発とバイオ応用, 新世代研究所水和ナノ構造・界面ナノ科学合同研究会 「固液界面の水和ナノ構造と生体高分子ダイナミクス」, 伊豆長岡温泉 天坊 2015.1.25, 静岡 (招待講演)
 32. Takayuki Uchihashi, High-speed atomic force microscope for studying dynamic interactions in biomolecular system The 3rd International Symposium on Dynamical Ordering of Biomolecular Systems for Creation of Integrated Functions, 合歓の郷, 2015.1.10-11, 三重(Invited talk)
 33. Takayuki Uchihashi, Single-molecule imaging of proteins at work with high-speed atomic force microscopy, 16th International Conference on Retinal Proteins, 琵琶湖ロイヤルホテル, 2014.10.5-10, 滋賀 (Invited talk)
 34. Takayuki Uchihashi, Cooperative Conformational Change in Ring-Shaped ATPase Observed by High-Speed AFM, 第 52 回日本生物物理学会年会 Japan-China-Taiwan Joint Symposium on Cooperativity in Supramolecular Machine, 札幌国際会議場, 2014.9.26, 北海道(Invited talk)
 35. Takayuki Uchihashi, High-speed atomic force microscopy for imaging of protein dynamics" Agilent Nanomeasure 2014, 2014.9.16-17, Beijing, China (Invited talk)
 36. 内橋貴之, 高速 AFM による生体試料のダイナミクス観察」, 日本膜学会第 36 回年会 境界領域シンポジウム「膜解析の最前線~生体膜・膜タンパク質から模擬膜, ソフトマターまで~, 早稲田大学 2014.5.13, 東京 (招待講演)
 37. Takayuki Uchihashi, Visualization of single molecule dynamics at work with high-speed atomic force microscopy, Single Protein Dynamics *in Cellulo* 2014: Spatio-Temporal, Structural and Quantitative Analyses, OIST シーサイドハウス, 2014.4.21-25, 沖縄(Invited talk)

【上久保裕生】

1. 上久保裕生, 光による蛋白質分子複合系の自由エネルギーLANDスケープの変調, 日本化学会 第 98 回春季年会 (2018), 2018.3.20, 千葉県船橋市 (特別企画講演)
2. 上久保裕生, 連続滴定溶液散乱による多成分系解析の標準化, 第 4 回タンパク質 X 線溶液散乱講習会, 2017.12.13, 茨城県つくば市 (招待講演)
3. 上久保裕生, 混合溶液中の蛋白質分子複合系の構造・相互作用評価～デバイス開発とその応用～, 日本学術

- 振興会 構造生物第 169 委員会 第 54 回研究会, 2017.11.29, 東京都文京区日 (招待講演)
4. 上久保裕生, 連続滴定溶液散乱測定による蛋白質分子複合系の多成分平衡状態の解析,
第 40 回溶液化学シンポジウム, 2017.10.9, 兵庫県姫路市 (招待講演)
 5. 上久保裕生, 複数混合溶液中に存在する生体高分子の構造・相互作用評価
分子・物質合成プラットフォーム, 大学連携研究設備ネットワーク主催「ソフトでウェットな素材のやさしい
観察」, 2017.10.6, 東京都江東区
 6. Hironari Kamikubo, Structural Exploring Multi-component Equilibrium in Biological Systems
The 24th Congress & General Assembly of the International Union of Crystallography, 2017.8.24,
Hyderabad, India (Invited talk)
 7. Hironari Kamikubo, Multi-component equilibrium in biological systems explored by using continuous
titration SAXS, Frontier Bioorganization Forum 2017 Dynamical ordering and integrated functions of
biomolecular systems, 2017.4.25, Taipei, Taiwan (Invited talk)
 8. Mikio Kataoka, Hironari Kamikubo, Mechanism of amyloid formation of human calcitonin
The International Chemical Congress of Pacific Basin Societies 2015, 2015.12.15-20, Honolulu, Hawaii,
USA (Invited talk)
 9. 上久保裕生, 水-蛋白質界面の電荷状態と蛋白質内部で生じる化学反応, 第 64 回高分子討論会, 2015.9.16,
宮城県仙台市 (招待講演)
 10. Hironari Kamikubo, Designing an Artificial Protein by Using a Kind of Building Blocks Responsible for
Structure and Function, IMS Asian International Symposium, 2015.6.12, Okazaki, Aichi (Invited talk)
 11. Hironari Kamikubo, Molecular actions in the light sensor protein, Photoactive Yellow Protein
The 7th Korea-Japan Seminars on Biomolecular Sciences: Experiments and Simulations, 2014.11.28,
Seoul Korea (Invited talk)
 12. Hironari Kamikubo, The potential uses of structure and function elements, The 14th KIAS Conference
on Protein Structure and Function, 2014.9.18, Seoul, Korea (Invited talk)
 13. Hironari Kamikubo, Functional modification of a protein by using element implantation, Sixth Korea-
Japan Seminars on Biomolecular Sciences: Experiments and Simulations, 2013.11.27, Okazaki, Japan
(Invited talk)

【佐藤啓文】

1. 山本武志, Predicting the structure of self-assembled supramolecular nanosystems via all-atom molecular
simulation, 第 98 回 日本化学会春季年会, アジア国際シンポジウム, 2018.3.23, 千葉 (招待講演)
2. 佐藤啓文, 分子レベルで見る自己集合過程の実時間追跡, 日本化学会第 98 春季年会, 日本大学理工学部 船
橋キャンパス, 2018.3.20, 千葉 (招待講演)
3. Hirofumi Sato, Understanding of Self-Assembly Process at Molecular Level, Frontier Bioorganization
Forum 2017: Dynamical ordering and integrated functions of biomolecular systems, Academia Sinica
2017.4.26, Taipei (Invited talk)
4. Hirofumi Sato, Chemical processes in condensed phase: Quantum chemistry and statistical mechanics
複雑・複合系の理論計算科学に関する 日・仏・スペイン合同シンポジウム, JFS-Joint Symposium on
Theoretical and Computational Science of Complex System, 京都大学福井謙一記念研究センター, FIFC,
Kyoto University, 2016.10.26-28, Kyoto (招待講演)
5. Hirofumi Sato, Molecular theory of chemical processes in condensed phase, International Symposium on
Multi-scale Simulation of Condensed-phase Reacting Systems (MSCRS2016), Noyori Conference Hall,
Nagoya University, 2016.10.10-13, Nagoya (Invited talk)
6. 佐藤啓文, 分子集団・分子集合の理論化学, 第 11 回新大先端化学セミナー, 新潟大学情報理工棟 2 階 第一
ゼミ室, 2016.9.26, 新潟 (招待講演)
7. Hirofumi Sato, Multi-scale couplings for chemical processes at molecular level: quantum chemistry and
statistical mechanics, Pacificchem, 2015.12.16, Honolulu, Hawaii, USA (Invited talk)
8. 佐藤啓文, 凝縮系の理論化学, 化学反応経路探索のニューフロンティア 2015, 2015.9.15, 品川 (招待講演)
9. 佐藤啓文, 動的秩序形成の分子理論, 日本化学会第 95 春季年会 特別企画「生命および人工分子システムに
おける動的秩序形成: 分子論的理解」, 日本大学理工学部 船橋キャンパス 2015.3.26, 千葉, 日本 (招待講演)
10. Takeshi Yamamoto, Molecular calculation of free energetics for nanocube self-assembly in solution, The
3rd International Symposium on Dynamical Ordering of Biomolecular Systems for Creation of
Integrated Functions, 2015.1.10-12, Ise, Japan (Invited talk)

【高田十志和】

1. 高田十志和, Selective Cyclic Polymer Synthesis via Topology Transformation”, 255th ACS National Meeting (Cyclic and Topological Polymer Symposium), 2018.3.18-22, New Orleans, LA, USA (Invited talk)
2. 高田十志和, 架橋高分子を強靭化するロタキサン架橋剤, 第 243 回ゴム技術シンポジウム,日本ゴム協会, 東部ビル, 2018.2.27, 東京 (招待講演)
3. 高田十志和, 固相でも動く分子スイッチと触媒不要のクリック反応剤, 第 63 回山口大学常盤台コロキウム, 山口大学工学部, 2018.2.2, 山口県宇部市 (招待講演)
4. 高田十志和, Stimuli-Responsive Polymers Driven by Rotaxane Molecular Switch
Special Lecture of East China Normal Univ., 2017.12.15, Shanghai, China (Invited talk)
5. 高田十志和, Rotaxane Cross-Linkers Endowing Polymers with Toughness
The 15th Pacific Polymer Conference (PPC-15), 2017.12.10-14, Xiamen, China (Invited talk)
6. 高田十志和, 「可動な架橋によるソフトマテリアルの強靭化」, 関西における日本ゴム協会設立 90 周年記念講演会, メルパルク大阪, 2017.12.8, 大阪 (招待講演)
7. 高田十志和, Mechanically Chiral Molecules: Synthesis, Optical Resolution, and Application
International Workshop on Chirality in Soft Matter, Toyota Physical and Chemical Research Institute, 2017.11.24-26, Nagoya, Japan (Invited talk)
8. 高田十志和, 「インターロック分子科学」, 光ナノサイエンス特別講演, 奈良先端科学技術大学院大学, 2017.11.2, 奈良県生駒市高山町 (招待講演)
9. 高田十志和, 「ロタキサン構造の可動性を活用する新材料開発」, 岡山大学合成有機化学研究室講演会, 岡山大学, 2017.11.4, 岡山市北区津島中 (招待講演)
10. 高田十志和, 「高分子材料とは -高分子の架橋と架橋高分子材料-」, 2017 年度若手社員のための高分子基礎講座 (高分子学会関東支部), 湘南セミナーハウス, 2017.10.19-21, 神奈川県平塚市 (招待講演)
11. 高田十志和, 「ゴム材料強靭化のためのロタキサン架橋」 日本ゴム協会衛生問題研究分科会, 日本電業会館, 2017.10.10, 東京都港区 (招待講演)
12. 高田十志和, Dynamical Ordering of Supramolecular Architecture Comprising Rotaxane-Linked Polymers, The 55th Annual Meeting of The Biophysical Society of Japan, Kumamoto University, 2017.9.19-21, Kumamoto, Japan (Invited talk)
13. Toshikazu Takata, Processive Macrocycle Catalyst for Efficient Polymer Reaction via Pseudorotaxane Intermediate, MMC-17 (IUPAC 17th International Symposium on Macromolecular Complexes), Waseda University, 2017.8.28-31, Shinjuku, Tokyo, Japan (Invited talk)
14. 高田十志和, 「分子の形と動きを制御し新機能物質を創成する」
有機合成化学協会東北支部岩手地区講演会, 岩手大学理工学部, 2017.7.13, 岩手県盛岡市 (招待講演)
15. 高田十志和, トポロジカル結合分子を合成する -分子スイッチ, 分子モーター, 生理活性, そして 分子トポロジー変換-, 第 28 回万有仙台シンポジウム, 仙台国際センター, 2017.6.24, 仙台市青葉区 (招待講演)
16. 高田十志和, Switchable Polyrotaxanes, Gordon Research Conference (Molecular Machines), Holderness School, 2017.6.11-16, New Hampshire, USA (Invited talk)
17. 高田十志和, ロタキサン分子スイッチと分子トポロジー変換, ホストゲスト超分子化学研究会, 立命館大学, 2017.6.4, 滋賀県草津市 (招待講演)
18. 高田十志和, Development of Dynamic Molecular and Macromolecular Systems based on Interlocked Structure (インターロック構造を基盤とする動的機能分子・高分子の開発)
第 97 回春季年会, 慶應大学日吉キャンパス, 2017.3.21, 横浜市港北区 (日本化学会賞受賞講演)
19. 高田十志和, Development of Dynamic Molecular and Macromolecular Systems based on Interlocked Structures, 日本化学会第 97 春季年会(2017) 慶應大学, 2017.3.16-19, 横浜市 (受賞講演)
20. 高田十志和, ロタキサンスイッチの高分子への応用 -トポロジー変換可能な高分子と強靭化-, 信州大学織維学部講演会 信州大学, 2016.9.6-8, 長野 (招待講演)
21. 高田十志和, Novel Cross-Linker Capable of Endowing Polymer with Toughness
9th International Symposium on High-Tech Polymer Materials (HTPM-9), 2016.7.11-15, Zhengzhou China (Invited Talk)
22. 高田十志和, Cross-linking That Endows Rubber with Toughness Using Rotaxane Cross-Linkers
International Polymer Conference of Thailand (PCT-6), 2016.6.30-7.1, Bangkok Thailand (Invited Talk)
23. 高田十志和, 高分子鎖の新しい連結手法, 接着学会粘着研究会 5 月度例会 東京大学, 2016.5.27, 東京 (招待講演)
24. 高田十志和, 架橋点における高分子鎖の可動性が生み出す機能と物性, 日本薬学会北陸支部特別講演会, 富山大学大学院医学薬学研究部・講演会, 2016.3.11, 富山 (招待講演)

25. Toshikazu Takata, "International Symposium on Center of Excellence for Innovative Material Sciences Based on Supramolecules" (超分子による革新的マテリアル開発の拠点形成国際シンポジウム), 金沢大学ミニ国際シンポジウム, 2016.3.10, 金沢 (Plenary Lecture)
26. 高田十志和, 動的結合を活用する高分子開発 -ポリウレタン修飾,環状ポリマー形成,ポリマー強靭化-, 第17回ポリウレタン研究会, 2016.2.12, 名古屋 (招待講演)
27. Toshikazu Takata, "Reversible Topology Transformation of Linear –Cyclic Polymer Using Rotaxane Protocol–", Pachifichem2015, Invited Lecture, 2015.12.15-20, Honolulu (Invited Talk)
28. 高田十志和, インターロック型超分子の特異空間を活かした合成反応,構造体構築, 静岡大学理学部講演会, 招待講演, 2015.11.20, 静岡 (招待講演)
29. 高田十志和, 擬ロタキサン構造を鍵とする0次反応経由高分子反応, 第16回リング・チューブ超分子研究会, 招待講演, 2015.10.26-27, 筑波 (招待講演)
30. Toshikazu Takata, Kazuko Nakazono, Daisuke Aoki, and Hiromitsu Sogawa, "Dynamic Polymer Networks Engineered by Rotaxane Cross-Linkers", 11th International Conference on Advanced Polymers via Macromolecular Engineering Registration (APME2015), 2015.10.18-22, 横浜(Invited talk)
31. 高田十志和, 可変な連結様式をもつブロックコポリマーの合成と構造・物性制御,第64回高分子討論会, 招待講演, 2015.9.15-17, 仙台 (招待講演)
32. Toshikazu Takata, "Role of Mechanical Linkage in Polymer Systems: Tough Polymer and Topology-Transformable Polymer", Japan-Taiwan Bilateral Symposium on Functional Polymer 2015 JTBS2015, Invited Lecture, 2015.9.2-5, 札幌 (Invited talk)
33. 高田十志和, ロタキサン型非共有結合連結がもたらす機能と物性, 名古屋大学応用化学系講演会, 招待講演, 2015.7.24, 名古屋 (招待講演)
34. Toshikazu Takata, "Dynamic Molecular Systems Utilizing Chirality and Switch Functions of Rotaxanes" (ロタキサンのキラリティーとスイッチ機能を活用する動的分子システム), モレキュラーキラリティー2015 (MC2015), 2015.6.12-13, 東京 (招待講演)
35. 高田十志和, 超分子ポリマーの組織化とトポロジー変換, 特別企画「生命および人工分子システムにおける動的秩序形成 : 分子論的理解」, 第95日本化学会春季年会,日本大学理工学部船橋キャンパス, 2015.3.26-29, 千葉県船橋市 (依頼講義)
36. 高田十志和, 形を変えるブロックコポリマー : 可変な連結様式の構築と特性制御,高田十志和,東北ポリマー懇話会,岩手大学,2015.3.16-17, 盛岡市上田 (招待講演)
37. Toshikazu Takata, Intelligent Polymer Materials Characterized by Rotaxane Linkages, Yamaga University International Commemorative Symposium of GMAP Center, Yamagata University, 2015.1.23-24, Yonezawa(Invited talk)
38. Toshikazu Takata, "Intelligent Polymer Materials Characterized by Rotaxane Linkages", International Commemorative Symposium of GMAP Center (Completion for Research Center) –Joint Symposium of Research Center of NGAP(YUCOE(E)), 2015.1.22-24, 米沢(Invited talk)
39. 高田十志和, 超分子連結の新手法と動的性質の活用による新材料創製,高田十志和,特別講義,日本大学生産工学部,2014年12月19日, 千葉県習志野市 (特別講演)
40. 高田十志和, 超分子ポリマーの組織化とトポロジー変換,高田十志和,国際高等研究所講演会「分子基盤に基づく生体機能への搖らぎとダイナミックネットワークの解明」2014年度第1回研究会,国際高等研究所,2014年12月14-15日, 京都府木津川市 (招待講演)
41. 高田十志和, ロタキサン構造を架橋点とする新しい架橋高分子の構築法,高田十志和,エラストマー討論会,愛知工業大学,2014.12.04-05, 豊田市 (依頼講演)
42. 高田十志和, 高機能化を目指す高分子の合成と修飾,高田十志和,大阪工研協会ニューフロンティア材料部会第94回例会,KKRホテル大阪,2014年11月13日, 大阪市 (招待講演)
43. Toshikazu Takata, Thermo-responsive Hydro Gels Networked by Rotaxane Cross-links, Toshikazu Takata, Takayuki Arai, Keumhee Jang, and Yasuhito Koyama, Polymer Network and Gel 2014 (PN&G2014), University of Tokyo, 2014.11.10-14, Tokyo(Invited talk)
44. 高田十志和, 架橋と表面修飾による機能高分子材料の創製,第5回 次世代機能化学素材・接合技術研究会,岩手大学ものづくり基盤技術開発コンソーシアム,ホテルルイズ,2014.11.5, 盛岡市 (招待講演)
45. Toshikazu Takata, Chiral Rotaxane Catalysts for Asymmetric Reactions, Toshikazu Takata, MC Asia 2014, Peking University, 2014.10.29-31, Beijing(Invited talk)
46. Toshikazu Takata, Macrocyclic Catalyst for Highly Efficient Hydroamination via Cavity-Threading of Substrate as The Key Process, CRC International Symposium on *Synthesis and Applications of Functional Molecules and Materials Utilizing Biomolecules as a Motif*, Hokkaido University 2014.9.30-

- 10.1, Sapporo (Keynote lecture)
47. 高田十志和, 高分子の効率的修飾 – グラフト化,表面修飾,架橋 –,ゴム技術フォーラム調査委員会, 東京工業大学, 2014.9.18, 東京 (基調講演)
48. Toshikazu Takata, Click Polymer Synthesis: Polymerization and Polymer Modification Using Nitrile *N*-Oxides, Toshikazu Takata and Yasuhito Koyama, Polycondensation 2014, Tokyo Institute of Technology, 2014.09.8-11, Tokyo (Invited talk)
49. Toshikazu Takata, Daisuke Aoki, and Satoshi Uchida, Transformable Polymer Architectures Functioned by Movable Linkages- Role of Rotaxane Linkage for Polymer Topology Change -, High Tech Polymer Materials (HTPM) 2014, Chinese Academy of Sciences, 2014.7.1-4, Beijing (Keynote lecture)
50. 高田十志和, 高分子の高次構造精密解析を支える分光分析技術 - らせんポリマーの合成と構造 -,高田十志和,高分子学会関東支部第80回武藏野地区高分子懇話会, 日本分光, 2014.6.13, 八王子 (招待講演)
51. 高田十志和, ロタキサン化学 - 空間連結と動的特性が醸す特異機能の創出 -,高田十志和,東北大学理学部・理学研究科一般雑誌会講演会,2014.6.5, 仙台 (招待講演)
52. 高田十志和, ロタキサン触媒の特異機能 - 協同効果と0次反応 -,高田十志和,大阪大学基礎工学研究科講演会,2014.5.2, 大阪 (招待講演)

【立川仁典】

1. 立川仁典, 歯車状両親媒性分子からなるナノキューブの安定性に関する理論的研究, 第98回日本化学会春季年会, 2018.3.20, 船橋 (招待講演)
2. Masanori Tachikawa, "Ab initio study of the effect of molecular vibrations on the positron-binding to polyatomic molecules", 8th Asia-Pacific Conference of Theoretical and Computational Chemistry (APCTCC8), 2017.12.15-17, Mumbai, India (Invited talk)
3. Masanori Tachikawa, "Path Integral Simulation for Hydrogen bonded systems: Protomic quantum nature and its isotope effects", 3rd Japan-Korea Joint Symposium on Hydrogen in Materials, Kyushu University, 2017.11.23-24, Fukuoka (Invited talk)
4. Masanori Tachikawa, "Ab initio theoretical study of the effect of molecular vibrations on the positron-binding to polyatomic molecules", 12th International Workshop on Positron and Positronium Chemistry (PPC12), 2017.8.28-9.1, Lublin, Poland (Invited talk)
5. Takako Mashiko, Shuichi Hiraoka, Umpei Nagashima, and Masanori Tachikawa, "Theoretical study on substituent and solvent effects for nanocube formed with gear-shaped amphiphile molecules", The 9th Conference of the Asian Consortium on Computational Materials Science (ACCMS2017), 2017.8.9-11, Kuala Lumpur, Malaysia, (Invited talk)
6. Takako Mashiko, Shuichi Hiraoka, Umpei Nagashima, and Masanori Tachikawa, "Theoretical study on substituent and solvent effects for nanocube formed with gear-shaped amphiphile molecules", Frontier Bioorganization Forum 2017, Academia Sinica, 2017.4.24-25, Taipei, Taiwan (Invited talk)
7. Takako Mashiko, Shuichi Hiraoka, Umpei Nagashima, and Masanori Tachikawa, Theoretical study on substituent and solvent effects for nanocube formed with gear-shaped amphiphile molecules, The 5th International Symposium on Dynamical Ordering of Biomolecular Systems for Creation of Integrated Functions The University of Tokyo, Komaba, 2017.1.21-22, Tokyo (Invited talk)
8. Masanori Tachikawa, Path integral simulation on muoniated acetone radical, The 11th General Meeting of ACCMS-VO, Tohoku University, 2016.12.21, Miyagi (Invited talk)
9. Masanori Tachikawa, Path integral simulation on muoniated acetone radical, The International Symposium of Quantum Beam Science, Ibaraki University, 2016.11.19, Ibaraki (Invited talk)
10. Masanori Tachikawa, Path Integral Simulation for Hydrogen bonded systems: Protomic quantum nature and its isotope effects, Thai-Japan Symposium in Chemistry, Chiang Mai University, 2016.11.15, Thailand (Invited talk)
11. Masanori Tachikawa, Quantum Monte Carlo calculation of Positron-attached Polyatomic Molecules, 14th International Workshop on Slow Positron Beam Techniques & Applications (SLOPOS14), 2016.5.22-27, Matsue, Japan (Invited talk)
12. Masanori Tachikawa, The International Conference of Computational Methods in Sciences and Engineering 2016 (ICCMSE 2016), 2016.3.17-21, Athens, Greece, Metropolitan (Invited talk)
13. Masanori Tachikawa, The Seventh Asia-Pacific Conference of Theoretical and Computational Chemistry (APCTCC7), 2016.1.25-28, Kaohsiung, Taiwan (Invited talk)
14. Yuki Oba, Tsutomu Kawatsu, and Masanori Tachikawa, International USMM & CMSI Workshop, Hongo Campus, 2016.1.5-9, TOKYO, (Japan) (Invited talk)

15. Masanori Tachikawa, "TBA", Seventh Asia-Pacific Conference of Theoretical and Computational Chemistry (APCTCC7), 2016.1.25-28, Kaohsiung, Taiwan (Invited talk)
16. Masanori Tachikawa, PACIFICHEM 2015, The International Chemical Congress of Pacific Basin Societies 2015, 2015.12.15-20, Honolulu, Hawaii, (USA) (Invited talk)
17. Masanori Tachikawa, 6th JCS, International Symposium on Theoretical Chemistry, 2015.10.11-15, Smolenice Castle, Slovakia (Invited talk)
18. Masanori Tachikawa, "TBA", The 8th Conference of Asian Consortium on Computational Materials Science (ACCMS-8), 2015.6.16-18, Taipei, Taiwan (Invited talk)
19. Masanori Tachikawa, "First-principles calculation for positron binding to molecules", International Symposium of Computational Chemistry (ICCMSE 2015), 2015.3.20-23, Athens, Greece. (Invited talk)
20. Masanori Tachikawa, "First-principles calculation for positron binding to molecules", XIXth International Workshop on Quantum Systems in Chemistry and Physics (QSCP-XIX), 2014.11.11-17, Tamsui, Taipei, Taiwan (Invited talk)
21. Masanori Tachikawa, "Multi-component molecular theory for hydrogen-bonded systems and positronic compounds", 5th French-Japanese Workshop on Computational Methods in Chemistry, 2014.6.30-7.1, Strasbourg, France (Invited talk)

【田中良和】

1. 田中良和, スルメイカヘモシアニンの結晶構造解析と結晶中へのタンパク質包摂の取り組み, 日本たばこ産業セミナー, 2018.1.22, 横浜 (招待講演)
2. 田中良和, 膜孔形成毒素の分子機構とヘモシアニンの構造解析, 山形大学蛋白質科学セミナー, 2018.1.15, 米沢 (招待講演)
3. Yoshikazu Tanaka, Minghao Chen, Shun Narai, Naoki Omura, Naoki Shigi, Min Yao, Investigating a novel tRNA thiolational modification mechanism involving an [4Fe-4S] cluster, 2nd Joint International Symposium of NSRRC and IPR, 2017.12.6-7, Taipei, Taiwan (Invited talk)
4. 田中良和, 分子量 4MDa の巨大蛋白質会合体へモシアニンの結晶への蛋白質の包摂, JST-ACCEL プロジェクト「革新的分子構造解析」公開シンポジウム, 2017.10.31, 東京 (招待講演)
5. 田中良和, 分子量 4MDa の巨大蛋白質会合体へモシアニンの結晶への蛋白質の包摂
第 17 回 リング・チューブ超分子研究会シンポジウム, 2017.6.9-10, 札幌 (招待講演)
6. Yoshikazu Tanaka, Pore formation mechanism of staphylococcal pore forming toxin, Max Planck Institute of Molecular Physiology Department 3 Lab Retreat Seminar, 2016.10.9-12, Germany (Invited talk)
7. Yoshikazu Tanaka, Molecular mechanism of staphylococcal pore forming toxin, Hokkaido University / Pusan National University Joint Symposium 2016, 2016.3.8-9, Busan, Busan, Korea (Invited talk)
8. 田中良和, X 線結晶構造解析により明らかになった黄色ブドウ球菌の膜孔形成毒素蛋白質の膜孔形成機構, 生理学研究所 研究会「膜システムの機能的・構造的統合」, 2015.9.1-2, 岡崎 (招待講演)
9. 田中良和, 山下大智, 菅原宇希, 金子淳, 神尾好是, 姚閔, Molecular mechanism of pore forming toxin from *Staphylococcus aureus*, 第 15 回日本蛋白質科学会年会, 2015.6.24-26, 徳島市 (招待講演)
10. 田中良和, 山下大智, 菅原宇希, 金子淳, 神尾好是, 田中勲, 姚閔, 結晶構造解析により明らかになった黄色ブドウ球菌由来膜孔形成毒素の分子機構, 毒素シンポジウム, 2014.9.3-5, 鳴門市 (招待講演)

【寺嶋正秀】

1. Masahide Terazima, Reaction Dynamics of Photo-induced Protein-DNA Interaction
Trombay Symposium on Radiation & Photochemistry, 2018.1.3-7, Mumbai, India (Invited talk)
2. Masahide Terazima, Time-resolved diffusion technique can detect conformation dynamics of photosensor proteins, Asia Oceania Conference on Photobiology, 2017.11.12-15, Seoul, Korea (Invited talk)
3. Masahide Terazima, Light induced heating and volume changes to reveal protein reactions
9th International Conference on Photoacoustic and Photothermal Phenomena, 2017.7.16-20, Bilbao, Spain (Invited talk)
4. Masahide Terazima, Transient fluctuations during protein reactions detected by time-resolved thermodynamics, 9th Korea-Japan Seminars on Biomolecular Science, 2016.11.14-16, Gyeongju, Korea
5. Masahide Terazima Advanced light sensing natural proteins that can detect light intensity, Asia Pacific Society for Materials Research, Sun Moon Lake, 2016.8.11-14, Taiwan
6. Masahide Terazima A biosensor to detect inter-protein interaction and domain-domain interaction, International Conference on Small Science, 2016.6.25-29, Prague, Czech Republic
7. Masahide Terazima, Spectrally Silent Dynamics of Photo-induced Protein-Protein Interaction Change

- Trombay Symposium on Radiation & Photochemistry & 6th Asia Pacific Symposium on Radiation Chemistry, 2016.1.5-9, Mumbai, India
8. Masahide Terazima, How to detect photoreactions of proteins: Blue light sensors, International Workshop on Radiation and Photochemistry, 2016.1.10-12, Pune, India
 9. Masahide Terazima, Conformation changes and water-protein interaction change for function of proteins, PACIFICHEM 2015, 2015.12.15-20, Hawaii.
 10. Masahide Terazima, Time-resolved detection of inter-protein interaction during protein reaction, PACIFICHEM 2015, 2015.12.15-20, Hawaii
 11. Masahide Terazima, Time-resolved detection of light induced heat during protein reactions, 18th International Conference on Photoacoustic and Photothermal Phenomena, 2015.9.6-10, Novi Sad, Serbia (全体講演)
 12. Masahide Terazima, Compressibility and reactivity of a protein reaction, 19th Symposium on Thermophysical Properties, Boulder, 2015.6.21-26, Colorado U.S.A.
 13. Masahide Terazima, Sensing of Protein Reactions using pulsed laser based transient grating, Frontiers in Optics 2014/Laser Science XXX (FiO/LS) meeting, 2014.10.19-23, Tucson, USA
 14. Masahide Terazima, Photosensing reaction dynamics of flavin proteins: Phototropin, 16th International Conference on Retinal protein, Nagahama Royal Hotel, Japan, 2014.10.5-10
 15. Masahide Terazima, Conformation change of a flavoprotein for signal transfer: phototropins, 18th International Flavin Symposium, 2014.7.27-8.1, Thailand
 16. Masahide Terazima, Time resolved measurements of thermodynamic properties during chemical reactions of bio-molecules, Thermophysical and mechanical properties of advanced materials, 2014.6.12-15, Cesme-Izmir,Turkey (Keynote lecture)
 17. Masahide Terazima, Time-resolved studies on protein reactions toward biological function, The Second KYOTO-BRISTOL Symposium, 2014.1.9-10, Kyoto, Japan
 18. Masahide Terazima, Spectrally Silent Dynamics of Photo-induced Protein-Protein Interaction, Korean-Japan Bilateral Symposium on Frontier Photoscience, 2013.11.24-27 (Invited talk)

【内藤晶】

1. Akira Naito, Photoirradiation and microwave irradiation NMR spectroscopy. Application to photoreceptor retinal proteins and liquid crystals, Taiwan-Japan Biomedical Symposium on Magnetic Resonance, 2017.10.15-17, Tainan, Taiwan (Invited talk)
2. Akira Naito, Microwave heating mechanism of liquid crystals as revealed by microwave irradiation NMR spectroscopy, Magnetism and Magnetic Materials, 2017.10.9-10, London, UK (Invited talk)
3. Akira Naito, Pressure and photo induced structural changes of photoreceptor proteins as revealed by solid-state NMR, IMBP2017, 2017.8.20-24, Kyoto, Japan (Invited talk)
4. 内藤晶, マイクロ波および光照射固体NMRの開発と液晶および光受容膜タンパク質への応用, 第 61 回 固体 NMR・材料フォーラム,理化学研究所,2017.5.12, 横浜 (招待講演)
5. Akira Naito, Microwave irradiation solid-state NMR spectroscopy to reveal microwave heating and energyflow mechanism of liquid crystals, The 7th Asia Pacific NMR Symposium, 2017.2.16-19, Bangalore, India (Invited talk)
6. Akira Naito, High resolution solid-state NMR spectroscopy. Development and application to structural biochemistry, 第 55 回 NMR 討論会, 2016.11.16-18, 広島 (特別講演)
7. Akira Naito, Y. Makino, K. Oshima, Y. Otani, A. Shigeta, I. Kawamura, T. Okitsu, A. Wada, S. Tuzi,, T. Iwasa, Photoreaction pathways of bacteriorhodopsin and its mutants as revealed by in-situ photo-irradiation solid-state NMR, ICRP2016, 2016.10.2-7, Potsdam, Germany (口頭発表)
8. T. Nagao, D. Mishima, N. Javkhaltugs, J. Wang, D. Ishioka, K. Yokota, K. Norisada, I. Kawamura, K. Ueda, A. Naito, Structure and orientation of antibiotic peptide alamethicin in membrane environment as revealed by chemical shift oscillation analysis of solid state NMR and MD simulation, ICBRBS2016, 2016.8.21-26, Kyoto (Invited talk)
9. Akira Naito, Photoreaction pathways of photoreceptor membrane proteins revealed by in-situ photo-irradiation solid-state NMR, 2015 International Chemical Congress of Pacific Basin Society (Pacificchem 2015), 2015.12.15, Honolulu, Hawaii, (USA) (Invited talk)
10. Akira Naito, Yoshiteru Makino, Hiroki Yomoda, Yuya Tomonaga, Tetsuro Hidaka, Izuru Kawamura, Yuki Sudo, Akimori, Wada, Takashi Okitsu, Naoki Kamo, Determination of photo-intermediates and photo-reaction pathways of photoreceptor membrane proteins by in-situ photo-irradiation solid-state

NMR, The 6th Asia-Pacific NMR Symposium, 2015.8.14, Hong Kong, China (Invited talk)

11. 内藤晶, 固体 NMR による抗菌ペプチドの生体膜結合構造と膜相互作用解析
第 15 回 日本蛋白質科学会年会, 2015.6.26, 徳島 (招待講演)
12. 内藤晶, 伊藤-渡邊ひかり, 上平-石島美弥, N. Lavkhantugs, 近藤正志, 中越雅道, 川村出, 辻暁, 斎藤肇, 上田一義, “固体 NMR と TEM によるヒトカルシトニンにおけるアミロイド線維形成と阻害機構の解明”, 第 53 回 NMR 討論会, 大阪大学コンベンションセンター, 2014.11.4-6, 大阪 (招待講演)
13. 内藤晶, 横野義輝, 重田安里寿, 四方田博紀, 川村出, 沖津貴志, 和田昭盛, 須藤雄氣, 加茂直樹, “光照射固体 NMR による光受容膜タンパク質の光反応経路の解明” 第 87 回日本生化学会大会, 2014.10.15-18, 国立京都国際会館, 京都 (招待講演)
14. A.Naito, Y. Makino, H. Yomoda, Y. Tomonaga, T. Hidaka, I. Kawamura, Y. Sudo, A. Wada, T. Okitsu, N. Kamo, “detection of photo-intermediates of microbial sensory rhodopsins by in situ photo-irradiation solid-state NMR”, 16th International Conference on Retinal Proteins, 2014.10.5-10, Nagahama, Japan (Invited talk)

【東雅大】

1. 東雅大, 凝縮系の励起状態反応ダイナミクスの定量的理解を目指して, 第 32 回量子系分子科学研究セミナー, 2018.2.20, 神戸 (招待講演)
2. 東雅大, 凝縮系の反応ダイナミクスを記述可能な計算手法の開発と応用, 北大理論化学研究会 : 実践理論化学の最前線, 2017.11.16, 札幌 (招待講演)
3. 東雅大, 高効率ポテンシャル関数生成手法による光捕集アンテナ中の色素の励起エネルギーの大きさと揺らぎの定量的評価, レア・イベントの計算科学, 2017.8.30, 熱海 (招待講演)
4. Masahiro Higashi, Toward quantitative understanding of excitation energy transfer in light-harvesting complexes, The 9th Conference of the Asian Consortium on Computational Materials Science, 2017.8.11, Kuala Lumpur, Malaysia. (Invited talk)
5. Masahiro Higashi, Theoretical investigation of excited-state reactions and properties in condensed phases, 前瞻跨領域基礎科學中心 尖端生物分子探測 専題演講, 2017.8.7, 新竹市, 台湾 (Invited talk)
6. Masahiro Higashi, Theoretical analysis of regio- and stereoselectivity of new organic reactions, International Symposium on Pure & Applied Chemistry 2017, 2017.6.9, Ho Chi Minh City, Vietnam (Invited talk)
7. Masahiro Higashi, Theoretical investigation of excited states of pigments in the light-harvesting antenna, Kickoff To Explore The New Field Of Molecular Soft Matter, 2017.3.14, Bangkok Thailand (Invited talk)
8. 東雅大, 新規有機反応における立体・位置選択性の理論的解明, IQCE 講演会「量子化学で探る化学の最先端 2016」, 2016.11.19, 東京 (招待講演)
9. Masahiro Higashi, Theoretical analysis of site energies and their fluctuations of pigments in the light-harvesting antenna, The 9th Korea-Japan Seminars on Biomolecular Science, 2016.11.14, Gyeongju, Korea (Invited talk)

【松村浩由】

1. 松村浩由, 細胞分裂タンパク質の構造研究, 大阪大学蛋白質研究所セミナー SPring-8 先端利用技術ワーカーショップ 「SPring-8 における蛋白質構造生物学研究の現状と将来」, 2017.8.3, Osaka (招待講演)
2. 松村浩由, 創薬に貢献するタンパク質結晶化技術, メディカルジャパン, インテックス大阪, 2017.2.15, 大阪市住之江区
3. 松村浩由, 離合集散する柔らかいタンパク質の構造機能解析, 大阪大学蛋白質研究所セミナー 「構造を基盤とする蛋白質科学における未解決問題」, 2016.3.1, 東京
4. 松村浩由, 細菌の細胞分裂ダイナミクスの構造機能相関解析, 分子基盤に基づく生体機能ネットワークとダイナミクスの解明, 第 1 回研究会, 国際高等研究所, 2014.12.14, 京都府木津川 (招待講演)
5. 松村浩由, 代謝経路における動的秩序, 第 1 回「動的秩序と機能」若手研究会, 2014.9.29, 蔵王, 宮城 (招待講演)

【松森信明】

1. Nobuaki Matsumori, Integrated analysis of membrane systems, International seminar on biophysics and chemical biology of biomembrane and lipid bilayers, 2017.10.9, Osaka (Invited talk)
2. Nobuaki Matsumori, NMR and Fluorescent Studies of Lipid Rafts, 9th Korea-Japan Seminars on Biomolecular Sciences: Experimental and Simulations. 2016.11.14, Korea (Invited talk)
3. 松森信明, 木下祥尚, 脂質ラフト形成基盤の解明を目指したスフィンゴミエリンの動的および NMR 構造解析, 日本化学会第 96 春季年会, 2016.3.24, 京田辺 (招待講演)

4. 松森信明, 脂質膜における動的挙動, 構造および相互作用解析, 蛋白研セミナ一生体膜上の生物化学, 2016.3.3, 吹田 (招待講演)
5. Nobuaki Matsumori, NMR studies of lipid rafts, Pacifichem2015, 2015.12.16, Honolulu, (Invited talk)
6. Nobuaki Matsumori, Small molecule interactions with membrane sterol, Pacifichem 2015, 2015.12.17, Honolulu (Invited talk)
7. 松森信明, 脂質ラフトにおける脂質分子の動的挙動および構造解析, 第 88 回日本生化学会大会フォーラム, 2015.12.3, 神戸 (招待講演)
8. 松森信明, 脂質膜の統合分析, 第 33 回九州コロイドコロキウム, 2015.11.21, 福岡 (招待講演)
9. 松森信明, 村田道雄, 脂質ラフトの NMR 解析, 第 53 回日本生物物理学会年会, 2015.9.14, 金沢(招待講演)
10. 松森信明, NMR による生体膜ならびに膜作用生理活性物質の解析, 日本生化学会大会, 2014.10.15, 京都 (招待講演)

【養王田正文】

1. Masafumi Yohda, Assembly of encapsulin nanocompartment from *Rhodococcus erythropolis* N771 and encapsulation of a guest protein, THE 15TH INTERNATIONAL CONFERENCE on QiR (Quality in Research), 2017.7.26, Bali, Indonesia(Invited talk)
2. 養王田正文, Protein folding mechanism of group II chaperonin, 第 44 回 生体分子科学討論会, 2017.6.23, 秋田 (招待講演)
3. Masafumi Yohda, Functional expression of olfactory receptors in yeast for the development of odorant sensor, International Symposium on Biomeomedical Engineering 2016, 2016.5.31, Indonesia (Invited talk)
4. Masafumi Yohda, Hiroshi Sekiguchi,Ayumi Nakagawa,Yohei Yamamoto,Toshihiko Oka,Kunihiro Kuwajima,Yuji C. Sasaki, Dissection of the ATP-dependent conformational change cycle of group II chaperonin, Japan-Italy Joint Symposium, New Trends in Science and Engineering of Enzyme and Microbiology for Sustainable Society, 2014.11.6, Nara (Invited talk)
5. 養王田正文, シークエンス技術の過去・現在・未来, 2014 年度生物工学フォーラム「先端技術による新たなバイオテクノロジー」, 2014.7.25, 埼玉 (招待講演)

A02 : 動的秩序の創生

【新井亮一】

1. 新井亮一, タンパク質ナノブロック戦略によるネオ・レクチン超分子複合体の創出を目指して, 生命科学系合同年次大会 (ConBio2017), 2017.12.6, 神戸 (招待講演)
2. 新井亮一, 人工タンパク質ナノブロックによる超分子複合体ナノ構造の創出, 第 6 回 CHUBU 懇話会, 2017.9.29, 長野 (招待講演)
3. 新井亮一, 人工タンパク質ナノブロックによる超分子複合体ナノ構造の創製, バイオ工学シンポジウム, 2017.7.3, 松山 (招待講演)
4. 木村尚弥, 小林直也, 新井亮一, タンパク質ナノブロックによる動的秩序構造形成
第 17 回日本蛋白質科学会年会, 2017.6.20, 仙台 (招待講演)
5. 新井亮一, 人工タンパク質ナノブロック(PN-Block)による自己組織化超分子ナノ構造複合体の創出, 第 17 回リング・チューブ超分子研究会シンポジウム, 2017.6.10, 札幌 (招待講演)
6. 木村尚弥, 小林直也, 新井亮一, タンパク質ナノブロックによる動的秩序構造形成, 日本化学会第 97 春季年会, 2017.3.16, 横浜 (招待講演)
7. Ryoichi Arai, Self-assembling supramolecular nanostructures created from *de novo* protein nanobuilding blocks, 11th Annual Symposium on Nanobiotechnology 2017, 2017.2.28, Kawasaki, Japan (Invited talk)
8. 新井亮一, 人工タンパク質ナノブロック (PN-Block) による自己組織化超分子ナノ構造複合体の創製：“ネオバイオ分子”創出へ向けて,平成 28 年度第 3 回富山大学テニュアトラックセミナー, 2017.2.3, 富山 (招待講演)
9. 小林直也, 木村尚弥, 新井亮一, 人工蛋白質ナノブロックによる多様な自己組織化超分子ナノ構造複合体：ネオバイオ分子の創生を目指して, 第 39 回日本分子生物学会年会, 2016.12.2, (招待講演)
10. 小林直也, 木村尚弥, 新井亮一, 人工タンパク質をブロックに見立てた超分子ナノ構造複合体の設計構築, 第 54 回日本生物物理学会年会, 2016.11.25, つくば (招待講演)
11. Naoya Kimura, Naoya Kobayashi, Ryoichi Arai, Self-assembling supramolecular nano-architectures created from *de novo* protein nano-building blocks, 第 68 回日本生物工学会大会, 2016.9.29, 富山 (Invited talk)
12. 新井亮一, タンパク質ナノブロックによる自己組織化超分子複合体の創製, 第 48 回若手ペプチド夏の勉強会, 2016.8.1, 八王子, 東京都 (招待講演)

13. 小林直也,木村尚弥, 新井亮一, Self-assembling supramolecular nanostructures created from *de novo* protein nanobuilding blocks, 分子研研究会 超機能分子の創成, 2016.6.28, 岡崎, 愛知県 (招待講演)
14. 小林直也,木村尚弥,新井亮一, 人工蛋白質ナノブロック(PN-Block)による自己組織化ナノ構造創製：“ネオバイオ分子” 創出へ向けて 第 16 回日本蛋白質科学会年会,2016.6.9, 福岡 (招待講演)

【飯野亮太】

1. Ryota Iino, Single-molecule dynamics of natural and engineered molecular motors
The 5th International Symposium on "Dynamical Ordering & Integrated Functions, 2017.1.21-22, Japan(Invited talk)
2. Ryota Iino, Watching dynamic motions of biological molecular machines, 7th RIES-Hokudai International Symposium, 2016.12.12-13, Japan(Invited talk)
3. 飯野亮太, マイクロ・ナノデバイスを用いた1分子・1細胞ナノバイオ計測, MNC2016 技術セミナー「マイクロ・ナノバイオ技術の最前線」, 2016.11.8, 京都 (招待講演)
4. Ryota Iino, Intermediate states during the stepping motion of kinesin-1 revealed by high-speed single-molecule imaging with gold nanoprobe, 4th Kanazawa Bio-AFM Workshop, 2016.10.16, Japan (Invited talk)
5. 飯野亮太, 金ナノプローブで生体分子の速いダイナミクスを観る, 第 25 回バイオイメージング学会学術集会シンポジウム, 2016.9.6, 名古屋 (招待講演)
6. Ryota Iino, Direct observation of intermediate states during the stepping motion of kinesin-1
Biophysical Society Thematic Meeting: Engineering Approaches to Biomolecular Motors: From *in vitro* to *in vivo*, 2016.6.15, Canada (Invited talk)
7. Ryota Iino, Direct observation of intermediate states during the stepping motion of kinesin-1
8th Japan-Korea Seminars on Biomolecular Science: Experiments and Simulation, 2016.2.15, Aichi (Japan) (Invited talk)
8. Ryota Iino, Single-molecule analysis of new molecular motors hydrolyzing crystalline polysaccharides, Pure and Applied Chemistry International Conference 2015 (PACCON2015), 2016.2.10, Bangkok (Thailand) (Invited talk)
9. Ryota Iino, Dynamics of linear and rotary molecular motors revealed by gold nanoprobe
Pacificchem 2015, Technical Symposia. Physical, Theoretical & Computational: Interplay between Chemistry and Dynamics in Biomolecular Machines, 2015.12.16, Hawaii (USA) (Invited talk)
10. Ryota Iino, Single-molecule high-speed imaging analysis of ATP-driven molecular motors,
第 53 回生物物理学会年会シンポジウム「Formation of spatiotemporal dynamic ordering mediated by ATP hydrolysis」, 2015.9.14, Kanazawa (Ishikawa) (Invited talk)
11. Ryota Iino, High speed single-molecule measurement of conformational dynamics of molecular motors probed by gold nanorod, International Symposium on "Studying the Function of Soft Molecular Systems", 2015.7.10, Tokyo (Japan) (Invited talk)
12. Ryota Iino, Dynamic motions of individual molecular motors, IMS Asian International Symposium "Supramolecular Dynamics at the interface of Chemistry and Biology", 2015.6.12, Aichi, Japan (Invited talk)
13. 飯野亮太, 回転型 ATPase によるイオンの輸送を考える, 分子研研究会「膜タンパク質内部のプロトン透過を考える」, 岡崎コンファレンスセンター, 2015.4.21, 岡崎 (招待講演)
14. 飯野亮太, 生体回転超分子モーターの作動メカニズム, 2015.3.26, (招待講演)
15. 飯野亮太, 金ナノプローブで探る生体分子モーターのダイナミクス, 日本化学会 第 95 春季年会 中長期企画 シンポジウム. 船橋 (日本大学) 2015.3.26-29 (招待講演)
16. Ryota Iino, Watching dynamic motions of individual molecular motors with gold nanoprobe
Pure and Applied Chemistry International Conference 2015 (PACCON2015), 2015.1.22, Bangkok, Thailand (Invited talk)
17. 飯野亮太, 生体分子モーターを測る, 壊す, 創る, シンポジウム「細胞のメゾスケール構造機能」, 京都大学, 2014.12.13, 京都 (招待講演)
18. 飯野亮太, 金ナノ粒子, 金ナノロッドを用いた生体分子モーターのマイクロ秒 1 分子計測
新学術領域「柔らかな分子系」第 7 回ワークショップ, 岡崎コンファレンスセンター, 2014.12.12, 岡崎市 (招待講演)
19. Ryota Iino, Single-molecule imaging analysis of molecular motors, The 7th Korea-Japan Seminars on Biomolecular Sciences, 2014.11.27, Seoul, Korea (Invited talk)
20. 飯野亮太, 生体分子モーターダイナミクスの 1 分子計測: 構造解析と理論予測との協奏を目指して, TCCI 第

- 5回研究会, 岡崎コンファレンスセンター, 2014.10.17-18, 岡崎市 (招待講演)
21. 飯野亮太, Importance of membrane pumps and channels: an introduction, 第 52 回生物物理学会年会シンポジウム "Which is important for biophysicists, pump or channel?", 札幌コンベンションセンター, 2014.9.25-27, 札幌 (招待講演)
 22. 飯野亮太, 1 分子計測技術で生体分子マシン, 人工分子マシンの“動き”を調べる・考える
第 63 回高分子討論会 S14. “動き”のある自己組織化材料: 動的応答・変化を示す材料の設計・機能・応用の最前線, 長崎大学, 2014.9.24, 長崎 (招待講演)
 23. Ryota Iino, "Motions of individual biomolecular motors probed by gold nanoparticle and nanorod International Symposium on Small Particles and Inorganic Clusters (ISSPIC XVII). 2014.9.10, Fukuoka, Japan (Invited talk)
 24. 飯野亮太, 細菌多剤排出ポンプ計測マイクロデバイス, 第 9 回トランスポーター研究会年会シンポジウム 「チャネル・トランスポーター研究の最前線 2 (機能評価系・創薬)」, 名古屋市立大学 2014.6.14, 名古屋 (招待講演)
 25. 飯野亮太, DNA を巻き取る分子リール, 第 66 回日本細胞生物学会大会シンポジウム「遺伝情報を司る DNA のふるまい」, 奈良県新公会堂, 2014.6.12, 奈良 (招待講演)

【井上雅彦】

1. Yuki Ohishi, Tetsuhiro Yoneda, Kentaro Masuda, Hajime Abe, Masahiko Inouye, Development of Cage-Shaped and Three-Armed Molecules with Pyridine–Acetylene–Phenol Units as Artificial Saccharide Receptors, The Second International Symposium on Toyama–Asia–Africa Pharmaceutical Network 2017.9.25–26, Jinan, China (Invited talk)
2. 井上将彦, タンパク代用物としての短鎖ヘリカルペプチドの創出, 第 34 回 有機合成化学セミナー, 2017.9.17-20, 金沢 (招待公演)
3. 井上将彦, 千葉順哉, アルキニルヌクレオチドからなる人工 DNA の創出と核酸関連酵素反応への展開, 日本化学会第 97 春季年会, 2017.3.16, 横浜 (特別企画講演)

【上野隆史】

1. Takafumi Ueno, "Design of Bioinorganic Materials Constructed by Coordination Chemistry" 6th Asian Conference on Coordination Chemistry, 2017.7.25, Melbourne, Australia (Invited talk)
2. Takafumi Ueno, "CO Releasing Bioinorganic Nanomaterials for CO Gas Biology", The International Advanced Drug Delivery Symposium 2017, 2017.4.6, Taipei (Invited talk)
3. 上野隆史, T4 ファージ由来蛋白質針による生体膜透過の動的秩序機構第 97 回日本化学会春季年会 特別企画「ハイブリッド自己組織化: 秩序形成における生命系と人工系の接点」慶應大学, 2017.3.16, 横浜 (特別企画講演)
4. 上野隆史, 高分子的設計で挑むタンパク質集合体機能の化学第 51 回高分子学会北海道支部研究発表会, 北海道大学, 2017.1.19, 北海道, 札幌 (Keynote 講演)
5. Takafumi Ueno, "In vivo functions of organometallopeptides", 5th Symposium on Advanced Biological Inorganic Chemistry, 2017.1.8, Kolkata (Invited talk)
6. Takafumi Ueno, "In Vivo and In Vitro Bioinorganic Functions Designed within Ferritin Cage", 8th Asian Biological Inorganic Chemistry Conference, 2016.12.6, Auckland (Invited talk)
7. Takafumi Ueno, "Multi-bioinorganic Functions Designed within Protein Cage", Japan-Korea-Taiwan Bioinorganic Chemistry Symposium 2016, 2016.9.30, Okazaki (Invited talk)
8. Takafumi Ueno, "Biosupramolecular materials designed by protein assemblies", 2016 International Symposium on Polymer and Related Materials, Harbin Engineering University, 2016.7.10, Harbin, China (Invited talk)
9. Takafumi Ueno, "Protein Crystals for Designing Biohybrid Solid Materials", 8th Japan-Korea Seminars on Biomolecular Science: Experiments and Simulation, IMS, 2016.2.17, Okazaki, (Invited talk)
10. Takafumi Ueno, "Design of Protein Assemblies toward Biohybrid Solid Materials", The 8th WPI-AIMR Joint Seminar, AIMR, Tohoku Univ. 2016.1.29, Miyagi (Invited talk)
11. Takafumi Ueno, "CO Releasing Proteins as Bioinorganic Tools for CO Gas Biology" Frontiers of Iron Chemistry in Biology (#268) / Pacificchem 2015, Regency I (Royal Hawaiian), 2015.12.16, Hawaii (Invited talk)

12. 上野隆史, 生体固体材料としてのタンパク質結晶, 第25回日本MRS_自己組織化材料とその機能XIII, 波止場会館, 2015.12.8, 横浜 (基調講演)
13. 上野隆史, バイオミネラル反応を基軸とする金属蛋白質の機能設計, 第 10 回バイオミネラリゼーションワークショップ, 2015.12.6, 東京大学 (基調講演)
14. Takafumi Ueno, "Metal Carbonyl-Protein Materials for Carbon Monoxide Delivery in Living Cells" 10th China-Japan Joint Symposium on Metal Cluster Compounds, 2015.10.25, Fuzhou, China, (Invited talk)
15. Takafumi Ueno, "CO-releasing Organometallics in Supramolecular Proteins", 17th International Conference on Biological Inorganic Chemistry, 2015.7.23, Beijing (Invited talk)
16. Takafumi Ueno, "Protein-based Supramolecular Coordination Materials", Supramolecular Dynamics at the Interface of Chemistry and Biology, IMS, 2015.6.13, Okazaki (Invited talk)
17. Takafumi Ueno, "Bioactive Organometallics in Supramolecular Proteins", The 2nd CMS International Symposium– Cutting-edge Technologies of Biomaterials and Bio-relatedMaterials Based on Well-defined Molecular Systems –, Tsinghua Univ., 2015.6.6, Beijing (Invited talk)
18. Takafumi Ueno, "Protein-based Supramolecular Coordination Materials", The 2nd CMS International Symposium– Cutting-edge Technologies of Biomaterials and Bio-relatedMaterials Based on Well-defined Molecular Systems –, Kyushu Univ., 2015.1.20, Kyushu Univ. Fukuoka, Japan (Keynote lecture)
19. Takafumi Ueno, "Designing Dynamic Function of Protein Assembly", The 3rd International Symposium on Dynamical Ordering of Biomolecular Systems for Creation of Integrated Functions, Nemunosato, 2015.1.11, Mie, Japan
20. Takafumi Ueno, "Bioinspired Inorganic Materials of Protein Assemblies", 7th Asian Biological Inorganic Chemistry Conference, Gold Coast, 2014.12.2, Australia,
21. Takafumi Ueno, "Bioactive Organometallics in Supramolecular Proteins", The 2nd Japan-UK Joint Symposium on Coordination Chemistry, Chuo Univ., 2014.9.18, Tokyo
22. Takafumi Ueno, "Design of Confined Space of Protein Assembly", The 3rd International Supramolecular System Symposium, 2014.8.26, Changchun, China (Keynote Lecture)

【片山勉】

1. 片山勉, 大腸菌をモデルとした染色体複製開始の分子機構と制御システムに関する研究, 日本遺伝学会第 89 回大会, 2017.9.14, 岡山 (木原賞受賞講演)
2. 片山勉, 大腸菌の複製開始複合体の分子構築と機能メカニズム, 2016 年度 国立遺伝学研究所研究会「単細胞システム細胞装置のダイナミズム」, 2017.3.27, 三島市
3. 片山勉, 大腸菌染色体の複製開始と分配の制御における不連続複製の役割, 岡崎フラグメント 50 周年シンポジウム 「DNA 複製の過去・現在・未来--ゲノム複製からエピゲノム複製へ」, 2016.12.21, 名古屋市 (招待講演)
4. 崎山友香里, 野口泰徳, 川上広宣, 片山勉, 大腸菌染色体の複製起点 *oriC* 上で形成される DnaA 多量体の機能動態の解明, 日本遺伝学会第 88 回大会 プレナリーワークショップ --昨年の BP 賞受賞講演から, 2016.9.8, 三島市 (招待講演)
5. 片山勉, 大腸菌の染色体複製起点 *oriC* における DnaA 複合体のダイナミクス, 日本進化学会 第 18 回東京大会 ワークショップ「生物進化に伴う DNA 複製装置の Dynamics」, 2016.8.27, 東京都目黒区 (招待講演)

【神谷由紀子】

1. 神谷由紀子, 人工核酸による RNA 干渉機構の理解と制御, 分子研シンポジウム 2016, 2016.5.27, 岡崎(招待講演)
2. 神谷由紀子, 人工核酸を用いた RNA 干渉機構の理解と制御法の開発, 第 160 回名古屋市立大学薬学談話会, 2016.5.11, 名古屋(招待講演)
3. Yukiko Kamiya, Fluorescent monitoring of intracellular trafficking of siRNA, 9th International Symposium on Nanomedicine (ISNM2015), 2015.12.12, Mie, Japan(Invited talk)
4. 神谷由紀子, 糖骨格を持たない人工核酸を用いた RNAi 機構の理解と高性能 siRNA の開発 第 13 回 糖鎖科学コンソーシアムシンポジウム 一糖鎖研究と多領域との統合一, 2015.10.19, 名古屋 (招待講演)
5. 神谷由紀子, 光応答性 DNA デバイスを用いた遺伝子発現を制御する人工システムの開発 国際高等研究所 研究プロジェクト「分子基盤に基づく生体機能への揺らぎとダイナミックネットワークの

解説」2014年度第1回研究会, 2014.12.14, 京都 (招待講演)

6. Yukiko Kamiya, Photoregulation of gene expression with functional oligonucleotide involving azobenzene derivatives, 7th Korea-Japan Seminars on Biomolecular Sciences:Experiments and Simulations, KIAS, 2014.11.26, Korea
7. 神谷由紀子,伊藤杏奈,高井順矢,伊藤浩,村山恵司,樋田啓,浅沼浩之, 非環状骨格を持つ人工核酸の導入による siRNA の高機能化, アンチセンス・遺伝子・デリバリーシンポジウム 2014.9.9 (招待講演)

【佐藤宗太】

1. 佐藤宗太, リチウムイオン電池の高容量負極材料 : ナノチャンネルをもつ大環状芳香族分子の結晶, 第 103 回 新電池構想部会, 2018.4.24, 大阪 (招待講演)
2. Sota Sato, "Chemistry of Cyborg Supramolecules: Approaches to Biomolecular Science Taking Advantage of Artificial Biomolecular Interfaces", The Chemical Society of Japan, The 98th Annual Meeting, Nihon University, 2018.3.22, Funabashi City (Invited talk)
3. Sota Sato, "Development of Dynamical Ordering of Artificial Molecules by Mimicking Biomolecular Systems", Dynamical Ordering of Biomolecular Systems for Creation of Integrated Functions The 6th International Symposium, Hamamatsu Act Tower, 2018.1.21, Shizuoka (Invited talk)
4. 佐藤宗太, "タンパク質を丸ごと閉じ込める巨大カプセル分子の化学", SEST2017 ミニシンポジウム「超分子化学の先端物質科学」, 東京工業大学, 2017.11.4, 東京都 (招待講演)
5. 佐藤宗太, "動的秩序化により合成したサイボーグ超分子", 日本化学会 第 97 春季年会, 慶應義塾大学, 2017.3.16, 横浜市 (招待講演)
6. Sota Sato, "Biomimetic Spherical Complexes Constructed by Dynamical Ordering", (招待講演) Symposium on Chemistry and Materials 2017, Institute of Multidisciplinary Research for Advanced Materials, Tohoku University, 2017.3.7, Sendai (Invited talk)
7. 佐藤宗太, "これならわかる「NMR を用いた反応追跡 2 実際の応用例」", 日本化学会第 6 回 CSJ 化学フェスタ 2016, タワーホール船堀, 2016.11.14-16, 東京都 (招待講演)
8. 佐藤宗太, "球状錯体の『中』の化学 : 世界最小のフルオラス溶媒からの展開", フルオラス科学研究会第 9 回シンポジウム, 名古屋大学, 2016.10.7, 名古屋市 (招待講演)
9. Sota Sato, "Carbon-rich active materials with macrocyclic nanochannels for high-capacity negative electrodes in all-solid-state lithium rechargeable battery", 平成 28 年度化学系学協会東北大会, いわき明星大学, 2016.9.10-11, いわき市
10. Sota Sato, "Biological Functions Implanted in Artificial Supramolecules", 分子研研究会 日韓生体分子科学セミナー, 岡崎統合バイオサイエンスセンター 2016.2.15-17, 岡崎市
11. 佐藤宗太, "巨大分子の精密合成 ~きちんと分子構造を押さえる~", ブルカー・ダルトニクス MS サミット, TKP ガーデンシティ横浜, 2015.11.26, 横浜市 (招待講演)
12. 佐藤宗太, "Hierarchically Self-Assembled Metal-Organic Complexes as Fusion Materials", Fusion Materials Symposium 2015, 東京大学, 2015.9.19, 東京都 (招待講演)
13. 佐藤宗太, "大環状芳香族分子の合成とデバイスへの応用", 日本薬学会東北支部主催 第 14 回化学系若手研究者セミナー, 東北大大学薬学部, 2015.9.5, 仙台市
14. 佐藤宗太, "磁場配向性錯体の自己組織化合成と NMR 解析への展開"分子研研究会「金属イオンをもちいた超構造体の構築と機能発現 -配位化学における境界領域研究の現状と展望-」, 自然科学研究機構岡崎コンファレンスセンター, 2015.3.15, 岡崎市 (招待講演)
15. 佐藤宗太, "高速回転する分子ベアリング", 国際高等研研究プロジェクト「分子基盤に基づく生体機能への搖らぎとダイナミックネットワークの解明」, 2014 年度第 1 回(通算第 3 回)研究会 (京都), 国際高等研究所, 2014.12.15, 木津川市 (招待講演)
16. 佐藤宗太, "巨大な中空球状錯体の自己組織化合成", 錯体化学若手の会北海道・東北支部第 6 回勉強会, 東北大大学, 2014.11.15, 仙台市 (招待講演)
17. 佐藤宗太, 「巨大な中空球状錯体を骨格として構築した生体分子インターフェースの開発」"錯体化学会第 64 回討論会, 中央大学, 2014.9.20, 東京都 (招待・受賞講演)
18. Sota Sato, "Synthesis and Biological Applications of Self-Assembled Huge Molecules", Asian International Symposium (Inorganic Chemistry) – Emerging Science in Inorganic Porous Materials ,2014.3.29, Nagoya University (Nagoya City) (Invited talk)

19. 佐藤宗太, "Development of dynamical ordering of artificial molecules by mimicking biomolecular systems", Dynamical Ordering of Biomolecular Systems for Creation of Integrated Functions The 2nd International Symposium, Campus Plaza Kyoto (Kyoto Prefecture), 2014.1.11, Kyoto(Invited talk)
20. 佐藤宗太, "生体分子系を模倣した動的秩序をもつ人工分子の開発", 生命分子システムにおける動的秩序形成と高次機能発現 第1回公開シンポジウム, 岡崎コンファレンスセンター, 2013.10.2, 岡崎市 (招待講演)
【杉安和憲】
1. 杉安和憲, 時間発展する超分子集合体, 高分子学会 湘北地区懇話会, 神奈川大学, 2017.6.16, 神奈川 (招待講演)
2. Kazunori Sugiyasu, Introduction to Supramolecular Polymerization, Gordon Research Conference, Self-Assembly & Supramolecular Chemistry, 2017.5.22, Les Diablerets, Switzerland (Invited talk)
3. 杉安和憲, 時間発展する超分子集合体, 錯体化学若手の会 関東支部勉強会, 錯体化学若手の会 ,2016.11.26 (招待講演)
4. Kazunori Sugiyasu, Living supramolecular polymerization, ECNIS Symposium at Ewha Womans Univ., 2016.10.9, Korea (Invited talk)
5. Kazunori Sugiyasu, "Dynamic Ordering of Supramolecular Assemblies Developed on a Complex Energy Landscape", 4th International Symposium on Dynamical Ordering of Biomolecular Systems for Creation of Integrated Functions, 2015.11.22-23, Fukuoka
6. Kazunori Sugiyasu, "Living Supramolecular Polymerization", International Conference on Advanced Polymers via Macromolecular Engineering, 2015.10.18-22, Yokoyama.
7. Kazunori Sugiyasu, "Living Supramolecular Polymerization", International Symposium for Photo- and Electro-Molecular Machine, 2015.10.5-9, Toulouse
8. 杉安和憲, 自己増殖する超分子集合体 Living Supramolecular Polymerization, 日本化学会第 95 春季年会, 2015.3.26, 千葉
9. Kazunori Sugiyasu, Living Supramolecular Polymerization, China-Japan Joint Symposium on Functional Supramolecular Architectures, 2014.12.12, China (Invited talk)
10. K. Sugiyasu, "Conjugated Polymers Sheathed within Their Own Cyclic Sidechains ", The 10th SPSJ International Polymer Conference (IPC 2014), 2014.12.3, Tsukuba, Japan (Invited talk)
11. Kazunori Sugiyasu, Living Supramolecular Polymerization, The 5th International Symposium of Advanced Energy Science, 2014.10.1, Kyoto(Invited talk)
12. 杉安和憲. 自己組織化をコントロールした超分子ポリマー, 高分子学会精密ネットワークポリマー研究会, 東工大, 2014.7.10, 東京
13. 杉安和憲, 超分子化学的に精密設計された高分子 and vice versa, 高分子若手研究会・夏の講演会, 2014.6.27, 北九州 (招待講演)

【鈴木大介】

1. Daisuke Suzuki, Masaya Takizawa, Haruka Minato, Yuka Sazuka, "Functional Hydrogel Microspheres at the Air/Water Interface", 255th ACS National Meeting, 2018.3.21, New Orleans, America (Invited talk)
2. Daisuke Suzuki, Masaya Takizawa, Yuka Sazuka, Haruka Minato, "Self-Organization of Soft Hydrogel Microspheres at the Air/Water Interfaces", The 6th Asian Symposium on Emulsion Polymerization and Functional Polymeric Microspheres, 2018.3.9, Fukui, Japan (Invited talk)
3. 鈴木大介, 「高分子ゲル微粒子の表面・内部構造制御と機能創出」, 日本接着学会粘着研究会 1 月度例会, 大阪市立工業研究所, 2017.1.27, 大阪 (招待講演)
4. Daisuke Suzuki, "Artificial Polymeric Microhydrogels toward Autonomous Soft Materials" The 5th International Symposium on Dynamical Ordering of Biomolecular Systems for Creation of Integrated Functions, 2017.1.21, Tokyo, Japan(Invited talk)
5. 鈴木大介, 「刺激応答性ヒドログル微粒子の開発に必要な評価技術」, 第 28 回散乱研究会, HULIC HALL, 2016.11.25, 東京 (招待講演)
6. 鈴木大介, 「顕微鏡法と散乱法を駆使したゲル微粒子の微細構造評価」, -日本化学会秋季事業- 第 6 回 CSJ 化学フェスタ 2016, タワーホール船堀, 2016.11.14, 東京 (招待講演)
7. 鈴木大介, 「ソフトゲル微粒子の次元構造とミクロ空間場における機能制御」, 第 16 回ナノテク高機能テキスタイル基盤技術研究会, 信州大学纖維学部, 2016.10.21, 長野 (招待講演)
8. 鈴木大介, 高分子ヒドログル微粒子を用いた空間・時間構造制御」, 第 157 回東海高分子研究会講演会(岐阜), 長良川観光ホテル石金, 2016.9.2, 岐阜 (招待講演)

9. Daisuke Suzuki, "Control of Spatio-temporal Structures for Polymeric Hydrogel Microspheres" The 5th Asian Symposium on Emulsion Polymerization and Functional Polymeric Microspheres (ASEPFP), 2015.10.25, Suzhou, China (Invited talk)
10. 鈴木大介, 「環境応答型高分子ゲル微粒子の構造と機能設計」, 計測自動制御学会中部支部シンポジウム, 信州大学繊維学部, 2015.9.24, 長野(招待講演)
11. 鈴木大介, 「高分子ヒドロゲル微粒子の微細構造と機能」, 第 84 回高分子若手研究会[関西]一次世代機能性材料の創出を担うポリマーサイエンス, 六甲山 YMCA, 2015.7.25, 兵庫, (招待講演)
12. Daisuke Suzuki, "Control of Spatio-Temporal Structures for Polymer Microspheres toward Advanced Soft Materials", Supramolecular Dynamics at the Interface of Chemistry and Biology, Institute for Molecular Science and Okazaki Institute for Integrative Bioscience, 2015.6.12, Aichi, Japan, (Invited talk)
13. 鈴木大介, 「機能性高分子ゲル微粒子の創製と構造評価」, 東京工業大学セミナー, 東京工業大学大岡山キャンパス, 2015.3.10, 東京 (招待講演)

【平岡秀一】

1. 平岡秀一, 自己集合過程の解明と平衡に支配されない自己集合, 日本化学会 第 98 春季年会, 日本大学 2018.3.20, 船橋 (招待講演)
2. Shuichi Hiraoka, Coordination Self-Assembly Process by QASAP, 新学術第 6 回国際シンポジウム, 2018.1.21, 浜松 (Invited talk)
3. Shuichi Hiraoka, How does coordination self-assembly take place?, New Frontiers of Supramolecular Chemistry in Non-equilibrium Systems, 東京工業大学 大岡山, 2018.1.11, Tokyo (Invited talk)
4. Shuichi Hiraoka, Coordination Self-Assembly Process and a Novel Method for the Formation of Coordination Self-Assemblies under Kinetic Control, 錯体化学会 第 67 回討論会, 2017.9.16, 北海道 (Invited talk)
5. Shuichi Hiraoka, Nanocube: Ultra-thermostable Self-assemblies In Water, International Symposium on Pure & Applied Chemistry 2017, 2017.6.9, Vietnam (Invited talk)
6. Shuichi Hiraoka, Nanocube: Ultra-Thermostable Self-Assemblies in Water, Frontier Bioorganization Forum 2017: Dynamical ordering and integrated functions of biomolecular systems, 2017.4.25, Taipei, (Invited talk)
7. Shuichi Hiraoka, Nanocube: Ultra-Thermostable Self-Assemblies in Water, Frontier Bioorganization Forum 2017: Dynamical ordering and integrated functions of biomolecular systems, 2017.4.22, 台湾 (Invited talk)
8. 平岡秀一, 超耐熱性自己集合体の秩序形成, 日本化学会第 97 春季年会, 2017.3.16, 神奈川 (招待講演)
9. Shuichi Hiraoka, Self-Assembly Process of Coordination Rings, Cages, and Capsules, International Symposium on Polymer and Related Materials, 2016.7.11, China (Invited talk)
10. 平岡秀一, 超分子錯体の自己組織化機構, 日本化学会第 96 春季年会特別企画, 同志社大学, 2016.3.24, 京田辺市 (招待講演)
11. Shuichi Hiraoka, Self-Assembly Process of Coordination Capsules and Cages, The 9 th Japan-China Joint Symposium on Functional Supramolecular Architectures, 2016.2.24, Okazaki, (Invited talk)
12. Shuichi Hiraoka, Self-Assembly Process of Coordination Cages and Capsules, 3rd Workshop of Center for Instrumental Analysis, Frontier of Instrumental Analysis using Magnetic Resonance Spectroscopy, 2015.10.30, Niigata (Invited talk)
13. 平岡秀一, メカニズム指向の自己組織化に挑戦! ~自己集合性金属錯体の形成機構の解明~, 第 5 回 CSJ 化学フェスタ 2015, タワーホール船堀, 2015.10.13, 東京 (招待講演)
14. 平岡秀一, 特別企画生命および人工分子システムにおける動的秩序形成:分子論的理解「金属錯体型自己組織化のメカニズム」, 日本化学会第 95 春季年会, 2015.3, 千葉 (招待講演)
15. 平岡秀一, 「金属錯体型自己組織化体の形成過程を実験的に追跡する」, 第 35 回生物物理学セミナー, 2013.12, 奈良 (招待講演)

【二木史郎】

1. 二木史朗, ペプチドを用いた生理活性分子の細胞内送達, 日本ケミカルバイオロジー学会第 13 回年会・日本農芸化学会合同シンポジウム, 2018.6.11-13, 東京 (招待講演)
2. 二木史朗, ペプチドを用いた細胞内デリバリー, 第 393 回 CBI 学会講演会「中分子創薬の実現にむけて」, 2018.3.16, 東京 (招待講演)
3. Shiroh Futaki, Cytosolic Antibody Delivery by Lipid-sensitive Endosomolytic Peptide , IRCCS-JST CREST Joint Symposium "Chemical sciences facing difficult challenges", 2018.1.24-26, 福岡 (Invited

talk)

4. Shiroh Futaki, Cytosolic delivery of biomacromolecules, 2018 International Symposium on Chemical Biology, 2018. 1. 10-12, Geneva, Switzerland (Keynote lecture)
5. Shiroh Futaki, Cytosolic antibody delivery by lipid-sensitive endosomalytic peptide, The Second A3 Roundtable Meeting on Chemical Probe Research Hub, 2017.11.23-26, Hangzhou, China (Invited talk)
6. Shiroh Futaki, Peptide-mediated Delivery of Biomacromolecules into Cells, 12th Australian Peptide Conference 2017, 2017.10.15-20, Queensland, Australia (Invited talk)
7. 二木史朗, 細胞内デリバリーペプチドの膜との相互作用, 17-1 バイオ・高分子研究会「生命を操るバイオ高分子設計」, 2017.9.22, 松山 (招待講演)
8. 二木史朗, ペプチドの膜との相互作用・膜透過, 平成 28 年度特別経費事業 (創薬人育成のための創薬実践道場教育構築事業), (多機能性人工エキソソーム(iTEX)医薬品化実践を通じた操薬人育成事業), 合同シンポジウム, 2017.2.13, 徳島 (招待講演)
9. 二木史朗, ペプチドの膜との相互作用・膜透過, 遺伝子・デリバリー研究会 第 16 回夏期セミナー, 2016.9.13, 長崎 (招待講演)
10. Shiroh Futaki, Yoshimasa Kawaguchi, Toshihide Takeuchi, Keiko Kuwata, Photocrosslinking approaches to identify cellular-uptake receptors of arginine-rich cell-penetrating peptides, 34th European Peptide Symposium and 8th International Peptide Symposium, 2016.9.4-9, Leipzig, (Keynote Lecture)
11. Shiroh Futaki, 14th Chinese International Peptide Symposium & 5th Asia-Pacific International Peptide Symposium (CPS-14 & APIPS-5) APIPS, Facilitated membrane translocation of arginine-rich peptides in the presence of curvature inducing peptides, 2016.7.7, 南京 (Invited talk)
12. 二木史朗, ペプチドを用いた細胞内デリバリー, 創薬懇話会 2016 in 蓼科: 次世代を担う若手のためのメデイシナルケミストリーフォーラム, 蓼科, 2016.7.1 (招待講演)
13. Shiroh Futaki, Tomo Murayama, Membrane-remodeling Amphiphilic Helical Peptides Accelerate Direct Penetration of Octaarginine, 20th Korea Peptide Protein Society (KPPS) Symposium, 2016.6.24-25, Yangyang, Korea (Invited talk)
14. 二木史朗, 見る・知る・作る: ペプチドと細胞膜との相互作用, 第 9 回東京大学化学生命工学専攻 ChemBio ハイブリッドレクチャー, 2016.6.10-8. 東京 (招待講演)
15. Shiroh Futaki, Peptide Mediated Delivery of Bioactive Proteins into Cells, 8th AsiaTIDES, 2016.2.24, 京都(Invited talk)
16. Shiroh Futaki, Membrane translocation of arginine-rich peptides and the effect of membrane curvature, The International Chemical Congress of Pacific Basin Societies 2015 (Pacificchem 2015), 2015.12.19, Honolulu, Hawaii, USA(Invited talk)
17. Shiroh Futaki, Exploiting hemolytic amphiphilic peptides as a design template for cytosolic antibody delivery, The International Chemical Congress of Pacific Basin Societies 2015 (Pacificchem 2015), 2015.12.17, Honolulu, Hawaii, USA (Invited talk)
18. Shiroh Futaki, Peptide mediated delivery of bioactive proteins into cells
The 7th International Peptide Symposium, 2015.12.9, Singapore (Invited talk)
19. Shiroh Futaki, A novel lysosomolytic peptide designed from spider venom 7th Peptide Engineering Meeting (PEM7-2015), 2015.12.6, Pune, India (Invited talk)
20. Shiroh Futaki, Converting a cytotoxic, hemolytic peptide to an intracellular delivery tool of bioactive proteins, NEA & ASIAHORCs Joint Symposium on Chemical Biology, 2015.11.10, 沖縄 (Invited talk)
21. 二木史朗, 「アミノ酸・ペプチドは体の調子を整える」, ひょうご講座 2015「健康をサイエンスする」, 2015.10.6, 神戸 (特別講師)
22. 二木史朗, 「細胞の営みを探る」, ひょうご講座 2015「健康をサイエンスする」, 2015.9.15, 神戸 (特別講師)
23. Shiroh Futaki, Peptide-based approaches for delivering exogenous molecules into cells
International symposium: "From cell-penetrating peptides to nanoparticles for cellular delivery"
2015.7.5, Paris (Invited talk)
24. Shiroh Futaki, Peptide-based approaches for delivering exogenous molecules into cells Asian Chemical Biology Conference 3 (ACBC3 2014), University Town (U-Town), 2014.12.16, Singapore, (Invited talk)
25. Shiroh Futaki, The Sino-Japan Workshop on Chemical Biology, Positive membrane curvature induced by epsin N-terminal peptide accelerates internalization of octaarginine, 2014.10.11, Beijing, China, (Invited talk)
26. Shiroh Futaki, Extramembrane control of peptide/protein assembly, Workshop on Artificial

- Photosynthesis: Engineering of Light-Harvesting Processes based on Peptide and Protein Science, 2014.10.6, Nagoya (Invited talk)
27. Shiroh Futaki, Positive membrane curvature induced by epsin N-terminal peptide accelerates internalization of octaarginine Swiss-Japanese Chemical Biology Symposium 2014, 2014.10.2, Bern, Switzerland (Invited talk)
 28. Shiroh Futaki, Engineering membrane curvatures by peptides, 15th Akabori Conference-Japanese-German Symposium on Peptide Science, 2014.9.8, Boppard, Germany(Invited talk)
- 【芳坂貴弘】
1. 芳坂貴弘, 非天然アミノ酸の導入によるタンパク質機能の人工的カスタマイズ, 第 16 回日本蛋白質科学会,2016.6.9, 福岡 (招待講演)
 2. 芳坂貴弘, 非天然アミノ酸導入技術のバイオ医薬への応用展開, 第 22 回ペプチドフォーラム, 2016.3.5, 金沢 (招待講演)
 3. Takahiro Hohsaka, Incorporation of nonnatural amino acids through expansion of the genetic code and its application to fluorescence analysis of proteins, BMB2015 (第 38 回分子生物学会年会・第 88 回生化学会大会合同大会) , 2015.12.2, 神戸(Invited talk)
 4. Takahiro Hohsaka, Incorporation of nonnatural amino acids by expanding the genetic code and its application to fluorescence analysis of proteins, The 14th KIAS Conference on Protein Structure and Function, 2014.9.18, Seoul, Korea
 5. 芳坂貴弘, 非天然アミノ酸の導入技術を利用したタンパク質の人工機能拡張 蛋白質科学会年会, 2014.6.26, 横浜 (招待講演)
 6. 芳坂貴弘, タンパク質への非天然アミノ酸導入技術の開発とその応用, 酵素工学研究会第 70 回講演会,2013.10.25, 東京 (招待講演)
- 【松浦友亮】
1. 松浦友亮, 「再構成型セルフリータンパク質合成系を用いたタンパク質配列空間と成分濃度空間探索」 2018 年度日本農芸化学会, 名城大学, 2018.3.15-18, 名古屋市
 2. 松浦友亮, 「リポソームディスプレイ法を用いた膜タンパク質の創生と解析」 2017 年度生命科学系学会合同年次大会 (ConBio2017) , 神戸ポートアイランド, 2017.12.6-9, 神戸市
 3. Tomoaki Matsuura, "Decorating liposome surface with biological molecule and its application" IGER International Symposium on Cell Surface Structures and Functions 2017, Nagoya University, 2017.11.30-12.1, Nagoya, Japan
 4. Tomoaki Matsuura, "In vitro membrane protein engineering by liposome display", The 17th KIAS Conference on Protein Structure and Function, Korea Institute for Advanced Study, 2017.9.21-22, Seoul, Korea
 5. Tomoaki Matsuura, "Evolutionary engineering and characterization of membrane proteins using liposome display" In 2017 KSBB Spring Meeting and International Symposium, Hyundai Hotel, 2017.4.6-7, Gyeongju, Korea
 6. Tomoaki Matsuura, "Evolutionary engineering and characterization of membrane proteins using liposome display" 2nd Korea-Japan Smart Biodesign Workshop: Technology exchange for green biotechnology, Sendai City Information & Industry Plaza, 2017.2.4, Sendai, Japan
 7. 松浦友亮, "Evolutionary engineering and characterization of membrane proteins using liposome display" 第 39 回日本分子生物学会年会 (パシフィコ横浜) , 2016.11.30-12.2, 神奈川
 8. 松浦友亮, 「リポソーム内で膜タンパク質を合成するシステムの構築と応用」 第 29 回植物脂質シンポジウム, 大阪大学会館, 2016.11.25-26, 豊中市
 9. 松浦友亮, 「セルフリータンパク質合成系を用いた進化分子工学技術の開発」, 富山国際会議場など, 2016.9.28-30, 富山(第 52 回生物工学奨励賞(斎藤賞) 受賞講演)
 10. 松浦友亮, 「リポソームディスプレイ法を用いた進化分子工学技術の開発と応用」 第 89 回日本生化学会大会 (仙台国際センター・東北大学川内北キャンパス 2016.9.25-27, 宮城
 11. 松浦友亮, 「In vitro 進化分子工学を用いたタンパク質デザイン」 第 56 回生物物理若手の会 夏の学校, 2016.9.2-5
 12. Tomoaki Matsuura, "The time development of the protein synthesis involves a collapse and regrowth of the reaction network" QBic Symposium 2015, High-dimensional data for the design principles of life (RIKEN Quantitative Biology Center, 2016.8.24-26, Osaka
 13. 松浦友亮, 「生体高分子を用いた構成的アプローチ」 研究会「物理学的普遍性から生物学的普遍性へ」(東京大学駒場キャンパス 21KOMCEE east 011 教室) 2016.8.4-5, 東京

14. 松浦友亮, 「*in vitro* で組み上げた生命システムから学べること」細胞システムの動態と論理 VIII, 理化学研究所, 2016.4.14-15, 和光
15. 松浦友亮, 「無細胞タンパク質合成系を用いた膜タンパク質進化分子工学」第 4 回 ネオバイオ分子研究会, 大阪府立大学 I-site, 2016.1.29, なんば・大阪市
16. 松浦友亮, 「全成分タンパク質合成反応モデルの構築とこれを用いた反応ダイナミクス解析」第 38 回日本分子生物学会年会・第 88 回日本生化学会大会合同大会, 神戸ポートアイランド, 2015.12.1-4, 神戸市
17. Tomoaki Matsuura, "Dynamics of biological systems constructed *in vitro*" The 4th International Symposium on Dynamical Ordering of Biomolecular Systems for Creation of Integrated Functions, Kyushu University, 2015.11.22–23, Fukuoka
18. 松浦友亮, 「バイオ界面を利用した *in vitro* 進化分子工学」日本生物工学会第 67 回大会, 城山観光ホテル, 2015.10.26-28, 鹿児島市
19. Tomoaki Matsuura, "The time development of the protein synthesis involves a collapse and regrowth of the reaction network" QBic Symposium 2015, High-dimensional data for the design principles of life, RIKEN Quantitative Biology Center, 2015.8.24-26, Osaka
20. 松浦友亮, 「無細胞タンパク質合成系を用いた膜タンパク質進化分子工学」第 15 回日本蛋白質科学会年会, あわぎんホール, 2015.6.24-26, 徳島市
21. Tomoaki Matsuura, "In Vitro Selection and Evolution of Membrane Proteins" Gordon Research Conferences in Proteins, 2015.6.15-20, Boston, U.S.A.
22. 松浦友亮, *in vitro* で生体分子を使って組み上げる生命システム. In 国際高等研研究プロジェクト「分子基盤に基づく生体機能ネットワークとダイナミクスの解明」第 1 回研究会プログラム 2014.12.14-15
23. Tomoaki Matsuura, In vitro evolution of alpha-hemolysin using a liposome display. Young Investigators Talk session in Protein Society Meeting 2014, 2014.7.27-30, SanDiego, U. S. A.
24. Tomoaki Matsuura, Engineering Membrane Proteins Entirely *in vitro* with Liposome Display Technology. In 2014 KSBB SPRING MEETING and INTERNATIONAL SYMPOSIUM, Hyundai Hotel, 2014.4.10, Gyeongju, Korea

【三宅弘之】

1. 三宅弘之, Stimuli-Responsive Helical Metal Complexes: Helicity Inversion, Expansion/Contraction, and Memorization, International Congress on Pure & Applied Chemistry (ICPAC) 2018, 2018.3.7-10, Siem Reap, (Cambodia)(Invited talk)
2. 三宅弘之, Coordination Chemistry-Based Dynamic Structure Switching in Helical Metal Complexes The 1st International Symposium on Coordination Ionic Compounds, 2017.12.15-16, 大阪 (Invited talk)
3. 三宅弘之, Coordination Chemistry Strategies for Dynamic Structural Switching, Invited Seminar of The institute of Chemistry and Biology of Membranes and Nano-objects (CBMN, UMR5248), CNRS, University of Bordeaux and Bordeaux Aquitaine National Polytechnic Institute, 2017.6.16, Bordeaux, France (Invited talk)
4. Hiroyuki Miyake, Dynamic Structural Switching in Asymmetric Coordination Chemistry", Invited Seminar at Department of Chemistry, Gyeongsang National University, 2016.12.1, Jinju, Korea (Invited talk)
5. 三宅弘之, Dynamic Supramolecular Architecture via Asymmetric Coordination Chemistry, 錯体化学会第 66 回討論会シンポジウム, 「Inorganic Molecular/Supramolecular Systems for Future Development of Photofunctional Materials」, 2016.9.10, 福岡 (Invited talk)
6. Hiroyuki Miyake, Dynamic Chiral Metal Complexes in Response to Combination of Multiple External Stimuli, Symmetry Festival, 2016.7.18-22, Vienna, Austria (Invited talk)
7. 三宅弘之, アミノ酸誘導体配位子からなる錯体ヘリシティーの創生と動的構造変換プログラミング, 分子研究会「金属錯体の非対称配位圈設計と異方集積化が拓く新物質創成科学」, 岡崎, 2016.3.5-6 (招待講演)
8. Hiroyuki Miyake, Dynamic Helicates based on Labile Metal Complexes, International Conference on Polymers and Advanced Materials POLYMAT-2015, Mexico, 2015.10.18-22 (Invited talk)
9. Hiroyuki Miyake, Chirality regulation in labile metal complexes via dynamic structural conversion, The Molecular Chirality Asia 2014, 2014.10.29-31, China (Invited talk)

A03 : 動的秩序の展開

【稻垣直之】

1. 稻垣直之, 分子の自己組織化による細胞の形態形成, 日本化学会第 98 春季年会, 2018.3.20, 千葉 (招待講演)
2. 稻垣直之, Molecular Bases for Gradient Reading and Force Generation of the Axon Guidance, 新領域「動的秩序と機能」第 6 回国際シンポジウム, 2018.1.20, 浜松 (招待講演)

3. 稲垣直之, 細胞移動装置の前駆体としてのアクチン波について, 第2回秩序化分子システムワークショッピング, 2017.12.26, 奈良 (招待講演)
4. 稲垣直之, Molecular Mechanics of Neuronal Axon Guidance, 2017年度生命科学系学会合同年次大会, 2017.12.9, 神戸 (招待講演)
5. 稲垣直之, 分子の集合・離脱が駆動する神経軸索ガイダンスの分子メカニクス, 第55回日本生物物理学会年会, 2017.9.19, 熊本 (招待講演)
6. 稲垣直之, 勝野弘子, 鳥山道則, 細川陽一郎, 水野健作, 池田和司, 作村諭一, 新たな細胞内分子輸送機構 Actin Wave と細胞形態形成, 第69回細胞生物学会大会, 2017.6.13, 仙台 (招待講演)
7. Naoyuki Inagaki, Molecular Mechanism for Axon Navigation in the Brain, Frontier Bioorganization Forum 2017 Dynamical ordering and integrated functions of biomolecular systems, 2017.4.24, 台湾 (Invited talk)
8. Naoyuki Inagaki, Molecular Mechanism for Axon Outgrowth and Neuronal Network Formation, Colorado State University Seminar, 2016.12.8, Denver, USA (Invited talk)
9. 稲垣直之, 蛍光ライブイメージングと牽引力顕微鏡による神経軸索ガイダンス機構の解析, 第25回日本バイオイメージング学会学術集会, 2016.9.5, 名古屋 (招待講演)
10. 勝野弘子, 鳥山道則, 細川陽一郎, 水野健作, 池田和司, 作村諭一, 稲垣直之, アクチンの重合・脱重合によって引き起こされる軸索内タンパク質輸送, シンポジウム「外的環境を感知する軸索内環境調節の分子基盤」, 第121回日本解剖学会総会・全国学術集会, 2016.3.28, 福島県郡山市 (招待講演)
11. Naoyuki Inagaki, Molecular Mechanism for Axon Outgrowth and Neuronal Network Formation, 7th Annual National Convention of Philippine Society for Developmental Biology, 2015.10.24, Philippines (Invited talk)
12. Naoyuki Inagaki, Molecular Mechanism for Axon Outgrowth and Neuronal Network Formation, The 4th International Conference on Pharmacy and Advanced Pharmaceutical Sciences, 2015.9.8, Indonesia (Invited talk)
13. 稲垣直之, ラージゲルプロテオミクスを基盤とした神経細胞の軸索形成とガイダンスの解析, 第65回日本電気泳動学会, 2014.10.24-25, 横浜市 (招待講演)
14. 稲垣直之, 馬場健太郎, 久保祐亮, 生体分子素子の自己組織化による神経軸索の伸長と調節 第14回日本蛋白質科学会, 2014.6.27, 横浜市 (招待講演)
15. 稲垣直之, Signal-force Transduction in Axon Outgrowth and Guidance 第66回日本細胞生物学会大会, 2014.6.11-13, 奈良市 (招待講演)
16. Y.Sakumura, N.Inagaki, Spontaneous symmetry breaking in neural morphology Society for Industrial and Applied Mathematics Annual Meeting, 2014.6.7-11, Chicago, USA (Invited talk)
17. 稲垣直之, 軸索伸長のためのシグナル・力変換機構, 第51回日本生物物理学会年会, 2013.10.29, 京都市 (招待講演)

【内山進】

1. 石井健太郎, 杉山正明, 加藤晃一, 内山進, 超分子質量分析による蛋白質複合体の形成過程の解明, 第17回日本蛋白質科学会年会, 2017.6.22, 仙台 (招待講演)
2. Susumu Uchiyama, Biophysical analysis for therapeutic proteins, Malvern Bioscience Conference 2016, 2016.7.13-15, Seoul and Tokyo (Invited talk)
3. 内山進, バイオ医薬品の溶液物性のモニタリングと最適化, 化学工学会関東支部, 最新のバイオ原薬製造プロセスの現状と今後の展望, 2016.6.30, 宇都宮 (招待講演)
4. 内山進, 蛋白質医薬の物理化学的特性解析における蛋白質科学の役割, 第16回蛋白質科学会年会, 2016.6.8, 福岡 (招待講演)
5. 内山進, バイオ医薬品の物理化学的特徴とその計測方法, 安定性試験研究会特別セミナー, 2016.5.27, 東京 (招待講演)
6. 内山進, 超高分解能 ESI-QTOF MS および MALDI-TOF/TOF MS による抗体医薬の分析 質量分析総合討論会, 2016.5.19, 大阪 (招待講演)
7. Susumu Uchiyama, "Advantages of Polymer-based Pre-fillable Syringes for Therapeutic Proteins", 2015 PDA Europe Universe of Pre-filled Syringes and Injection Devices, 2015.11.3, Vienna, Austria.
8. 内山進, "Solution behavior of TLR9 studied by analytical ultracentrifugation", 第52回日本生物物理学年会 金沢大学, 2015.9.14, 金沢
9. 内山進 「医薬品の熱測定」, 2015年8月25~26日, 第75回熱測定講習会, 京都府立大学, 2015.8.25-26, 京都
10. 内山進 「水素重水素交換質量分析法による蛋白質の高次構造解析」, 第34回日本糖質学会年会, 東京大学, 2015.7.31, 東京

11. 内山進, 「抗体医薬などのバイオ医薬品の物理化学的評価」, 日本薬学会第 135 会年会, 2015.3.27, 神戸 (招待講演)
12. Susumu Uchiyama, "Biophysical studies on antibody aggregation" JAACT international symposium, 2014.11.12, Kitakyu-shu (Keynote lecture)
13. 内山進, 「抗体の物理化学的特性と抗体の凝集・機能性との関係」, 第 87 回 日本生化学会大会, 2014.10.18, 京都 (招待講演)
14. 内山進, In solution characterizations of protein-protein interactions by analytical ultracentrifugation and mass spectrometry 第 52 回日本生物物理学会年会, 2014.9.26, 札幌 (招待講演)
15. Susumu Uchiyama, "Biophysical factors governing antibody aggregation", Workshop on Protein Aggregation and Immunogenicity., 2014.7.24, Breckenridge, Colorado (招待講演)
16. 内山進, 蛋白質相互作用・超分子複合体の質量分析 第 14 回蛋白質科学会年会 ランチョンセミナー, 2014.6.26, 横浜 (招待講演)
17. 内山進, Analytical ultracentrifugation for quantitative intermolecular interaction studies, 第 14 回蛋白質科学会年会 ワークショップ, 2014.6.26, 横浜 (ワークショップ主催)

【老木成稔】

1. Shigetoshi Oiki, Ion Permeation and Gating of the KcsA Potassium Channel Reconstituted in the Manipulated Membrane, Ion Channel Modulation Symposium 2017, 2017.6.21, Cambridge, UK (Keynote lecture)
2. Sumikama, T. and Oiki, S.: A 3-D movie of ion permeation through ion channel at the atomic levelfilmed by computer simulation. 94th Annual Meeting of the Physiological Society of Japan, 2017.3.28-30, Hamamatsu
3. 老木成稔：“人工膜マニピュレーション”第 38 回生体膜と薬物の相互作用シンポジウム ミニシンポジウム 1 「人工膜を用いた研究最前線」名古屋市立大学 田辺通りキャンパス 2016.11.17-18, Nagoya
4. 老木成稔：カスタムメード膜でのチャネル研究法 カスタムメード膜でのチャネル研究法 第 63 回中部生理学会 特別プログラム「研究実験方法論」 岡崎コンファレンスセンター 2016.11.4-5, Okazaki
5. 老木成稔 チャネル研究のための人工膜法 自然科学研究機構プロジェクト・生理学研究所概算要求プロジェクト終了シンポジウム「次ステージ機能生命科学の展望」,2016, 岡崎
6. 老木成稔, チャネル研究のための人工膜法. 3 プロジェクト合同 終了シンポジウム「次ステージ機能生命科学の展望」, 岡崎コンファレンスセンター,2016.3.10, 岡崎
7. Shigetoshi Oiki, "Channel Function Reconstitution and Reanimation", The 45th NIPS International symposium, Co-sponsored by "The Journal of Physiology" "Cutting-edge approaches towards the functioning mechanisms", 2014.11.26-28, Okazaki (Invited talk)
8. 老木成稔, 「チャネル膜蛋白質 KcsA の一分子構造・機能ダイナミクスと膜」 膜学会「膜解析の最前線～生体膜・膜タンパク質から模擬膜,ソフトマターまで～」,2014.5.13, 東京 (招待講演)
9. Shigetoshi Oiki, "Spatio-Temporal Interplay of the Single KcsA Potassium Channel in the Membrane" OIST International Workshop: Single Protein Dynamics in Cellulo: Spatio-Temporal, Structural and Quantitative Analyses, 2014.4.21-25, Okinawa. (招待講演)

【岡本裕幸】

1. Yuko Okamoto, Generalized-ensemble simulations of biological molecular assembly and artificial molecular assembly, The 6th International Symposium on Dynamical Ordering of Biological Systems for Creation of Integrated Functions, 2018.1.20-21, Hamamatsu, Japan (invited talk)
2. Yuko Okamoto, Classical and quantum molecular simulations in generalized ensemble, International Workshop on Molecular Simulations, 2017.11.17-20, Nanjing, China(invited talk)
3. Yuko Okamoto, Generalized-ensemble algorithms for materials and biomolecular simulations Molecular Simulation Studies in Material and Biological Sciences (MSSMBS2017), 2017.9.7-10, St. Petersburg, Russia (Keynote lecture)
4. Yuko Okamoto, Generalized-ensemble algorithms for advanced materials simulations IUMRS-ICAM 2017, The 15th International Conference on Advanced Materials, 2017.8-27-9.1, Kyoto, Japan (invited talk)
5. Yuko Okamoto, Energy landscape of biomolecular systems studied by generalized-ensemble simulations, Energy Landscapes 2017, 2017.8.13-17, Goa, India, August (invited talk)
6. Yuko Okamoto, Efficient sampling methods for classical and quantum simulations, TSRC Workshop on the Chemistry and Dynamics in Complex Environments (CHEM-DiCE), 2017.6.26-30, Telluride, Colorado, U.S.A.(invited talk)

7. Yuko Okamoto, Biomolecular simulations in generalized ensemble, Frontier Bioorganization Forum 2017: Dynamical Ordering and Integrated Functions of Biomolecular Systems, 2017.4.24-26, Taipei, Taiwan(invited talk)
8. Yuko Okamoto, Generalized-ensemble simulations of complex systems, 2016 NCTS March Workshop on Critical Phenomena and Complex Systems, 2016.3.28-29, Hsinchu, Taiwan (invited talk)
9. Yuko Okamoto, Protein dynamics studied by generalized-ensemble simulations, The 251st American Chemical Society National Meeting, Symposium under the Computers in Chemistry (COMP) Division “30 Years of Protein Dynamics in Silico”, 2016.3.13-17, San Diego, California, U.S.A. (invited talk)
10. Yuko Okamoto, Enhanced sampling methods for exascale computational chemistry, The 2015 International Chemical Congress of Pacific Basin Societies (PACIFICHEM2015), Symposium under the Physical, Theoretical & Computational Session, “Challenges and Opportunities for Exascale Computational Chemistry”, 2015.12.15-20, Honolulu, Hawaii, U.S.A. (invited talk)
11. Yuko Okamoto, Generalized-ensemble algorithms for enhanced configurational sampling, Algorithms in Structural Bioinformatics: Sampling in Biomacromolecular Systems (AlgoSB Winter School - 2015), 2015.11.29-12.4, Cargèse, Corsica, France (invited talk)
12. 岡本祐幸, 「拡張アンサンブル法による量子化学シミュレーション」,分子研研究会「理論計算分子科学ワークショップ」, 2015.10.22, 岡崎市,日本 (招待講演)
13. Yuko Okamoto, Generalized-ensemble simulations of classical and quantum molecular systems, The 6th Japan-Czech-Slovak International Symposium for Theoretical Chemistry, 2015.10.11-15, Smolenice, Slovakia (invited talk)
14. Yuko Okamoto, Generalized-ensemble simulations, Hands-on Workshop on Computational Biophysics at Okazaki, 2015.9.9-11, Okazaki, Japan (invited talk)
15. 岡本祐幸, 「古典系および量子系の拡張アンサンブルシミュレーション」,スーパーコンピュータワークショップ, 2015.9.7-8, 岡崎市,日本(招待講演)
16. Yuko Okamoto, Generalized-ensemble simulations of complex systems, 2015 NCTS International Workshop on Critical Phenomena and Complex Systems, 2015.8.15-17, Hsinchu, Taiwan (invited talk)
17. Yuko Okamoto, Generalized-ensemble algorithms for calculations of ligand binding affinity, BIRS Workshop: Free-Energy Calculations. A Mathematical Perspective, 2015.7.19-24, Oaxaca, Mexico, (invited talk)
18. Yuko Okamoto, Generalized-ensemble algorithms for enhanced sampling and free energy calculations Summer Snowmass Biophysics Workshop, Free Energy Calculations: Three Decades of Adventure in Chemistry and Biophysics, 2015.7.5-9, Snowmass, Colorado, U.S.A. (invited talk)
19. Yuko Okamoto, Efficient sampling methods for identifying transition states TSRC Workshop on the Chemistry and Dynamics in Complex Environments (CHEM-DiCE), Telluride, 2015.6.23-26, Colorado, U.S.A. (invited talk)
20. Yuko Okamoto, Efficient sampling methods for complex systems, The 19th International Annual Symposium on Computational Science and Engineering (ANSCSE19), 2015.6.17-19, Ubon Ratchathani, Thailand (Keynote lecture)
21. 岡本祐幸, 「拡張アンサンブル法による生体分子シミュレーション」, バイオスーパーコンピューティング名古屋 2015,2015.1.22, 名古屋 (招待講演)
22. Yuko Okamoto, Lecture 1: “An introduction to replica-exchange molecular dynamics simulation” Lecture 2: “Generalized-ensemble algorithms”, The 11th Thai Summer School of Computational Chemistry 2015 Workshop:Replica Exchange Molecular Dynamics Simulation, 2015.1.4-7, Nan, Thailand (invited talk)
23. Yuko Okamoto, “Protein folding and ligand binding simulations by generalized-ensemble algorithms” The 7th Korea-Japan Seminars on Biomolecular Sciences – Experiments and Simulations, 2014.11.26-28, Seoul, Korea (invited talk)
24. 岡本祐幸, 「分子シミュレーションにおける拡張アンサンブル法」, 第8回分子シミュレーションスクール－基礎から応用まで－, 2014.10.14-17, 岡崎 (招待講演)
25. Yuko Okamoto, “Protein folding and unfolding simulations by generalized-ensemble algorithms” Dushanbe Symposium on Computational Materials and Biological Sciences (DSCMBS-2014), 2014.9.23-28, Dushanbe, Tajikistan (Keynote lecture)

26. Yuko Okamoto, "Drug design by generalized-ensemble simulations", The 6th Japan-Russia International Workshop on Molecular Simulation Studies in Material and Biological Sciences (MSSMBS-2014), 2014.9.21-22, Moscow, Russia (Keynote lecture)
27. Yuko Okamoto "Computer simulations of protein folding, ligand binding, and proton transfer" 2nd International Conference on Computational Science and Engineering (2nd ICCSE 2014), 2014.8.21-23, Ho Chi Minh City, Vietnam (invited talk)
28. Yuko Okamoto, "Enhanced configurational sampling methods for spin systems and biomolecular systems" XXVI IUPAP Conference on Computational Physics, CCP2014, 2014.8.11-14, Boston, Massachusetts, U.S.A. (Keynote lecture)
29. Yuko Okamoto, "Enhanced sampling techniques for spin and biological systems", The 10th AIMS Conference on Dynamical Systems, Differential Equations and Applications Special Session "Enhanced Sampling Techniques in Simulation of Complex Systems", 2014.7.7-11, Madrid, Spain (invited talk)
30. 岡本祐幸, 「生体分子集団の相互作用と自由エネルギー計算」, 第 14 回日本蛋白質科学会年会, 2014.6.25-27, 横浜 (招待講演)
31. 岡本祐幸, 「拡張アンサンブル法による分子シミュレーション」, 第 3 回産学連携シンポジウム「HPC の利用と成果と人材」, 2014.1.31, 名古屋 (招待講演)
32. Yuko Okamoto, "Reweighting techniques for Monte Carlo and molecular dynamics simulations" The 13th KIAS Protein Folding Winter School, High 1 Resort, 2014.1.19-24, Korea (Invited talk)
33. Yuko Okamoto, "Generalized-ensemble simulations of biological molecular assembly and artificial molecular assembly" The 2nd International Symposium on Dynamical Ordering of Biological Systems for Creation of Integrated Functions, 2014.1.11-12, Kyoto, Japan (Invited talk)
34. Yuko Okamoto, "Computer modeling of complex systems by generalized-ensemble simulations" The 2013 Hong Kong Workshop in Computer Modeling of Complex Processes, 2013.12.17-20, Hong Kong (Invited talk)
35. 岡本祐幸, 「拡張アンサンブル法による生体分子の高次構造と機能の解明」, 物性研スパコン共同利用・CMSI 合同研究会 (第 4 回 CMSI 研究会) ,2013.12.11-13, 柏 (招待講演)
36. Yuko Okamoto, Generalized-ensemble algorithms for molecular simulations, The 3rd International Conference on Molecular Simulation (ICMS 2013), 2013.11.18-20, Kobe, Japan (Invited talk)
37. Yuko Okamoto, "Enhanced-sampling simulations of spin and biological systems", West-Lake International Workshop on Statistical Physics and Complex Systems, 2013.11.8-10, Hangzhou, China (Keynote lecture)
38. Yuko Okamoto, "Generalized-ensemble algorithms for first-principles electronic structure calculations", 16th Asian Workshop on First-Principles Electronic Structure Calculations (ASIAN-16), 2013.10.27-30, Beijing, China (Invited talk)
39. 岡本祐幸, 「生体分子集団および人工分子集団の相互作用と大規模構造転換」 新学術領域研究「動的秩序と機能」第 1 回公開シンポジウム,岡崎,2013.10.2 (招待講演)
40. Yuko Okamoto, "Generalized-ensemble simulations of spin and biological systems", The 1st International Symposium on Computational Materials and Biological Sciences, 2013.9.10-12, Tokyo, Japan, (Invited talk)
41. Yuko Okamoto, "Simulational physics in generalized ensemble",VII Brazilian Meeting on Simulational Physics, 2013.8.6-10, Joao Pessoa, Brazil (Invited talk)
42. Yuko Okamoto, "Generalized-ensemble algorithms for free energy calculations", Free Energy Calculations: Three Decades of Adventure in Chemistry and Biophysics, 2013.7.15-19, Snowmass, Colorado, U.S.A. (Invited talk)

【奥村久士】

1. 奥村久士, アミロイド線維の生体分子動力学シミュレーション, 企業研究会第 31 期 CAMM フォーラム 本例会, 2018.1.12, 東京 (招待講演)
2. Hisashi Okumura, Molecular dynamics simulations for aggregation and disaggregation of amyloid- β peptides, EMN Meeting on Computation and Theory 2017, 2017.11.6-10, Dubai, United Arab Emirates (Invited talk)
3. 奥村久士, 分子動力学シミュレーションによるアミロイド線維の形成過程と破壊過程, 近畿化学協会コンピュータ化学部会例会, 2017.10.27, 大阪 (招待講演)
4. Hisashi Okumura, Simulational studies of A β amyloid fibrils by equilibrium and nonequilibrium molecular dynamics method, 21st International Annual Symposium on Computational Science and

- Engineering, 2017.8.3-4, Pathum Thani, Thailand (Invited talk)
5. Hisashi Okumura, "All-atom molecular dynamics simulations of A_β amyloid fibrils", Institute for Protein Research (IPR) Seminar, Osaka University, Suita, 2017.1.26-27, Osaka, Japan (Invited talk)
 6. Hisashi Okumura, "Equilibrium and nonequilibrium molecular dynamics simulations of A_β amyloid fibrils", 10th International Conference on Computational Physics, Holiday Inn, Sands Cotai Central, 2017.1.16-20, Macau, China (Invited talk)
 7. Hisashi Okumura, "Dynamical ordering of amyloid fibril studied by molecular dynamics simulations", Thai-Japan Symposium in Chemistry, Chiang Mai University, 2016.11.14-16, Chiang Mai, Thailand (Invited talk)
 8. Hisashi Okumura, "Molecular dynamics simulations to study dynamical ordering of amyloid fibril", 2016 NCTS October Workshop on Critical Phenomena and Complex Systems, National Tsing Hua University, 2016.10.5, Hsinchu, Taiwan (Invited talk)
 9. Hisashi Okumura, "Molecular dynamics simulations of amyloid fibrils", 2016 NCTS March Workshop on Critical Phenomena and Complex Systems, National Tsing Hua University, 2016.3.28-29, Hsinchu, Taiwan, (Invited talk)
 10. 奥村久士, 「分子動力学シミュレーションによるアミロイド線維の離合集散」, 日本化学会特別企画「どこまで明らかになったか?自己組織化のメカニズム:アミロイド形成から人工系」, 同志社大学京田辺キャンパス, 2016.3.24-27, 京都 (招待講演)
 11. Hisashi Okumura, "Pressure induced structural change of proteins by molecular dynamics simulations", Eighth Japan-Korea Seminars on Biomolecular Sciences, Institute for Molecular Science, 2016.2.15-17, Okazaki, Japan (Invited talk)
 12. 奥村久士, 「分子動力学シミュレーションで探るアミロイドβペプチドの凝集,離散」国立長寿医療研究センター, CAMD セミナー, 2016.2.18, 愛知
 13. Hisashi Okumura, "Molecular dynamics simulations of proteins under high pressure", Pure and Applied Chemistry International Conference 2016, Bangkok International Trade & Exhibition Centre, 2016.1.9-11, Bangkok, Thailand (Invited talk)
 14. Hisashi Okumura, "Molecular dynamics simulations for oligomerization and disruption of amyloid-β fibril", The International Chemical Congress of Pacific Basin Societies 2015 (Pacificchem 2015), Hawaii Convention Center, 2015.12.15-20, Honolulu, Hawaii, USA (Invited talk)
 15. Hisashi Okumura, "Nonequilibrium molecular dynamics simulation of amyloid-fibril disassembly by supersonic cavitation", International workshop on complex phenomena from molecule to society, University of Tokyo, 2015.11.24-25, Tokyo, Japan (Invited talk)
 16. 奥村久士, 「各種統計アンサンブルの生成法」, 第9回分子シミュレーションスクール—基礎から応用まで, 分子科学研究所, 2015.10.13-16, 愛知
 17. 奥村久士, 「アミロイド線維の分子動力学シミュレーション」第2回新学術領域研究「動的秩序と機能」若手研究会, 西浦温泉ホテルたつき, 2015.10.5-7, 愛知, (招待講演)
 18. S..G..Itoh, New molecular dynamics simulation methods to enhance conformational sampling for biomolecules, 東京大学物性研究所 理論セミナー, 2015.5.8, 柏 (Invited talk)
 19. Hisashi Okumura, Nonequilibrium and generalized-ensemble molecular dynamics simulations for amyloid fibril, 11th International Conference of Computational Methods in Sciences and Engineering, 2015.3.20, Athens, Greece (Invited talk)
 20. 奥村久士, アミロイド線維の形成初期過程と破壊の分子動力学シミュレーション
スーパーコンピュータワークショッピング 2015, 2015.1.29, 岡崎 (招待講演)
 21. Hisashi Okumura, All-atom molecular dynamics simulations for amyloid fibril assembly and disassembly, The 3rd International Symposium "Dynamical ordering of biomolecular systems for creation of integrated functions", 2015.1.11, Shima (Invited talk)
 22. Hisashi Okumura, Thermodynamics and free energy calculation, 11th Thai Summer School of Computational Chemistry "Replica exchange molecular dynamics simulation", 2015.1.7, Nan (Thailand) (Invited talk)
 23. Hisashi Okumura, Molecular dynamics simulation and temperature replica-exchange method
11th Thai Summer School of Computational Chemistry "Replica exchange molecular dynamics simulation", 2015.1.6, Nan (Thailand) (Invited talk)
 24. Hisashi Okumura, All-atom molecular dynamics simulations of amyloid-fibril disruption and peptide oligomerization Mini Symposium, 2014.12.17, Okazaki (Invited talk)

25. 奥村久士, 生体分子系, 液体系における分子動力学シミュレーション手法の開発と応用
第 28 回分子シミュレーション討論会, 2014.11.13, 仙台 (学術賞受賞講演)
26. 奥村久士, Molecular dynamics simulations of A β amyloid fibrils, 岡崎統合バイオサイエンスセンターリトリート, 2014.11.5, 岡崎 (招待講演)
27. 奥村久士, アミロイド線維の破壊と形成初期過程の分子動力学シミュレーション
第二回 CUTE シンポジウム, 2014.10.30, 津 (招待講演)
28. 奥村久士, 各種統計アンサンブルの生成法, 第 8 回分子シミュレーションスクール—基礎から応用まで—, 2014.10.15, 岡崎 (招待講演)
29. Hisashi Okumura, Molecular dynamics simulations for amyloid fibril disruption and dimerization of amyloid- β peptides 2nd International Conference on Computational Science and Engineering,), 2014.8.22 Ho Chi Minh City, Vietnam (Invited talk)
30. Hisashi Okumura, Generalized-ensemble molecular dynamics simulations, 2014 UST-Sokendai Joint Seminar on Computational Sciences, 2014.7.31, Daejeon (Korea) (Invited talk)
31. 奥村久士, キャビテーションによるアミロイド破壊の非平衡分子動力学シミュレーション
山手イブニングセミナー, 2014.6.6, 岡崎 (招待講演)
32. Hisashi Okumura, Generalized-ensemble algorithms to determine free-energy landscape of proteins 10th International Conference of Computational Methods in Sciences and Engineering, 2014.4.4, Athens, Greece (Invited talk)
33. S. G. Itoh and H. Okumura, Free-energy calculation by the replica-permutation method for biomolecules, 10th International Conference of Computational Methods in Sciences and Engineering, 2014.4.4, Athens, Greece (Invited talk)

【加藤晃一】

1. Takumi Yamaguchi, Design and creation of neoglycoconjugates for the application of biofunctional oligosaccharides, JAIST Japan-India Symposium on Materials Science 2018 2018.3.5, 能美(Invited talk)
2. 佐藤匡史, 加藤晃一, 糖タンパク質品質管理メカニズムの構造基盤解明, 第 46 回結晶成長国内会議 (JCCG-46) 2017.11.28, 浜松 (Invited talk)
3. Koichi Kato, Structural aspects of glycosylation as potential drug target, The 19th Symposium on Advanced Concepts in New Drug Developmemt : 1st Ewha-NCU Joint Symposium, 2017.11.16, Seoul(Invited talk)
4. Koichi Kato, Isotope Labeling Approaches for Carbohydrates and Glycoproteins, 第 56 回 NMR 討論会, 2017.11.15, 八王子(Invited talk)
5. Koichi Kato, Dynamic ordering of biomolecular systems coupled with creation of integrated functions CU-IMS collaborative seminar, 2017.10.31, Bangkok (Invited talk)
6. Koichi Kato, Dynamic ordering of biomolecular systems coupled with creation of integrated functions The NANOTEC-IMS Joint Research Meeting, 2017.10.30, Pathum (Invited talk)
7. Koichi Kato, Maho Yagi-Utsumi, Saeko Yanaka, Tatsuya Suzuki, Hirokazu Yagi, Tadashi Satoh, and Takumi Yamaguchi, Structural mechanisms of carbohydrate functions, Taiwan-Japan Biomedical Symposium on Magnetic Resonance, 2017.10.15, Tainan (Invited talk)
8. Kensuke Kurihara, Constructive biology approach to artificial cells: Artificial cells based on vesicles from nonliving materials, 6th International Conference in Biology, Chemistry and Agronomy, 2017.9.27, Guadalajara, Mexico (Keynote lecture)
9. 加藤晃一, 抗体を源流とする生命分子構造学の展開, 静岡県立大学薬学部 第 264 回 月例薬学セミナー, 2017.9.22, 静岡
10. 矢木宏和, 谷中冴子, 輿語理那, 鈴木達哉, 山口拓実, 杉山正明, 加藤晃一, 統合的な構造生物学アプローチによる糖タンパク質および糖鎖の構造ダイナミクスの解析, Integrative structural biology approaches for understanding conformational dynamics of oligosaccharides and glycoproteins 第 55 回 日本生物物理学会年会, 2017.9.20, 熊本
11. 加藤晃一, 糖鎖の生命分子科学の探究, 第11回 分子科学討論会, 2017.9.17, 仙台
12. Saeko Yanaka, Stable isotope labeling for NMR observation of antibody glycoproteins in serum environments, Seminar at Max Planck Institute for Biophysical Chemistry, 2017.9.14, Göttingen (Invited talk)
13. Koichi Kato, Structural views of fate determination of glycoproteins in cells, Seminar at Max Planck Institute for Biophysical Chemistry, 2017.9.14, Göttingen (Invited talk)

14. Saeko Yanaka, Integrative approach for exploration and creation of dynamical ordering of biomolecular systems, 14th HORIZONs in Molecular Biology, 2017.9.11-14, Göttingen (Invited talk)
15. 矢木宏和, 糖タンパク質糖鎖の構造解析法の開発と糖鎖機能解析への応用, 第 2 回 G-CHAIN セミナー, 2017.9.8, 岐阜
16. 佐藤匡史, 鈴木達哉, Gengwei Yan, 谷中汎子, 矢木宏和, 山口拓実, 加藤晃一, レクチンによる動的な糖鎖の認識機構, 第 4 回レクチン研究会, 2017.9.4, 東京 (招待講演)
17. Hirokazu Yagi, The characterization of the laminin-binding glycans on alpha-dystroglycan catalyzed by several causative gene products of dystroglycanopathy, The 1st International Conference on the Glycobiology of Nervous System, 2017.9.2, Seoul (Invited talk)
18. 栗原頸輔, 松尾宗征, 化学的人工細胞モデルで探る生命起源, 日本進化学会第 19 回年次大会, 2017.8.24, 京都 (招待講演)
19. 矢木宏和, 糖タンパク質糖鎖の機能解明のための構造解析技術の開発と応用, 第 1 回触発型有機化学研究会, 2017.8.5, 名古屋 (招待講演)
20. 加藤晃一, プロテアソーム分子集合の構造生物学, 大阪大学蛋白質研究所セミナー・SPring-8 先端利用技術ワークショップ, SPring-8 における蛋白質構造生物学研究の現状と将来 2017.8.4, 吹田 (招待講演)
21. 加藤晃一, 抗体と糖鎖の NMR, NMR 共用プラットフォームシンポジウム - 高磁場 NMR を核としたプラットフォームの新たな展開 -, 2017.8.2, 横浜 (招待講演)
22. 加藤晃一, 生体分子の集合離散が織りなす細胞機能研究の最前線, 第 69 回 日本細胞生物学会大会, 2017.6.13, 仙台 (招待講演)
23. 加藤晃一, 生命分子システムの動的秩序形成の探査・創生・展開 —領域内共同研究の成果を中心に—, 新学術領域研究「動的秩序と機能」全体班会議, 2017.6.3, 恩納村, 沖縄 (招待講演)
24. Kensuke Kurihara, Strategy for origin of life research using chemical artificial cells ELSI International Workshop “Reconstructing the Phenomenon of Life –To Retrace the Emergence of Life –”, 2017.5.30, Tokyo (Invited talk)
25. 矢木・内海真穂, タンパク質の動的構造と分子集合メカニズムの理解を目指して, 第 169 回 薬学談話会, 2017.5.24, 名古屋 (招待講演)
26. 矢木宏和, 糖タンパク質における糖鎖の生物機能と高次構造への影響, 第 58 回 日本生化学会中国四国支部例会, 2017.5.20, 香川 (招待講演)
27. 矢木宏和, タンパク質における糖鎖修飾の生物学的機能と高次構造への影響, 第 1 回名古屋市立大学若手イブニングセミナー, 2017.4.26, 名古屋 (招待講演)
28. Maho Yagi-Utsumi, Arunima Sikdar, Tadashi Satoh and Koichi Kato, Versatile structural architectures of archaeal homolog of proteasome assembly chaperone, Frontier Bioorganization Forum 2017: Dynamical ordering and integrated functions of biomolecular systems, 2017.4.25, Taipei (Invited talk)
29. Koichi Kato, Structural insights into dynamic orchestration of biomolecular systems Frontier Bioorganization Forum 2017: Dynamical ordering and integrated functions of biomolecular systems, 2017.4.24, Taipei (Invited talk)
30. 加藤晃一, 谷中汎子, 矢木宏和, バイオ医薬品への構造生物学的アプローチ, 日本薬学会第 137 年会, 2017.3.27, 仙台 (招待講演)
31. 加藤晃一, タンパク質の運命決定と機能発現の分子構造ダイナミクス研究, 細胞・システム作動機構の理解に向けた, 生体タンパク質分子の構造と機能のダイナミクス研究の拠点形成 平成 28 年度末 シンポジウム, 2017.3.13, 岡崎 (招待講演)
32. Koichi Kato, Structural glycobiology by stable isotope-assisted NMR spectroscopy, Advances Isotopic Labeling Meeting for Integrated Structural Biology 2017, 2017.3.7, Grenoble (Invited talk)
33. Koichi Kato, Structural glycobiology by stable isotope-assisted NMR spectroscopy, Advances Isotopic Labeling Meeting for Integrated Structural Biology 2017..3.7, Grenoble (Invited talk)
34. 栗原頸輔, “化学的人工細胞が提案する生命起源への戦略”, ABC プロジェクトミニ WS 「低温度星周りの光合成」, 2017.3, 東京 (招待講演)
35. Hirokazu Yagi, Development and application of glycosylation-profiling techniques for functional glycomics in the nervous system, 11th International Symposium on Cell Surface Macromolecules, 2017.2.26, Punjab(Invited talk)
36. Hirokazu Yagi, Development and application of glycosylation-profiling techniques for functional glycomics in the nervous system, 11th International Symposium on Cell Surface Macromolecules, 2017.2.26, Punjab (Invited talk)

37. Koichi Kato, Maho Yagi-Utsumi, Saeko Yanaka, Tatsuya Suzuki, Hirokazu Yagi, Tadashi Satoh, and Takumi Yamaguchi, NMR views of functional roles of glycoconjugates of biological and pharmaceutical interest, 7th Asia Pacific NMR Symposium & 23rd Annual Meeting of NMRS-India, 2017.2.18 Bangalore(Invited talk)
38. Koichi Kato, Maho Yagi-Utsumi, Saeko Yanaka, Tatsuya Suzuki, Hirokazu Yagi, Tadashi Satoh, and Takumi Yamaguchi, NMR views of functional roles of glycoconjugates of biological and pharmaceutical interest, 7th Asia Pacific NMR Symposium & 23rd Annual Meeting of NMRS-India 2017.2.18, Bangalore (Invited talk)
39. 加藤晃一, 生命分子システムの秩序形成のダイナミクス, 次世代脳プロジェクト, 2016.12.19, 国立 (招待講演)
40. 栗原顕輔, “化学で創る人工細胞”,細胞を創る研究会 9.0, 2016.11, 東京 (招待講演)
41. Saeko Yanaka, Rina Yogo, Hirokazu Yagi, and Koichi Kato, Characterization of antibody interactions in serum environments, The 9th Korea-Japan Seminars on Biomolecular Sciences: Experiments and Simulations, 2016.11.15, Gyeongju (Invited talk)
42. Koichi Kato, Structural insights into proteasome orchestration mechanisms The 9th Korea-Japan Seminars on Biomolecular Sciences: Experiments and Simulations, 2016.11.14, Gyeongju (Invited talk)
43. Maho Yagi-Utsumi, Tadashi Satoh, and Koichi Kato, Structural basis for interactions of molecular chaperones with intrinsically disordered proteins The 9th Korea-Japan Seminars on Biomolecular Sciences: Experiments and Simulations, 2016.11.14, Gyeongju (Invited talk)
44. Koichi Kato, Structural basis for dynamic orchestration of proteasomes, The 42nd Naito Conference, 2016.10.6, 札幌 (Invited talk)
45. Koichi Kato, Carbohydrate dynamics that determine protein fates and functions, IGER International Symposium on Science of Molecular Assembly and Biomolecular Systems 2016, 2016.9.30, 名古屋 (Invited talk)
46. Koichi Kato, Takumi Yamaguchi, and Tadashi Satoh, Structural basis of glycoprotein-fate determination in cell, The 16th KIAS Conference on Protein Structure and Function, 2016.9.23, Seoul (Invited talk)
47. Koichi Kato, Structural views of glycofunctions of biological and pharmaceutical interest, The 5th International Symposium on Drug Discovery and Design by NMR, 2016.8.29, Yokohama (Invited talk)
48. K.Kurihara, “Constructive biology approach to artificial cell”, OIBB Summer School 2016, Okazaki, 2016.8, Japan (Invited talk)
49. Koichi Kato, Dynamic ordering of biomolecular and artificial systems, OIIB Summer School 2016, 2016.8.19, 岡崎 (Invited talk)
50. 加藤晃一, バイオ医薬品の構造解析 : 基礎と応用, 技術情報協会セミナー, 2016.8.5, 東京 (招待講演)
51. 佐藤匡史, 加藤晃一, 糖鎖認識を介したタンパク質品質管理機構の構造基盤平成 28 年度日本応用糖質科学会 東日本シンポジウム, 2016.7.22, 東京 (招待講演)
52. Koichi Kato, Structural views of glycan-dependent determination of glycoprotein fates in cells 28th International Carbohydrate Symposium, 2016.7.19, New Orleans (Keynote Lecture)
53. Koichi Kato, Platform for Integrative Biological Sciences 28th International Carbohydrate Symposium, 2016.7.17, New Orleans (Invited talk)
54. 加藤晃一, 糖鎖の生命分子構造学の探査・創生・展開, 生体機能関連化学部会若手の会 第 28 回サマースクール, 2016.7.15, 西浦温泉 (招待講演)
55. 栗原顕輔, “柔らかい分子集合体で創る人工細胞”, 第一回オルガネラ生理学研究会, 2016.7, 岡崎 (招待講演)
56. 加藤晃一, 多彩な糖鎖の構造・ダイナミクス・相互作用の分子科学, 第 43 回生体分子科学討論会, 2016.6.24, 名古屋 (招待講演)
57. 佐藤匡史, Tong Zhu, 年森隆泰, 山口拓実, 加藤晃一, 糖鎖を目印としたタンパク質品質管理機構の構造基盤, 第 16 回日本蛋白質科学会年会, 2016.6.9, 福岡 (招待講演)
58. Koichi Kato, Structural views of glycoprotein quality control in cells 2nd KU-IMS Symposium, 2016.6.3, Bangkok (Invited talk)
59. 加藤晃一, 糖鎖の生命分子構造学 第 371 回 CBI 学会講演会, 2016.4.23, 大阪 (招待講演)
60. 加藤晃一, 「糖鎖をみる」, 第 4 回バイオイメージング研究会, 2015.5.8, 東京
61. 加藤晃一, 「生命分子の自己組織化のダイナミクス」, 日本化学会第 95 春季年会 特別企画「生命および人工分子システムにおける動的秩序形成 : 分子論的理解」, 2015.3.26, 船橋

62. 加藤晃一, 「複合糖質の構造生物学と創薬」, 第 4 回岐阜構造生物学・医学・論理的創薬研究会シンポジウム, 2015.3.5, 岐阜
63. K. Kato, "Conformational dynamics and interactions of oligosaccharides and glycoconjugate," The 4th International Symposium on Drug Discovery and Design by NMR, 2015.2.4, 横浜
64. K. Kato, "Structural views of glycans in physiological and pathological contexts," CU-IMS workshop, 2015.1.24, Bangkok, Thailand,
65. K. Kato, "Biophysical exploration of biomolecular systems characterized by conformational dynamics and dynamical assembly," National Chiao Tung University Seminar, 2014.12.12, Hsinchu, Taiwan
66. T. Satoh, T. Toshimori, K. Suzuki, T. Yamaguchi, G. Yan, T. Zhu and K. Kato, "Structural basis for recognition of the terminal glucose tag of N-glycans as fate-determinant of glycoproteins in cells," The 7th Korea-Japan Seminars on Biomolecular Sciences: Experiments and Simulations, 2014.11.26-27, Seoul, Korea
67. K. Kato, T. Yamaguchi, M. Yagi-Utsumi, H. Yagi and T. Satoh, "A multilateral approach for structural characterization of dynamic organization of flexible biomolecules," Okazaki Institute for Integrative Bioscience Retreat, 2014.11.5, Okazaki
68. 加藤晃一, 「X 線と NMR による抗体の構造解析」, 第 87 回日本生化学会大会, 2014.10.18, 京都
69. 加藤晃一, 「タンパク質機能の制御に関する N 型糖鎖構造の多様性と多型性」, 第 87 回日本生化学会大会, 2014.10.17, 京都
70. 加藤晃一, 「NMR と SANS による生命分子のダイナミクス研究」, 平成 26 年度第 1 回生物構造学研究会, 2014.10.3, 東京
71. K. Kato, "NMR exploration of dynamic conformations and interactions of oligosaccharides and glycoconjugates," The 6th Iberoamerican NMR meeting // IV Iberian NMR meeting // VII Reunion Bienal del GERMN, 2014.9.24, Alcala de Henares, (Spain) (Keynote lecture)
72. K. Kato, "Structural basis for fate determination and functional regulation of proteins mediated by sugar chains," The Cordeliers Research Center Seminar, 2014.9.19, Paris, France
73. K. Kato, "Structural views of physiological and pathological roles of glycans," Academia Sinica Institutional Lecture, 2014.9.11, Taipei, Taiwan
74. 加藤晃一, 「生命分子の動的秩序形成におけるミクロ-マクロ相関の探査と設計原理の探求」, 新学術領域「動的秩序と機能」平成 26 年度全体班会議, 2014.8.4-7, 小松
75. K. Kato and T. Satoh, "Dynamic ordering in proteasomal subunit assembly," 第 14 回日本蛋白質科学年会, 2014.6.26, 横浜
76. K. Kato, "Dynamic orchestration of proteasomes," University of Cambridge Cambridge Seminar, 2014.5.23, Cambridge (UK)
77. K. Kato, "Structural views of glycosylation as potential drug target," The 4th Asia Pacific Protein Association (APPA) Conference, 2014.5.18, Jeju (Korea)
78. 加藤晃一, 「NMR を応用した糖鎖の動的構造解析」, よこはま NMR 構造生物学研究会 第 49 回ワークショップ, 2014.3.20, 横浜
79. 加藤晃一, 「超高磁場 NMR 分光法を中心としたタンパク質の高次構造・相互作用解析」, 第 2 回 ISIT ナノ・バイオフォーラム, 2014.3.5, 福岡
80. 加藤晃一, 「生命分子の揺らぎと秩序形成」, 独) 日本学術振興会 分子系の複合電子機能第 181 委員会, 2014.2.25, 木津川,
81. K. Kato, "NMR approaches for elucidating the functional roles of carbohydrate chains," 20th Symposium of National Magnetic Resonance Society (NMRS-2014), 2014.2.3, Assam, India
82. K. Kato, "Conformational dynamics of oligosaccharides characterized by paramagnetism-assisted NMR spectroscopy in conjunction with molecular dynamics simulation," The 10th International Symposium on Biochemical roles of Eukaryotic Cell Surface Macromolecules, 2014.1.21, Kolkata India
83. K. Kato and T. Satoh, "Exploration of micro-macro relationships in dynamic ordering of biomolecular systems and their underlying design principles," The 2nd International Symposium on Dynamical Ordering of Biomolecular Systems for Creation of Integrated Functions, 2014.1.11, Kyoto, Japan
84. K. Kato, "Mechanistic insights into dynamic orchestration of proteasomes," Pure and Applied Chemistry International Conference (PACCON) 2014, 2014.1.8, Khon Kaen, Thailand
85. K. Kato, "Dynamic assembly of proteins involved in the ubiquitin-/proteasome-mediated protein degradation system," Joint IMS-KU Workshop on Molecular Sciences towards Green Sustainability, 2014.1.6, Bangkok, Thailand

86. K. Kato, "Structural biology of glycoconjugates: Sugar chains as therapeutic targets," Yonsei-IMS Seminars on Biomolecular Sciences: Protein Structure and Diseases, 2013.12.16, Busan, Korea
87. K. Kumoi, T. Satoh, K. Murata, T. Hiromoto, T. Mizushima, Y. Kamiya, M. Noda, S. Uchiyama, M. Sugiyama, H. Yagi and K. Kato, "An archaeal homolog of proteasome assembly chaperone forms a homotetramer and functions as proteasome activator," Sixth Korea-Japan Seminars on Biomolecular Science: Experiments and Simulation, 2013.11.25, Okazaki, Japan
88. 加藤晃一, 「タンパク質社会の秩序維持における糖鎖の役割」, お茶の水女子大学 糖鎖科学教育研究センター第6回公開シンポジウム, 2013.11.23, 東京,
89. 山口拓実, 加藤晃一, 「生命分子アッセンブリーにおける分子間相互作用のダイナミクスの精密解析」, 山田研究会・統合バイオサイエンスシンポジウム, 2013.11.20, 田原
90. 加藤晃一, 「糖鎖を見る: 分子模型からムービーへ」, 生理学研究所研究会「構造の多様性に立脚した糖鎖機能の解明に向けて」, 2013.11.15, 岡崎,
91. 加藤晃一, 「糖鎖の機能解明を目指したNMRアプローチ」, 第52回NMR討論会, 2013.11.12, 金沢
92. K. Kato, "NMR characterization of dynamic conformational ensembles of oligosaccharides and intermolecular interactions in glycolipid clusters," 5th Asia-Pacific NMR Symposium 2013, 2013.10.28, Brisbane (Australia),
93. 山口拓実, 「常磁性効果を活用した糖鎖の立体構造解析-NMRと分子動力学計算によるコンフォメーション空間の探査-」, 日本分光学会NMR分光部会 平成25年度講習会, 2013.10.16, 名古屋,
94. 加藤晃一, 「生命分子の動的秩序形成におけるミクロ・マクロ相關の探査と設計原理の探求」, 新学術領域研究「生命分子システムにおける動的秩序形成と高次機能発現」第1回公開シンポジウム, 2013.10.2, 岡崎
95. 加藤晃一, 「バイオ医薬品の構造をみる」, 日本バイオイメージング学会 第22回学術集会, 2013.9.16, 東京
96. K. Kato, "Atomic Anatomy of Antibody as Glycoprotein," World Biopharma Week China Focus 2013, 2013.9.5, Shanghai (China)
97. 加藤晃一, 「NMRを用いた生命分子のダイナミクス解析」, 第2回立命館大学BKC生体分子ネットワークセミナー, 2013.8.20, 草津
98. 加藤晃一, 「生命分子システムにおける動的秩序形成と高次機能発現」, 国際高等研研究プロジェクト「分子基盤に基づく生体機能ネットワークとダイナミクスの解明」第2回研究会, 2013.8.9, 木津川
99. 山口拓実, 加藤晃一, 「NMRを用いた糖鎖のコンフォメーション揺らぎとクラスター特性の解析」, 国際高等研研究プロジェクト「分子基盤に基づく生体機能ネットワークとダイナミクスの解明」第2回研究会, 2013.8.8, 木津川,
100. 加藤晃一, 「立体構造からみた糖鎖の不均一性」, 第32回 日本糖質学会年会, 2013.8.7, 大阪,
【菊地和也】
- 菊地和也, "GLUT4の糖鎖機能を可視化する局在痕跡イメージングプローブ", 日本農芸化学会2018年度大会, 2018.3.16, 名古屋 (招待講演)
 - Kazuya Kikuchi, "Real-time Intravital Imaging of pH Variation Associated with Osteoclast Activity Using BODIPY Based Two Photon Excitation Probes", 14th International Conference on Applied Bioinorganic Chemistry (ISABC14), 2017.6.8, Toulouse, France (Invited talk)
 - Kazuya Kikuchi, "Real-time Intravital Imaging of pH Variation Associated with Osteoclast Activity Using BODIPY Based Two Photon Excitation Probes", *The 8th Asian Biological Inorganic Chemistry Conference (AsBIC VIII)*, 2016.12.5-9, Auckland, New Zealand (Invited talk)
 - Kazuya Kikuchi, "Intracellular Protein Labeling by Functional Probes with Tunable Chemical Switches", *Molecular Sensors & Molecular Logic Gate 2016*, 2016.7.24-28, Bath, U.K.
 - Kazuya Kikuchi, "Real-time Intravital Imaging of pH Variation Associated with Osteoclast Activity Using BODIPY Based Two Photon Excitation Probes", *ICPP-9, 9th International Conference on Porphyrins and Phthalocyanines*, 2016.7.3-8, Nanjing, China
 - Kazuya Kikuchi, "Development of Multifunctional 19F MRI Contrast Agents with Fluorine-encapsulated Silica Nanoparticle", Pacifichem2015, 2015.12.15-20, Honolulu HI, U.S.A., (Invited talk)
 - Kazuya Kikuchi, "Highly Sensitive 19F MRI Probes with Tunable Chemical Switches Using Metal Complexes", ICBIC-17, International Conference on Biological Inorganic Chemistry, 2015.7.20-24, Beijing, China (Invited talk)
 - Kazuya Kikuchi, "Development of Multifunctional 19F MRI Contrast Agents with Fluorine-encapsulated Silica Nanoparticle", 13th International Symposium on Applied Bioinorganic Chemistry (ISABC13), 2015.6.12-15, Galway, Ireland (Invited talk)

9. Kazuya Kikuchi, Development of Highly Sensitive ^{19}F MRI Probes with Tunable Chemical Switches, The 7th Asian Biological Inorganic Chemistry Conference (AsBIC VII), 2014.12.4, Gold Coast, Australia (Invited talk)
10. Kazuya Kikuchi, Intracellular Protein Labeling by Functional Probes with Tunable Chemical Switches Labeling and Nanoscopy, 2014.9.26, Heidelberg, Germany (Invited talk)
11. 菊地和也, 生体内において、分子や細胞の機能を追跡可能な分光情報に変換する化学プローブ, 有機合成夏季セミナー－明日の有機合成化学－, 2014.8.28, 大阪 (招待講演)
12. Kazuya Kikuchi, Development of Highly Sensitive ^{19}F MRI Probes with Tunable Chemical Switches Gordon Research Conferences, Metals in Medicine, 2014.6.17, New Hampshire, USA (Invited talk)

【佐甲靖志】

1. Rho Maeda, Takeshi Sato, Kenji Okamoto, Yasushi Sako, Lipid-protein cooperativity in the regulation of juxtamembrane domain dimer formation in epidermal growth factor receptor 第 55 回日本生物物理学会年会, 2017.9.20, 熊本(Invited talk)
2. Kenji Okamoto, Ryo Maeda, Yasushi Sako, Single-molecule FRET measurement for EGFR-RAS-MAPK signal transduction pathway, Deciphering complex energy landscape and kinetic network from single molecules to cells: a new challenge to make theories meet experiments, 2017.9.3-8, Dijon (France) (Invited talk)
3. 佐甲靖志, 細胞膜受容体の運動・会合体形成による情報伝達, 第 69 回日本細胞生物学会大会, 2017.6.13, 仙台 (招待講演)
4. 佐甲靖志, 細胞膜受容体の 1 分子可視化, 第 25 回日本バイオイメージング学会学術集会, 2016.11.21, 名古屋 (招待講演)
5. 佐甲靖志, 細胞膜受容体の動的会合体形成と機能, 13 回 JCGG シンポジウム, 2015.10.1, 名古屋
6. 佐甲靖志, 細胞膜受容体の会合と機能の 1 分子イメージング, 日本薬学会第 135 年会 シンポジウム「タンパク質の会合・凝集：計測、メカニズムから制御まで」, 2015.3.26, 神戸 (招待講演)
7. 佐甲靖志, 細胞内反応ネットワークの 1 分子計測, 日本学術会議フォーラム「生命情報ビッグデータ時代における新しい生命科学」, 2014.8.29, 東京 (招待講演)
8. Yasushi Sako, Experimental aspects of intracellular dynamics of cell fate decision, The Joint Annual Meeting of The Japanese Society for Mathematical Biology and The Society for Mathematical Biology 2014, Osaka, 2014.7.30, Osaka (Invited talk)
9. Yasushi Sako, Single-molecule analysis of cell signaling reactions, Single Protein Dynamics *in Cellulo* SPDC 2014, 2014.4.25, Onna, (Invited talk)

【笹井理生】

1. Masaki Sasai, Fluctuating three-dimensional genome architecture and gene regulation, Gordon Research Conference on Protein Folding Dynamics, 2016.1.10-15, Galveston, Texas, USA (Invited talk)
2. Masaki Sasai, Tomoki Terada, Qing-Miao Nie, Dynamical energy landscape perspective on protein functioning, Deciphering molecular complexity from protein functions to cellular network: Symposium at the PacifiChem 2015 Meeting, 2015.12.15-16, Honolulu, Hawaii, USA (Invited talk)
3. Tokuda Naoko, Shin Fujishiro, Masaki Sasai, Chromatin Domains and Heterogeneous Transcription Activities, International Symposium on Chromatin Structure, Dynamics, and Function, 2015.8.23-26, 兵庫 (Invited talk)
4. Masaki Sasai, Eukaryotic gene regulation and chromosome architecture CSRC Workshop on Kinetics of Enzymes and Molecular Machines, The Beijing Computational Science Research Center, 2015.8.18-19, Beijing, China (Invited talk)
5. Masaki Sasai, Eukaryotic chromatin folding and gene regulation, 3rd International Workshop on Theoretical and Computational Physics, 2015.7.27-30, Đà Lạt, Vietnam, (Invited talk)
6. 笹井理生, 分子モーターの時空間スケール, 第 6 回分子モーター討論会, 東大駒場, 2015.6.13-14, 東京 (招待講演)
7. Masaki Sasai, Multi-funnel view of protein folding, The First Korean-Polish Conference on "Protein Folding: Theoretical and Experimental Approaches", Korea Institute for Advanced Study, 2015.5.24-28, Seoul, Korea (Invited talk)
8. Masaki Sasai, Epigenetic dynamics of gene regulation, Architecture, dynamics, and functionality of molecular biosystems, 分子科学研究所, 2015.3.31, Okazaki, (Invited talk)
9. 笹井理生, 蛋白質機能発現の動的エネルギーとスケープ理論, 第 359 回 CBI 学会研究講演会, 東京大

学, 2015.2.24, 東京 (招待講演)

10. 笹井理生, 遺伝子制御ダイナミクスの理解へ向けて, 第 20 回交通流のシミュレーションシンポジウム, 名古屋大学, 2014.12.4, 名古屋 (基調講演)
11. Masaki Sasai, Qing-Miao Nie, and Tomoki P. Terada, Dynamical energy landscape view of actomyosin motors, The 7th Korea-Japan Seminars on Biomolecular Sciences, Korea Institute for Advanced Study, 2014.11.27, ソウル (Invited talk)
12. Masaki Sasai, Toward a theory of gene regulation dynamics, 京大生命動態システム科学推進拠点事業大学院生セミナー, 京都大学, 2014.10.24, 京都 (招待講演)
13. Masaki Sasai, Allosteric dynamics of proteins Macromolecular dynamics: structure, function and diseases, Kavli Institute for Theoretical Physics, 2014.7.1, Beijing, (Invited talk)
14. Masaki Sasai, Dynamical Landscapes of motors and enzymes Macromolecular dynamics: structure, function and diseases, Kavli Institute for Theoretical Physics, 2014.6.27, Beijing (Invited talk)
15. 笹井理生, 徳田直子, 藤城新, ゲノム立体構造の動力学シミュレーション
第 14 回日本蛋白質科学会年会, 2014.6.25, 横浜 (招待講演)
16. Masaki Sasai, Naoko Tokuda, Shin Fujishiro, Fluctuating genome structure and gene regulation 8th IUPAP International Conference on Biological Physics, 2014.6.20, Beijing (Invited talk)

【佐藤健】

1. 佐藤健, 小胞体における輸送小胞形成の時空間制御, 第 13 回 日本糖鎖科学コンソーシアム, 2015.10.19, 名古屋
2. 佐藤健, 出芽酵母をモデルとした小胞体における輸送小胞形成の時空間制御, 第 186 回酵母細胞研究会, キリンビール(株) 横浜工場, 2014.7.11 (招待講演)

【申惠媛】

1. Shin, H.-W., Membrane dynamics and phospholipid flippase activity. Membrane Lipid Transporter Symposium 2018, 2018.9.27, Kyoto iCeMS (Invited talk)
2. 申惠媛, リン脂質フリップ活性による細胞膜の形状変化と細胞機能, 第 91 回日本生化学会大会. シンポジウム, 2018.9.25, 京都 (招待講演)
3. 申惠媛, 細胞膜脂質フリップペーゼの基質特異性と活性調節メカニズム, 大阪大学蛋白質研究所セミナービーム上の生物化学 2018, 2018.3.2, 大阪 (招待講演)
4. Shin, H.-W., Specific substrates for P4-ATPases and their regulation in mammalian cells. The 15th International Conference on Na, K-ATPase and Related Transport ATPases. 2017.9.27, Otsu, Japan (Invited talk)
5. 申惠媛, メンブレントラフィックによる P4-ATPase の活性調節機構, 第 89 回日本生化学会大会, 2016.9.27, 仙台 (招待講演)

【真行寺千佳子】

1. Chikako Shingyoji, Mechanical activity of dynein and its dynamical regulation underlying oscillatory movement of sperm flagella. International Workshop Dynein 2017, Awaji Yumebutai International Conference Center, 2017.10.29-11.1, Hyogo
2. Chikako Shingyoji, Mechanical activity of dynein and its dynamical ordering underlying oscillatory movement of sperm flagella, The 3rd International Symposium on Dynamical Ordering of Biomolecular Systems for Creation of Integrated Functions, 2015.1.10, Shima, Mie (Invited talk)

【杉山正明】

1. Masaaki Sugiyama, Integrative approach to protein structure and dynamics Protein structure and dynamics under high density condition = crowding condition, The 8th Taiwan-Japan Joint Meeting on Neutron and X-ray Scattering, 2018.3.14-17, Taiwan (Invited talk)
2. Masaaki Sugiyama, Study on intra- and inter-protein dynamics by neutron solution scattering The 17th KIAS Conference on Protein Structure and Function, 2017.9.21-23, Korea (Invited talk)
3. Masaaki Sugiyama, Investigation on domain motion of protein by neutron scattering Frontier Bioorganization Forum :Dynamical ordering and integrated functions of biomolecular systems, 2017.4.24-26, Taiwan (Invited talk)
4. Masaaki Sugiyama, Neutron Scattering for Bio-molecular Systems, Grand Challenges in Small-angle Scattering, 2017.3.19-21, Okazaki, Japan (Invited talk)
5. Masaaki Sugiyama, Subunit Kinetics in α -Crystallins, The 9th Korea-Japan Seminars on Biomolecular Sciences, 2016.11.14-16, Gyeongju, Korea. (Invited talk)
6. Masaaki Sugiyama, "Subunit Kinetics in Dynamical Proteins", Japan-Korea meeting on Neutron Science,

- 2016.1.8, Busan, Korea. (Invited talk)
7. Masaaki Sugiyama, "Subunit Kinetics in α -Crystallins", International Conference on the Lens 2015, 2015.12.6-11, Kona, Hawaii. (Invited talk)
 8. 杉山正明, 「中性子線が拓く化学の未来」, 中性子小角散乱の基礎と応用, 日本化学会, 第 95 回春季年会, 2015.3.29, 船橋市 (招待講演)
 9. 杉山正明, 中性子の中性子による中性子のため生物溶液散乱, 第 6 回 MLF シンポジウム, 2015.3.18, つくば (招待講演)
 10. 杉山正明, コントラスト変調及び重水素化で見るタンパク質の溶液中の静的・動的構造 第 37 回溶液化学シンポジウム, 2014.11.12, 佐賀 (招待講演)

【茶谷絵里】

1. 茶谷絵理, 線維前駆中間体の観察によるアミロイド核形成の機構解明と制御, 蛋白質異常凝集研究会, 2017.12.1, 大阪
2. 茶谷絵理, アミロイド形成における蛋白質分子集合プロセスの観察, 第 17 回 日本蛋白質科学会年会, 2017.6.22, 仙台
3. Eri Chatani, Naoki Yamamoto, Nucleation dynamics in amyloid formation as investigated by the analysis of prefibrillar intermediates, 新学術領域「動的秩序と機能」第 5 回国際シンポジウム, 2017.1.22, 東京(Invited talk)
4. Eri Chatani, Naoki Yamamoto, Investigation of protein aggregation processes leading to amyloid formation, Japan-Taiwan Medical Spectroscopy International Symposium, 2016.12.5, Awaji, Japan (Invited talk)
5. Eri Chatani, Takato Hiramatsu, Yukari Itakura, Roumiana Tsenkova, Investigation of Water Molecular System Dynamics in the Early Stages of Amyloid Formation, AQUAPHOTOMICS: UNDERSTANDING WATER in BIOLOGY 2nd INTERNATIONAL SYMPOSIUM, 2016.11.28, Kobe, Japan,(Invited talk)
6. Eri Chatani, Investigating early steps in amyloid fibril formation, 日中生物物理学会ジョイントシンポジウム : 蛋白質設計とバイオテクノロジーへの応用, 第 54 回日本生物物理学会年会, 2016.11.25, 茨城 (招待講演)
7. Eri Chatani, Naoki Yamamoto, Molecular Exploration of Prefibrillar Intermediates That Lead to Formation of Amyloid Fibrils, Indo-Japan Joint Seminar on "Frontiers in Molecular Spectroscopy: From Fundamentals to Applications on Material Science and Biology", 2016.11.14, Kanpur, India,
8. 茶谷絵理, 線維前駆中間体の観察によるアミロイド線維の伝播およびその発現機構の解明 第 63 回 日本生化学会 近畿支部例会, 2016.5.21, 神戸 (招待講演)
9. 茶谷絵理, アミロイド線維形成にみられるタンパク質初期会合メカニズムの解明, 日本化学会第 96 春季年会, 2016.3.24, 京都 (招待講演)
10. 茶谷絵理, 平松貴人, 板倉由佳利, 増田裕輝, Roumiana Tsenkova, アミロイド線維の伝播核形成過程における水の関与性, BMB2015, 神戸, 2015 年 12 月 1 日 (招待講演)
11. 茶谷絵理, Roumiana Tsenkova, アミロイド線維の核形成に伴う水の動態解明, 第 31 回近赤外フォーラム, 筑波, 2015.11.26 (招待講演)
12. 茶谷絵理, アミロイド線維形成初期に見られる水構造の変化, 京都大学原子炉実験所「第 8 回タンパク質の異常凝集とその防御・修復機構に関する研究会」, 2015.11.19, 大阪 (招待講演)
13. Eri Chatani, Exploring the early stages of protein association in the formation of amyloid fibrils IPR and ANU joint symposium "Protein structure and function", 2015.11.16, Australia (Invited talk)
14. Eri Chatani, Yuki Masuda, Yutaro Tsuchisaka, Takato Hiramatsu, Yukari Itakura, and Roumiana Tsenkova, Exploring the early stages of generation of amyloid fibrils from the viewpoint of water molecular system dynamics, The 15th KIAS Conference on Protein Structure and Function, 2015.9.18, Korea(Invited talk)
15. Eri Chatani, Recent progress in understanding the early stages of generation of amyloid fibrils Kwansei Gakuin-Kobe University Joint Symposium: International Symposium on Recent Advanced in Near Infrared Spectroscopy, 2015.9.5, Sanda, Hyogo (Invited talk)
16. Eri Chatani, Investigation of molecular mechanism for the generation of amyloidogenic nuclei 第 15 回 日本蛋白質科学会年会, 2015.6.25, 横浜 (招待講演)
17. Naoki Yamamoto, Eri Chatani, Atsuo Tamura, and Keisuke Tominaga, Temperature and Hydration Dependence of the Complex Dielectric Spectra of Proteins at GHz and THz Regions Obtained by THz Time-domain Spectroscopy and Dielectric Spectroscopy, AWEST2015-The 3rd Awaji International

- Workshop on "Electron Spin Science & Technology: Biological and Materials Science Oriented Applications", 2015.6.17, Awaji, Hyogo
18. 茶谷絵理, アミロイド線維形成における蛋白質初期集合メカニズムの解明, 日本薬学会第 135 回年会, 2015.3.26, 兵庫
 19. 茶谷絵理, アミロイド伝播核の形成メカニズムの解明, 京都大学原子炉実験所・専門研究会 第 7 回タンパク質の異常凝集とその防御・修復機構に関する研究会, 2014.12.11, 大阪
 20. Eri Chatani, Association mechanism of protein molecules in the formation of amyloid fibrils Indo-Japan Joint Workshop on "Frontiers in Molecular Spectroscopy: Fundamentals and Applications to Material and Biology", 2014.11.26, 奈良
 21. Eri Chatani, Understanding early events in protein assembly for the amyloidogenic nucleation Japan-Hungary Seminar "Mechanism and regulation of aberrant protein aggregation", 2014.11.20, 大阪
 22. Eri Chatani, Investigation of early association of protein molecules during the amyloidogenic nucleation, 2nd Awaji International Workshop on "Electron Spin Science & Technology: Biological and Materials Science Oriented Applications", 2014.6.16, 兵庫

【寺内一姫】

1. 寺内一姫, 時計タンパク質 KaiC の探究, 藍藻の分子生物学 2017, 2017.12.1, 千葉 (招待講演)
2. 寺内一姫, ブルーネイティブ電気泳動による時計タンパク質 KaiC の動的構造解析, 第 68 回日本電気泳動学会総会, 2017.11.25, 広島 (招待講演)
3. 寺内一姫, シアノバクテリアがおしえてくれること, 日本光合成学会若手の会 第 13 回セミナー, 2017.5.28, 大津 (招待講演)
4. 寺内一姫, 試験管内の概日時計 : 発振子の正体を追って 北山陽子メモリアルシンポジウム・光合成生物の概日時計機構, 2017.3.11, 名古屋 (招待講演)
5. 寺内一姫, 3 つのタンパク質で再構成できるシアノバクテリアの生物時計, 第 53 回日本生物物理学会年会 シンポジウム「ATP 加水分解が介在する時空間動秩序の形成」, 2015.9.14, 金沢 (招待講演)
6. 寺内一姫, シアノバクテリアの概日時計から学ぶ, 微細藻類研究会 2014, 2014.12.23, 岡崎 (招待講演)
7. 大山克明, シアノバクテリア時計タンパク質 KaiABC の概日リズム生成機構解明に向けて
第 11 回光合成学会若手の会セミナー, 2014.12.7, 草津 (招待講演)

【水野健作】

1. 水野健作, 藤原佐知子, 大橋一正, 動物細胞の力覚応答における細胞骨格の役割, 第 39 回 日本分子生物学会年会, 2016.12.2, 横浜 (招待講演)
2. Kensaku Mizuno, Dynamic reordering of actin cytoskeleton in mechanical force-induced cell responses, The 4th International Symposium on Dynamic Ordering of Biomolecular Systems for Creation of Integrated Functions, 2015.11.24, Fukuoka, Japan (Invited talk)
3. Kensaku Mizuno, Roles of actin dynamic ordering in mechanosensing and cell proliferation
The 53rd Annual Meeting of Japan Biophysics Society, 2015.9.14, Kanazawa, Japan (Invited talk)
4. 水野健作, 増殖抑制シグナル依存的な一次纖毛形成機構, 第 67 回 日本細胞生物学会年会, 2015.7.1, 東京 (招待講演)

【村田和義】

1. Kazuyoshi Murata, Structural analysis of Pithovirus sibericum by Cryo-EM, 第 12 回日本ゲノム微生物学会年会, 京大桂キャンパス, 2018.3.7, 京都 (招待講演)
2. 村田和義, 宮崎直幸, ソンチホン, 岡本健太, ピソウイルスのクライオ電子顕微鏡による構造解析, 2017 年度生命科学系学会合同年次大会, 神戸ポートピアホテル, 2017.12.6, 神戸 (招待講演)
3. Kazuyoshi Murata, Single Particle Zernike Phase Contrast Cryo-Electron Microscopy Combined with Direct Detector for Analyzing Membrane Proteins and their Dynamics, 第 89 回日本化学会大会, 2016.9.25, 仙台 (Invited talk)
4. 村田和義, クライオ電子顕微鏡によるタンパク質の単粒子構造解析, 日本バイオイメージング学会大 25 回 学術集会, 2016.9.6, 名古屋 (招待講演)
5. 村田和義, 超高圧電子顕微鏡トモグラフィーの生物応用, 日本顕微鏡学会第 72 回学術講演会, 2016.6.14, 仙台 (招待講演)
6. Kazuyoshi Murata, Dynamical ordering of virus particle for creation of integrated functions
The 4th International Symposium "Dynamical ordering of biomolecular systems for creation of integrated functions", 2015.11.22, Fukuoka, Japan (Invited talk)
7. Kazuyoshi Murata, Visualizing the Infection Process of T7-like Cyanophage in the Marine

Cyanobacterium Prochlorococcus by Cryo-Electron Tomography IGER International Symposium on Frontiers in Biological Research with Advanced Electron Microscope Technologies, 2015.1.16, Nagoya (Invited talk)

8. Kazuyoshi Murata, Advanced biological applications for high-voltage electron microscopes, International Symposium on Analytical Science and Technology, 2014.11.20, Daejeon, Korea

【山本量一】

1. 松田拓也, 山本量一, 細胞分裂を考慮した自発運動粒子モデルによる創傷治癒のシミュレーション, 研究会「凝縮系の理論化学」, 2018.3.13, 那覇市 (招待講演)
2. 多羅間充輔, 山本量一, 基盤上を運動する細胞の力学モデル, 研究会「凝縮系の理論化学」, 2018.3.13, 那覇市 (招待講演)
3. 山本量一, A particle-based minimal model for crawling and proliferating cells on substrate, 日本生物物理学会第 55 回年会, 2017.9.19, 熊本市 (招待講演)
4. Ryoichi Yamamoto, "The emergence of collective motion: from glass to biological systems" CerSJ-GOMD Joint Symposium on Glass Science and Technologies, 2016.11.16, Kyoto, Japan (Invited talk)
5. Ryoichi Yamamoto, "Collective Motion of Cells Crawling on a Substrate" The 4th International Conference on Molecular Simulation (ICMS2016), 2016.10.23-26, Shanghai, China (Invited talk)
6. 山本量一, 「自己組織化する細胞集団のモデリングとシミュレーション」, 化学工学会 第 48 回秋季大会特別シンポジウム「SP-3 新時代に向けたバイオプロセスイノベーション」, 2016.9.8, 徳島市 (招待講演)

班友

【澤田知久】

1. Tomohisa Sawada, Makoto Fujita, Peptide-entangled polyhedral by folding and assembly, 43rd International Conference of Coordination Chemistry, 2018.7.30, 仙台(Invited talk)
2. Tomohisa Sawada, Concerted folding-and-assembly of short peptides via coordination The 2018 Wolf Prize Symposium, 2018.5.30, Israel (Invited talk)
3. Tomohisa Sawada, Creation of Higher-order Peptidic Nanostructures by Concerted Folding and Assembly, 日本化学会第 98 回春季年会, 2018.3.20, 千葉 (Invited talk)
4. Tomohisa Sawada, Metal-peptide nanostructures with unique entanglements, Symposium on Frontier of Biofunctional Chemistry, 2017.12.12, Yokohama (Invited talk)
5. 澤田知久, 配位結合とペプチド配座を利用した新奇ナノ構造の創出, 最先端光計測とライフサイエンスの近未来—Bio. Phys. Chem. 三重点の探索—, 2017.9.6, 仙台 (招待講演)
6. 澤田知久, フォールディング&アセンブリー：ユニークなペプチド性ナノ空間の構築に向けた新戦略, 日本化学会東北支部山形地区講演会, 2016.11.24, 山形 (招待講演)
7. 澤田知久, 生体に学ぶ自己組織化：人工系における相互誘導適合, そしてフォールディング&アセンブリー 第 5 回 お茶の水女子大学 イブニングセミナー, 2014.6.28, 東京 (基調講演)

【重田育照】

1. Yasuteru Shigeta, "An Enhanced Sampling Method for Searching Conformational Changes of Proteins and Supramolecules", 22nd International Workshop on Quantum Systems in Chemistry, Physics and Biology (QSCP-XXII), 2017.10.16-24, Hunan Province, P.R. China.
2. Yasuteru Shigeta, "Cumulant Mechanics for Quantum and Statistical Physics", The 4th International Conference on Molecular Simulation (ICMS), 2016.10.23-26, Shanghai, China
3. Yasuteru Shigeta, "Integrated approach toward rational design of enzyme", 2016 2nd Japan-Thai workshop on theoretical and computational chemistry, Yokohama City University, 2016.9.21-22, Yokohama, Japan
4. Yasuteru Shigeta, "Simple conformational search methods for understanding biological functions", Shanghai Workshop on Frontiers in Molecular Biophysics, 2016.7.23-26, Shanghai, China
5. Yasuteru Shigeta, "Efficient Conformational Search Methods for Protein Folding Problems", The ninth Congress of the International Society for Theoretical Chemical Physics (ISTCP 2016), Grand Forks, 2016.7.17-22, North Dakota, USA
6. Yasuteru Shigeta, "A consistent scheme for accurately estimating acid dissociation constant (pK_a) and redox potential", 4th Changsha Workshop on Theoretical and Computational Chemistry 2016, 2016.6.10-12, Changsha, Hunan, China
7. Yasuteru Shigeta, "Inverse Histogram-based Sampling Algorithm for Protein-folding Problems", The Seventh Asia-Pacific Conference of Theoretical and Computational Chemistry (APCTCC 7), Kaohsiung, 2016.1.25-28, Taiwan

8. Yasuteru Shigeta, "Simple Conformational Search Algorithms For Protein Folding", 6th Czech-Slovakia-Japan Theoretical Chemistry meeting, 2015.10.11-14, Bratislava, Slovakia
9. 重田育照, “理論研究によるチトクロム c 酸化酵素の酸化還元と共に役割したプロトン輸送機構：入り口と出口”, 生物物理学会シンポジウム「膜を介したプロトン透過機構」, 2015.9.13-15, 金沢, 石川
10. 重田育照, “チトクロム c 酸化酵素におけるプロトン輸送の構造的仕掛け”, 分子研研究会「膜タンパク質内部のプロトン透過を考える」, 分子研, 2015.8.20-21, 愛知
11. 重田育照, “第一原理計算に基づく酵素機能解析：チトクロム c 酸化酵素のプロトンポンプを例として”, 東京大学物性研究所 「機能物性融合科学シリーズ(3)「反応と輸送」」, 2015.6.24-26, 柏, 千葉
12. 重田育照, “第一原理計算によるタンパク質の pKa と構造機能相関”, 日本物理学会シンポジウム「プロトネーション イントゥ ダークネス：生体分子機能理解の為の水素位置情報」, 早稲田大学, (注 平成 26 年度) 2015.3.21-24, 東京
13. 重田育照, 「高分散分子動力学法によるタンパク質でのレイアウトの検出」, レイアウトの理論科学ワークショップ, February 16th 2015, 日本原子力研究開発機構システム計算科学センター, 2015.2.16, 千葉 (招待講演)
14. 重田育照, “理論と実験の協奏的アプローチによる複合スピニル起子変換制御”, 新学術領域研究「高次複合光応答」第 2 回公開シンポジウム, January 23rd-24th 2015, 千里ライフサイエンスセンター, 2015.1.23-24, 大阪
15. 重田育照, “酵素活性制御に向けた多階層量子計算手法の応用”, 理研シンポジウム「生体分子系量子化学計算の最前線」, 理化学研究所, 2015.1.22-23, 和光
16. 重田育照, 「宇宙生命連携研究による物質進化過程の探索」27 回理論懇親シンポジウム, 国立天文台, 2014.12.24-26, 東京
17. Yasuteru Shigeta, “A Molecular Design of Nonlinear Optical Properties and Conductivity Switches on the Basis of Open-shell Nature”, 2014 Workshop on Innovative Nanoscale Devices and Systems (WINDS), 2014.11.30-12.5, Hawaii, USA
18. Yasuteru Shigeta, “Protein Folding Processes Detected by Enhanced Sampling Techniques”, 19th International Workshop on Quantum Systems in Chemistry and Physics, 2014.11.11-17, Tamsui, Taiwan
19. Yasuteru Shigeta, “Towards Theoretical Design of Catalytic Activities of Enzymes”, The Asia Hub for e-Drug Discovery Symposium 2014, Sichuan University (SCU), 2014.11.11-12, Chengdu, China.
20. Yasuteru Shigeta, “First-principles analysis on enzymatic degradation of nylon”, 2nd World Congress on Petrochemistry and Chemical Engineering, 2014.10.27-29, Las Vegas, USA
21. 重田育照, 「第一原理計算に基づくタンパク質機能デザイン」, 第 8 回 FMO 研究会 (CBI 学会 2014 年大会), タワーホール船堀, 2014.10.28-30, 東京
22. 重田育照, “CMD の思想に基づく生命物理学の研究”, 第 25 回コンピュテーションナル・マテリアルズ・デザイン (CMD) ワークショップ, 国際高等研究所, 2014.9.1-5, 京都
23. 重田育照, “計算化学による pKa の高精度算出法”, 第 1 回水和ナノ構造研究会, 奥平温泉, 2014.8.28-29, 岩手
24. Yasuteru Shigeta, “Computational Studies on Redox Potential of Metal Complexes and Model Cofactors”, International Conference on Synthetic Metals, 2014.6.30-7.5, Turku, Finland

【塚崎智也】

1. Takamitsu Haruyama, Yasunori Sugano, Yoshiki Tanaka, Hiroki Konno, Tomoya Tsukazaki, High-speed AFM observation of membrane protein embedded in Nanodisc, 第 55 回 日本生物物理学会年会, 2017.9.20, 熊本 (Invited talk)
2. Tomoya Tsukazaki, Arata Furukawa, Kunihito Yoshikai, Takaharu Mori, Hiroyuki Mori, Yusuke V. Morimoto, Yasunori Sugano, Shigehiro Iwaki, Tohru Minamino, Yuji Sugita, Yoshiki Tanaka, Protein Translocation Motor SecDF, 第 55 回 日本生物物理学会年会, 2017.9.19, 熊本 (Invited talk)
3. Tomoya Tsukazaki, Snapshots of the proton-driven protein translocation motor, EMBO conference | Protein translocation and cellular homeostasis, 2017.3.19, クロアチア・ドブロブニク
4. 塚崎智也, 新生鎖を膜透過させる Sec 蛋白質の X 線結晶構造解析, 第 16 回 日本蛋白質科学会年会, 2016.6.9, 福岡市 (招待講演)
5. Tomoya Tsukazaki, Structure of YidC reveals a mechanism of Sec-independent membrane protein insertion, The 25th Hot Spring Harbor International Symposium Cutting Edge of Technical Innovations in Structural and Systems Biology, 九州大学, 2015.11.14, 福岡市 (Invited talk)
6. Tomoya Tsukazaki, Structure and dynamics of Sec protein-conducting channel, 第 53 回日本生物物理学年会, 金沢大学, 2015.09.15, 金沢市 (Invited talk)
7. 塚崎 智也, 膜蛋白質結晶化技術の向上により解明された Sec/YidC による蛋白質輸送機構 第 3 回 生体分子サイエンスセミナー(東工大・すずかけキャンパス), 2015.08.31 (招待講演)

8. Tomoya Tsukazaki, Structures of Membrane Protein Insertase YidC, Gordon Research Conference 2015, Membrane Protein Folding, 2015.6.21, アメリカ, ボストン (Invited talk)
9. 塚崎智也, タンパク質を膜へと組み込む膜タンパク質 YidC の立体構造とその機能 第 1197 回ウイルス研究所セミナー/共同利用・共同研究拠点セミナー, 京都大学 2014.12.10, 京都市 (招待講演)
10. 塚崎智也, 熊崎薫, 千葉志信, 武本瑞貴, 古川新, 伊藤維昭, 石谷隆一郎, 濡木理
タンパク質の膜への組み込みに関わる膜タンパク質 YidC の結晶構造と作業機序
平成 26 年度日本結晶学会年会, 東京大学, 2014.11.3, 東京都文京区 (招待講演)
11. Tomoya Tsukazaki, Structure of YidC reveals a mechanism of Sec-independent membrane protein 2014 Joint Symposium, Integrative Microbiology, University of Minnesota, 2014.10.6, MN, USA
12. 塚崎智也, タンパク質を膜へと組み込む YidC の立体構造とその機能, 日本化学会 会員増強のための講演会, 山形大学, 2014.8.8, 山形市 (招待講演)
13. 塚崎智也, 春山 隆充, 菅野 泰功, 田中 良樹, 紺野 宏記, Sec タンパク質膜透過装置の活写にむけて, 第 14 回 日本蛋白質科学会年会, 2014.6.25, 横浜 (招待講演)
14. 塚崎智也, プロトン駆動型タンパク質分泌モーター SecDF, 分子研研究会「膜タンパク質内部のプロトンの透過を考える」, 2014.4.21, 岡崎市 (招待講演)
15. 塚崎智也, 熊崎薫, 千葉志信, 武本瑞貴, 古川新, 伊藤維昭, 石谷隆一郎, 濡木理, タンパク質膜組込み insertase YidC の立体構造と分子機構, 日本化学会第 95 春季年会, 日本大学, 2014.3.26, 船橋市 (招待講演)

【長田裕也】

1. 長田裕也, 高分子主鎖のキラリティスイッチングに基づいた新規機能性材料の創出, 2017 年液晶交流会, 弘前大学 文京町キャンパス, 2017.9.12, 弘前 (招待講演)
2. Yuuya Nagata, Development of New Chirality-switchable Materials Based on the Solvent-dependent Helix Inversion of Poly(quinoxaline-2,3-diyl)s, 10th China-Japan Joint Symposium on Functional Supramolecular Architectures, Wuhan University, 2017.5.14, China (Invited talk)
3. 長田裕也, 高分子主鎖のキラリティスイッチングに基づく新機能性材料の開拓, 第 65 回高分子学会年次大会, 神戸国際会議場・展示場, 2016.5.26, Kobe (招待講演)
4. 長田裕也, Development of Innovative Chiral Materials Based on Solvent-Dependent Helix Inversion of Macromolecular Backbone, 日本化学会 第97春季年会, 慶應義塾大学, 2016.3.17, 横浜市 (Invited talk)

【原野幸治】

1. 原野幸治, 分子科学研究ツールとしての原子分解能電子顕微鏡, 第 5 回錯体化学若手の会九州・沖縄支部勉強会, 2017.12.9, 熊本 (招待講演)
2. 原野幸治, 高分解能電子顕微鏡を駆使した分子集合体科学の探求, 2017 年度中国四国支部高分子講演会, 2017.12.5, 鳥取 (招待講演)
3. 原野幸治, 分子科学研究ツールとしての実時間原子分解能電子顕微鏡イメージング, 日本顕微鏡学会「その場観察研究部会」第 2 回研究会, 2017.11.17, 岡崎 (招待講演)
4. 原野幸治, 有機分子の結晶化を観察できるメガネ!-電子顕微鏡の最前線-, 第 7 回 CSJ 化学フェスタ, 2017.10.19, 東京 (招待講演)
5. 原野幸治, 原子分解能電子顕微鏡で明らかにする動的分子科学, 第 5 回バイオ関連化学シンポジウム若手フォーラム, 2017.9.6, 東京 (招待講演)
6. 原野幸治, 分子科学研究のための原子分解能顕微鏡動画撮影技術, AIKOC-1, 2017.8.5, 名古屋 (招待講演)
7. 原野幸治, 原子分解能電子顕微鏡で分子が動く, 集まる, 反応する様子をとらえる, 統合物質創製化学研究推進機構 第 1 回若手の会, 2017.7.28, 犬山 (招待講演)
8. Koji Harano, Functional Hybrid Vesicles Made of [60]Fullerene Amphiphiles Japan-China Joint Interdisciplinary Symposium on Coordination-based Hybrid Materials, 2017.6.24, Okazaki (Invited talk)
9. Koji Harano, The New Frontiers of Organic and Supramolecular Chemistry Explored by High-resolution Electron Microscopy, Department Lecture, 2017.5.19, Zurich (Invited talk)
10. Koji Harano, The New Frontiers of Organic and Supramolecular Chemistry Explored by High-resolution Electron Microscopy, Intergroup Seminar, 2017.5.17, Zurich (Invited talk)
11. 原野幸治, 有機分子の原子分解能顕微鏡動画を撮影する, 日本鉄鋼協会高温プロセス部会資源・エネルギーフォーラム, 2017.1.18, 東京 (招待講演)
12. 原野幸治, 高分解能電子顕微鏡で切り開くナノ・メゾ分子集合体科学, 立教大学特別講義, 2016.12.19, 東京 (招待講演)

13. 原野幸治, 結晶が生まれる様子が見える!分子の動画撮影, 第6回CSJ化学フェスタ, 2016.11.15, 東京
(招待講演)

【前田大光】

1. Hiromitsu Maeda, π -Electronic Ion-Pairing Materials That Exhibit Stimuli-Responsive Behaviors International Congress on Pure & Applied Chemistry (ICPAC) 2018, 2018.3.7, Cambodia (Invited talk)
2. 前田大光, イオンペアリング π 電子系集合体の創製, 早稲田大学大学院講演会, 2018.2.27, 東京(招待講演)
3. Hiromitsu Maeda, π -Electronic Ion-Pairing Materials That Exhibit Stimuli-Responsive Behaviors 分子科学研究所講演会, 2018.1.24, 岡崎 (Invited talk)
4. 前田大光, イオンペアリング π 電子系集合体の創製, 東京工業大学理学院化学系講演会(超分子分析化学セミナー), 2018.1.11, 東京 (招待講演)
5. 前田大光, イオン応答性 π 電子系からイオンペア集合体への新たな展開, 大須賀研講演会, 2017.12.28, 京都
(招待講演)
6. 前田大光, π 電子系集合体の新展開: ポルフィリンを基盤としたデザインと合成, 新潟大学講演会, 2017.12.15, 新潟 (招待講演)
7. 前田大光, イオン応答性 π 電子系の超分子化学: イオンペア集合体の創製と新展開, 静岡大学理学部化学科
講演会, 2017.11.22, 静岡 (招待講演)
8. 前田大光, イオン応答性分子の合成を契機とした π 電子系集合体の創製と新展開, 東京大学「工学とバイオ」
セミナー, 2017.11.17, 東京 (招待講演)
9. 前田大光, 超分子化学: 個々の分子を凌駕した物性・機能性の発現, 東京工科大学 サステイナブル応用化学
特別講義, 2017.11.6, 八王子 (招待講演)
10. Hiromitsu Maeda, Ion-Pairing Assemblies Exhibiting Stimuli-Responsive Behaviors, ストラスブール
大学講演会, 2017.10.26, France (Invited talk)
11. Hiromitsu Maeda, Anion-Responsive π -Electronic Molecules with Dynamic Conformation Changes
ストラスブール大学講演会, 2017.10.24, France (Invited talk)
12. Hiromitsu Maeda, Dimension-Controlled Ion-Pairing Assemblies Comprising Charged Metal Complexes
of π -Electronic Systems, 254th ACS National Meeting, 2017.8.20, USA (Invited talk)
13. Hiromitsu Maeda, π -Electronic Ion-Pairing Supramolecular Assemblies, International Symposium on
Pure & Applied Chemistry (ISPAC) 2017, 2017.6.8, Vietnam (Invited talk)
14. 前田大光, π 電子系集合体の新展開: 分子デザインと合成, 有機合成化学協会中四国支部 パネル討論会,
2017.5.20, 東広島 (招待講演)
15. Hiromitsu Maeda, Ion-Pairing Assemblies Exhibiting Photo-Responsive Crystal-Crystal Phase
Transitions, China-Japan Joint Symposium on Functional Supramolecular Architectures, 2017.5.14,
China, (Invited talk)
16. 前田大光, π 電子系イオンからなる次元制御型集合体の創製, 日本化学会第97春季年会 特別企画,
2017.3.19, 横浜 (招待講演)
17. 前田大光, イオン応答性 π 電子系の超分子化学, 機能性分子創製の最前線, 2017.3.15, 東京, (招待講演)
18. 前田大光, 超分子化学: 個々の分子を凌駕した物性・機能性の発現, 信州大学CSTプログラム 最新の科学情
報を学ぶ特別授業, 長野, 2017年2月17日 (招待講演)
19. 前田大光, イオン会合能を有する π 電子系の合成と超分子集合化
滋賀医科大学講演会, 大津, 2017年1月24日 (招待講演)
20. Hiromitsu Maeda, Supramolecular Assemblies Comprising π -Electronic and Photoresponsive Ions, 9th
Asian Photochemistry Conference (APC2016), 2016.12.4, シンガポール (Invited talk)
21. Hiromitsu Maeda, Supramolecular Assemblies Comprising π -Electronic Ions, 第2回超然プロジェクト
(金沢大学) シンポジウム, 2016.10.27, 金沢 (招待講演)
22. Hiromitsu Maeda, π -Electronic Ion-Pairing Supramolecular Assemblies, 産業技術総合研究所(関西セン
ター) 第2回RIECENセミナー, 2016.7.27, 池田
23. Hiromitsu Maeda, π -Electronic Ion-Pairing Supramolecular Assemblies, International Symposium on
Polymer and Related Materials, 2016.7.13, 長春, China (Keynote Lecture)
24. Hiromitsu Maeda, π -Electronic Ion-Pairing Supramolecular Assemblies, International Symposium on
Polymer and Related Materials, 2016.7.10, ハルビン, China (Invited talk)
25. Hiromitsu Maeda, π -Electronic Ion-Pairing Supramolecular Assemblies, 2016 International Workshop
on Nanomaterials and Nanodevices, 2016.7.8, 北, China (Keynote Lecture)
26. Hiromitsu Maeda, π -Electronic Ion-Pairing Dimension-Controlled Assemblies, The 8th Japanese-
Italian Liquid Crystal Workshop (JILCW2016), 2016.7.6, 京都 (Invited talk)

27. Yohei Haketa, Yuya Bando, Hiromitsu Maeda, Ion-Pairing Assemblies Based on “Genuine” π -Electronic Ions, 12th International Workshop on Supramolecular Nanoscience of Chemically Programmed Pigments, 2016.6.18, 草津(Invited talk)
28. Hiromitsu Maeda, Pyrrole-Based Ion-Pairing Supramolecular Assemblies, 229th meeting of the Electrochemical Society (ECS), 2016.5.30, San Diego, USA (Invited talk)
29. 前田大光, 光応答性イオンからなる次元制御型集合体の創製, 日本化学会第96春季年会 特別企画 2016.3.27, 京田辺 (招待講演)
30. Hiromitsu Maeda, Supramolecular ion-pairing assemblies based on anion-responsive π -systems 251st ACS National Meeting, 2016.3.16, サンディエゴ(Invited talk)
31. Hiromitsu Maeda, Supramolecular Assemblies Comprising Ionic π -Electronic Systems, Japan-China Joint Symposium on Functional Supramolecular Architectures, 2016.2.25, 岡崎(Invited talk)
32. 前田大光, π 電子系イオンを基盤とした次元制御型集合体の創製, 2015年度日本接着学会中部支部接着マスター講座, 2016.1.29, 名古屋 (招待講演)
33. Hiromitsu Maeda, Supramolecular Ion-Pairing Assemblies Based on Anion-Responsive π -Systems MATCON 2016 - International Conference on Materials for the Millennium, 2016.1.15, コーチ (Keynote lecture)
34. Hiromitsu Maeda, Ion-pairing assemblies based on anion-responsive π -electronic molecules Pacifichem 2015, 2015.12.20, ホノルル(Invited talk)
35. Hiromitsu Maeda, π -Electronic Ion-Pairing Supramolecular Assemblies, Japan-Euro Joint Workshop for Photo- and Electro-Molecular Mechanics, 2015.10.6, トゥールーズ, (Invited talk)
36. 前田大光, イオン応答性 π 電子系を基盤とした次元制御型集合体の創製, 第4回液晶若手シンポジウム, 2015.9.10, 東京 (招待講演)
37. Hiromitsu Maeda, π -Electronic Ion-Pairing Supramolecular Assemblies 分子研講演会, 2015.7.29, 岡崎(Invited talk)
38. Ryohei Yamakado, Hiromitsu Maeda, π -Electronic Systems That Provide Various Anion Complexes and Resulting Ion-Pairing Assemblies, 11th International Workshop on Supramolecular Nanoscience of Chemically Programmed Pigments, 2015.5.30, 草津(Invited talk)
39. Hiromitsu Maeda, Ion-Pairing Assemblies Comprising Pyrrole-Based π -Electronic Systems 227th meeting of the Electrochemical Society (ECS), 2015.5.24, シカゴ(Invited talk)
40. 前田大光, 「 π 電子系イオンを基盤とした超分子集合体の創製」, 日本化学会第95春季年会 特別企画, 2015.3.29, 船橋 (招待講演)
41. Hiromitsu Maeda, “Supramolecular Assemblies Comprising Ionic π -Electronic Systems” ANGEL Kick-Off Meeting 2015, 2015.1.28-29, 米沢 (Invited talk)
42. Hiromitsu Maeda, “Ionic and Ion-Responsive π -Electronic Systems and Their Supramolecular Assemblies”, 分子科学研究所講演会, 2015.1.20, 岡崎(Invited talk)
43. Hiromitsu Maeda, “Supramolecular Assemblies Comprising Ionic π -Electronic Systems”, China-Japan Joint Symposium on Functional Supramolecular Architectures, 2014.12.10-13, 天津, 中国 (Invited talk)
44. 前田大光, 「 π 電子系イオンを基盤とした超分子集合体の創製」, お茶の水女子大学理学部化学科講演会, 2014.12.3, 東京 (招待講演)
45. 前田大光, 「 π 電子系を構成ユニットとした超分子集合体の創製」, 東京理科大学理学部応用化学科講演会（「分子集積化学」非常勤講義）, 2014.11.11, 東京 (招待講演)
46. 前田大光, 「発光性 π 電子系におけるキラリティ誘起の新戦略」, 第4回CSJ化学フェスタ2014, 2014.10.16, 東京
47. 前田大光, 「 π 電子系イオンの次元制御型集合化」, 第8回超分子若手懇談会, 2014.10.9-10, 加賀 (招待講演)
48. Hiromitsu Maeda, “Assemblies of π -Conjugated Ionic Species Providing Electronic Materials”, International Union of Materials Research Societies, International Conference in Asia (IUMRS-ICA 2014) 2014, 2014.8.24-30, 福岡 (招待講演)
49. Hiromitsu Maeda, “Supramolecular Assemblies Comprising Anion-Binding and Metal-Coordinating π -Conjugated Ionic Building Units”, 248th ACS National Meeting, 2014.8.10-14, サンフランシスコ (Invited talk)
50. 前田大光, 「イオン応答性 π 電子系を基盤とした超分子集合体の創製」, 機能超分子セミナー, 2014.7.23, 東京 (招待講演)

51. Hiromitsu Maeda, "Ion-Based Assemblies of Pyrrole-Based π-Systems", Eighth International Conference on Porphyrins and Phthalocyanines (ICPP-8) ,2014.6.22-27, イスタンブル (Invited talk)
52. 前田大光, 「イオン応答性 π 電子系を基盤とした超分子集合体の創製」, 第 12 回ホスト・ゲスト化学シンポジウム, 2014.5.31-6.1, 東京 (招待講演)

(3) 図書

A01 : 動的秩序の探査

【秋山修志】

1. 秋山 修志, 「概日時計因子の構造や動態を調べる意義とは?」, 生物物理 **56**, 266-270, (2016)
- 【秋山良】

1. R. Akiyama, "Theoretical Studies of Strong Attractive Interaction Between Macro-anions Mediated by Multivalent Metal Cations and Related Association Behavior: Effective Interaction Between ATP-Binding Proteins Can Be Regulated by Hydrolysis", The Role of Water in ATP Hydrolysis Energy Transduction by Protein Machinery, Makoto Suzuki ed. Springer (Japan), pp 53-67, 10.1007/978-981-10-8459-1_4, (2018)
2. 安池智一, 秋山良, “エントロピーからはじめる熱力学”, 放送大学教育振興会, p.p.1-300, ISBN978-4-595-31643, 2016.3.20

【池谷鉄兵】

1. T. Ikeya, *Y. Ito, "Advances in NMR Data Acquisition and Processing for Protein Structure Determination, Experimental Approaches of NMR Spectroscopy -Methodology and Application to Life Science and Materials Science", Springer, 63-99, (2018)
2. 葛西卓磨, 池谷鉄兵, 木川隆則, “生命分子のNMR計測・解析への応用”, 電子情報通信学会誌, Vol.99, pp439-443, (2016)

【岩田耕一】

1. 岩田耕一, 「ラマン分光法」, 日本光生物学協会 光と生命の事典 編集委員会 編, 朝倉書店), ISBN978-4-254-17161-7. 「光検出器」, pp1-224 (2016)
2. Hiro-o Hamatuchi and Koichi Iwata, "Time-resolved Raman Spectroscopy", Encyclopedia of Spectroscopy and Spectrometry, 3rd Edition, Elsevier, 分担執筆, in press, (2016)
3. 濱口宏夫, 岩田耕一 共編, 「ラマン分光法」, 日本分光学会分光法シリーズ 1, 講談社, ISBN978-4-06-156901-0. 第 3 章「ラマン分光の実際」(51 から 87 ページ) および付録 A から付録 C (145 から 180 ページ) を分担執筆, (2015)
4. 岩田耕一, “光化学の事典” 3-2 節「さまざまな光化学反応」③「結合解離反応」, 光化学の事典編集委員会 編, 朝倉書店, 東京 (分担執筆), pp1-418 (2014)

【内橋貴之】

1. T. Uchihashi, H. Watanabe and N. Kodera, "Optimum Substrates for Imaging Biological Molecules with High-speed Atomic Force Microscopy", Methods Mol. Biol., 1814, 15-179, (2108)
2. T. Uchihashi, "High-Speed Atomic Force Microscopy", Compendium of Surface and Interface Analysis (The Surface Science Society of Japan, Eds) (Springer), pp. 263-267, 10.1007/978-981-10-6156-1, (2018)
3. 内橋貴之, “光と生命の辞典” 第 5 章 「光による生命現象の計測」 177 節 高速原子間力顕微鏡, (20016), 真嶋哲郎, 七田芳則, 飯野盛利, 藤堂剛 編, 朝倉書店, 東京, ISBN: 978-254-17161-7 C3545, pp1-436 (2015)
4. T. Uchihahshi, N. Kodera, and T. Ando (章著), “Development of High-speed AFM and its Biological Applications”, K. Takeyasu ed., Atomic Force Microscopy in Nanobiology, Chap. 8, pp. 143-176. Pan Stanford Publishing, 10.4032/9789814411592, (2014)

【上久保裕生】

1. 上久保裕生, 光による生命現象の計測 (X 線小角散乱), 第 5 章, 「光と生命の事典」, 編集: 真嶋哲郎・飯野盛利・七田芳則・藤堂剛, 朝倉書店, 東京, pp1-436 (2016)

【佐藤啓文】

1. 佐藤啓文, “揺らぎ・ダイナミクスと生体機能: 物理化学的視点から見た生体分子” (DOJIN BIOSCIENCE SERIES) 第 7 章「理論」2 節「分子性液体の積分方程式理論」, 寺嶋正秀編, 化学同人, 京都, ISBN:9784759815108, pp103-135 (2013)

【高田十志和】

- 青木大輔, 高田十志和, “高分子鎖の可動な連結を利用するトポロジー変換と特性制御”, 高分子, 65(5), pp 217-219, (2017)
- 青木大輔, 高田十志和, “高分子鎖の機械的な連結による分岐-線状トポロジー変換-新しい刺激応答高分子の創製”, 日本ゴム協会誌, 90(6), pp 283-289, (2017)
- 高田十志和, 曾川洋光, 新版ゲルテクノロジーハンドブック, (架橋点可動型), エヌティーエス出版, 東京, pp1-908 (2014) ISBN 978-4-86469-074-4

【立川仁典】

- 飯沼裕美, 大場優生, 河村成肇, 高妻孝光, 菅原洋子, 高柳敏幸, 立川仁典, 「新しい量子ビーム・ミュオン分光と理論的アプローチ」 J. Comput. Chem. Jpn., vol. 16, A12-A17, (in Japanese), (2017)
- 川島雪生, 澤田啓介, 中嶋隆人, 立川仁典, 酢酸-リリン酸アニオンクラスターの分子間水素結合における核の量子揺らぎの効果に関する理論的研究 J. Comput. Chem. Jpn. 「量子水素の科学」特集号, vol. 15, 203-209, (in Japanese), (2016)
- 木下郁雄, 北幸海, 立川仁典, 橘勝, 「カーボンナノウォールの電子状態と水素吸着」 J. Comput. Chem. Jpn. 「量子水素の科学」特集号, vol. 15, 177-183, (in Japanese), (2016)
- 宇田川太郎, 常田貴夫, 立川仁典, 「多成分密度汎関数のための電子-核相関汎関数の開発」 J. Comput. Chem. Jpn. 「量子水素の科学」特集号, vol. 15, 143-147, (in Japanese), (2016)
- 立川仁典, 「特集「量子水素の科学」に寄せて」 J. Comput. Chem. Jpn. 「量子水素の科学」特集号, vol. 15, A51, (in Japanese), (2016)
- 立川仁典, 北幸海, 小山田隆行, 「原子・分子の陽電子束縛機構と対消滅機構解明のための高精度第一原理計算 (Accurate ab initio calculation for the elucidation of the mechanism of positron binding and pair-annihilation in atoms and molecules)」 陽電子科学, vol. 7, 1-11, (2016),

【田中良和】

- 加藤早苗, 松井崇, *田中良和, “3.8MDa の超巨大酸素運搬蛋白質ヘモシアニン会合体の結晶構造,” 生化学 Vol.90, 238-243, 10.14952/SEIKAGAKU.2018.900238, (2018)
- 田中良和 Essential タンパク質科学 第2章 タンパク質のドメイン, p59-94 南江堂, (2016)
- Yamashita, D., Sugawara, T., Tanaka, I., Tanaka, Y., and Yao, M., Pore formation mechanism of staphylococcal pore forming toxin, PF activity report part A, 2014 Highlight, 46-47, (2015)
- 田中良和, 陳明皓, 姚閔, tRNA ジヒドロウリジン合成酵素の分子機構 生化学 86 (3), 395-399, (2014)
- 田中良和 環境と微生物の事典 第6章 p135 人体環境での鉄の獲得 朝倉書店, (2014)

【寺嶋正秀】

- *寺嶋正秀, 光熱変換現象の時間分解検出と化学反応解析への応用, 分光研究, 67, 22-34, (2018)
- 黒井邦巧, 寺嶋正秀, 「圧力印加過渡回折格子法による反応中のタンパク質圧縮率の時間分解計測」, 熱測定, 43, 66-71, (2016)
- 寺嶋正秀, 「分子基盤に基づく生体機能ネットワークとダイナミクス」, 国際高等研究所アニュアルレポート 2015, (2016)
- 中曾根祐介, 寺嶋正秀, 青色光センサータンパク質フォトトロピンの光制御機構」, 物性科学センター誌, 29, 3-11, (2016)
- 寺嶋正秀 (分担執筆), 「過渡回折格子法」, 光と生命の事典, pp356-357, 2016, 朝倉書店 ISBN978-4-254-17161-7, (2016)
- Masahide Terazima, “Time-Resolved Detection of Protein Fluctuations During Reactions”, Molecular Science of Fluctuations Toward Biological Functions, Eds., M.Terazima, M.Kataoka, R.Ueoka, 1-28, Springer. ISBN 978-4-431-55840-8, 分担執筆, (2016)
- 寺嶋正秀, 時間分解熱力学量で見るタンパク質揺らぎと反応, パリティ, Vol.30, No.08, 52-55, (2015)
- 黒井邦巧, 寺嶋正秀, タンパク質反応と揺らぎ, 生物物理学会誌, 55, 235-241, (2015)
- 寺嶋正秀, タンパク質の揺らぎが生体反応を決める, 寺嶋正秀, 現代化学, 534, 42-46, (2015)
- 寺嶋正秀, 馬場正昭, 松本吉泰 共著, 現代物理化学, 化学同人 ISBN978-4-7598-1809-3, (2015)
- 寺嶋正秀 (分担執筆), 蛍光蛋白質, 「光化学の事典」, 光化学協会光化学の事典編集委員会 編, 朝倉書店, ISBN978-4-254-14096-5, C3543, (2014)
- 寺嶋正秀, “揺らぎ・ダイナミクスと生体機能: 物理化学的視点から見た生体分子” (DOJIN BIOSCIENCE SERIES) 第1章「揺らぎと生体反応概論」, 寺嶋正秀編, 化学同人, 京都, ISBN:9784759815108, pp3-11 (2013)
- 寺嶋正秀, “揺らぎ・ダイナミクスと生体機能: 物理化学的視点から見た生体分子” (DOJIN BIOSCIENCE SERIES) 第2章「分光法」, 寺嶋正秀編, 化学同人, 京都, ISBN:9784759815108, pp12-27 (2013)

14. 寺嶋正秀, “揺らぎ・ダイナミクスと生体機能: 物理化学的視点から見た生体分子” (DOJIN BIOSCIENCE SERIES)第 10 章「新しい時間分解測定: 拡散係数と熱力学量」, 寺嶋正秀編, 化学同人, 京都, ISBN:9784759815108, pp159-169 (2013)
15. 寺嶋正秀, 揺らぎから観た生体分子科学, Molecular Science 誌, 第 7 卷, A0063 (1-10 ページ) (2013)
【内藤晶】
 1. 功刀滋, 内藤 晶, “生命科学のための物理化学 15 講,” 講談社サイエンティフィク 編集, pp 1-244, ISBN978-4-06-153898-6, (2018)
 2. *I. Kawamura, K. Norisada, A. Naito, “Structure Determination of Membrane Peptides and Proteins by Solid-state NMR”, Experimental Approaches of NMR Spectroscopy. Methodology and Application of Life Science and Materials Science, The NMR Society of Japan. Ed. Springer, pp 135- 170, DOI: [10.1007/978-981-10-5966-7_9](https://doi.org/10.1007/978-981-10-5966-7_9), (2018)
 3. *A. Naito, Y. Makino, Y. Tasei, I. Kawamura, “Photoirradiation and Microwave Irradiation NMR Spectroscopy,” Experimental Approaches of NMR Spectroscopy –Methodology and Application of Life Science and Material Science-. The NMR Society of Japan ed. Springer, pp 135-170, [10.1007/978-981-10-5966-7_5](https://doi.org/10.1007/978-981-10-5966-7_5), (2018)
 4. *A. Naito, Y. Makino, I. Kawamura, “In-situ Photo Irradiation Solid-state NMR Spectroscopy Applied to Retinal-binding Membrane Protein,” Mod. Magn. Reson., pp.537-557, Webb ed. Springer, 1-22, [10.1007/978-3-319-28275-6_51-1](https://doi.org/10.1007/978-3-319-28275-6_51-1), (2017)
 5. *内藤晶, 固体高分解能 NMR(Narrow is beautiful)の構造生命化学研究への展開 NMR (Bulletin of the nuclear magnetic society of japan) 日本核磁気共鳴学会 Vol. 7, pp 8-10, (2016)
 6. 内藤晶, “光と生命の事典”第 5 章「光による生命現象の計測」, 第 176 節「NMR 分光法」, 日本光生物学協会編, 朝倉書店, ISBN: 978-4-254-17161-7 c3545, pp1-436 (2016)
 7. A. Naito, I. Kawamura, N. Javkhlanlugs, “Recent solid-state NMR studies of membrane-bound peptides and proteins.” Annu. Rep. NMR Spectrosc., Graham A. Webb, ed. Academic Press Vol 86, pp 333-411, [10.1016/bs.arnmr.2015.06.001](https://doi.org/10.1016/bs.arnmr.2015.06.001), (2015)
 8. *A. Naito, I. Kawamura, “Photoactivated Structural Changes in Photoreceptor Membrane Proteins as Revealed by in situ Photoirradiation Solid-State NMR Spectroscopy”n Chater 20 in “Advances in Biological Solid-State NMR”, Eds. F. Separovic, A. Naito, (Royal Society of Chemistry, UK), ISBN: 978-1-84973-910-8, pp387-404 (2014)
 9. 内藤晶, ”最新マイクロ波エネルギーと応用技術”第 1 章「マイクロ波の基礎」, 第 3 節「電磁場相互作用の NMR による研究」, 吉川昇 編, 産業技術サービスセンター, 東京, ISBN:978-4-915957-94-9 C3053, pp 1-960 (2014)

【松森信明】

1. 松森信明, 村田道雄, 「エリスロマイシンと膜脂質」, 天然物の化学 II (科学のとびら 64), 上村大輔編, pp162-168, 東京化学同人, ISBN: 9784807915057, (2018)
2. N. Matsumori, *M. Murata, “NMR Studies on Natural Product—Stereochemical Determination and Conformational Analysis in Solution and in Membrane”, Experimental Approaches of NMR Spectroscopy, The Nuclear Magnetic Resonance Society of Japan (eds) pp 383-414 Springer Singapore, https://doi.org/10.1007/978-981-10-5966-7_14, (2018)

【養王田正文】

1. M. Yohda, “Aspartate Racemase: Function, Structure, and Reaction Mechanism”, D-Amino Acids :Physiology, Metabolism, and Application, T. Yoshimura, T. Nishikawa and H. Homma ed. Springer, Japan, pp 323-337, [10.1007/978-4-431-56077-7_21](https://doi.org/10.1007/978-4-431-56077-7_21), (2016)

A02 : 動的秩序の創生

【飯野亮太】

1. *A. Nakamura, R. Iino, “Visualization of functional structure and kinetic dynamics of cellulases Glycobiophysics”, K.Kato, Y. Yamaguchi, editors, Springer,in press, (2018)
2. *R. Iino, S. Sakakihara, Y. Matsumoto, K. Nishino, “Large Scale Femtoliter Droplet Array for Single Cell Efflux Assay of Bacteria”, Methods in Molecular Biology, A. Yamaguchi ed., Springer, 1700, 331-341, DOI: [10.1007/978-1-4939-7454-2_18](https://doi.org/10.1007/978-1-4939-7454-2_18), (2018)
3. *R. Iino, S. Sakakihara, Y. Matsumoto, K. Nishino, “Large Scale Femtoliter Droplet Array for Single Cell Efflux Assay of Bacteria”, Methods in Molecular Biology, A. Yamaguchi ed., Springer, in press, (2017)
4. *R. Iino, S. Sakakihara, Y. Matsumoto, K. Nishino, “Single-cell Detection and Collection of Persister

- Bacteria in a Directly Accessible Femtoliter Droplet Array”, Methods in Molecular Biology, 1333, 101-109, 10.1007/978-1-4939-2854-5_9, (2015)
5. *飯野亮太, “生体分子機械の作動原理”, 自己組織化マテリアルのフロンティア, p67-74, フロンティア出版, (2015)
 6. 飯野亮太, “薬剤排出トランスポーター活性のマイクロデバイスによる計測”, 化学療法の領域, 31, 440-448, (2015)
 7. R. Iino, S. Sakakihara, Y. Matsumoto, K. Nishino, “Single-cell detection and collection of persister bacteria in a directly accessible femtoliter droplet array”, Methods in Molecular Biology, in press , (2015)
 8. 飯野亮太, “化学フロンティア 23 1 分子ナノバイオ計測：分子から生命システムを探る革新的技術”, III 部 Topics 5 “デジタル PCR とデジタル ELISA”, 野地博行編, 化学同人, 京都, 1-240 (2014)
 9. 内橋貴之, 飯野亮太, 安藤敏夫, 野地博行, 高速 AFM による F1-ATPase 分子回転の直接可視化, 生化学. 86, 127-136, 日本生化学会, 東京, pp1-240 (2014)
 10. 飯野亮太, 中村彰彦, 五十嵐圭日子, 鮫島正浩“, 1 分子計測からわかるエクソ型セルラーゼの分子機構”, 生物物理. 54, 318-320, 日本生物物理学会, 大阪, (2014)

【上野隆文】

1. 稲葉央, 安部聰, *上野隆史, 「超分子タンパク質を用いて金属の反応を操る」, 化学, 70, 41-46, (2015)
2. Takafumi Ueno (Guest Editor), Special Issue: Artificial Metalloenzymes, Isr. J. Chem., 55, DOI: 10.1002/ijch.201410018, (2015)
3. 藤田健太, 上野隆史, 「細胞への一酸化炭素ガス分子放出を指向した蛋白質集合体の機能化」酵素工学ニュース, 73, 14-16, (2015)
4. 稲葉央, 上野隆史, 「タンパク質分子針の動的機能と細胞制御」, 生物物理, 55, 89-91, (2015)
5. 安部聰, 上野隆史, 「タンパク質結晶の分子設計によるバイオ固体材料の開発」, 化学工業, 66, 264-272, (2015)
6. 藤田健太, 上野隆史, 展望「X線結晶構造解析が解き明かす一カゴタンパク質のガス放出」日本アイソトープ協会"ISOTOPE NEWS", 64, 2-6, (2015)

【片山勉】

1. 神崎秀嗣, 片山勉, 児玉 悟, 新藏礼子, 矢島孝浩, “「微生物と人類の共存」をゲノムの視点から ---「日本遺伝学会第 88 会大会」の 1 ワークショップ報告”, 生物の科学 遺伝, Vol. 71, No. 1, pp77-85, (2017)
2. *T. Katayama, “Initiation of DNA Replication at the Chromosomal Origin of E. coli, oriC”, DNA Replication: From Old Principles to New Discovery, H. Masai and M. Foiani ed. SpringerNature, , (Singapore), 79-98, 10.1007/978-981-10-6955-0, (2017)

【神谷由紀子】

1. 神谷由紀子, 村山恵司, 横田 啓, 浅沼浩之, “非環状骨格型人工核酸 : aTNA, SNA”, “核酸医薬の創製と応用展開”, 監修: 和田 猛, p79-86, シーエムシー出版, (2016)
2. Y. Zhang, T. Yamaguchi, M. Yagi-Utsumi, Y. Kamiya, Y. Sakae, Y. Okamoto, *K. Kato, “Conformational Dynamics of Oligosaccharides Characterized by Paramagnetism-assisted NMR Spectroscopy in Conjunction with Molecular Dynamics Simulation”, Advances in Experimental Medicine and Biology , A.Chakrabarti and A.Surolia ed., Springer, Switzerland, Vol. 842, pp 389-401, 10.1007/978-3-319-11280-0_14, (2015)

【佐田和己】

1. K. Sada, T. Ishiwata, K. Kokado, “Advances in Organic Crystal Chemistry: Comprehensive Reviews 2015”, “Topochemical Polymerizations & Crystal Cross-linking of Metal Organic Frameworks”, in press, Rui Tamura, Mikiji Miyata, eds., Springer, Germany, (2015)
2. K. Sada, Y. Furukawa, K. Kokado, “Encyclopedia of Polymeric Nanomaterials” “Polyacrylonitrile”, in press, Shiro Kobayashi, Klaus Müllen, eds., Springer, Germany, (2015)
3. 小門憲太, 佐田和己, “クリックケミストリー —基礎から実用まで—”, 第 17 章「多孔性配位高分子のクリックケミストリー」, pp.161-173, 高田十志和, 小山靖人, 深瀬浩一編, シーエムシー出版, 東京, (2014)
4. S. Amemori, K. Kokado, K. Sada, “Synergy in Supramolecular Chemistry”, Chapter 5, “Chemo-sensitive Soft Matters Based on Thermo-sensitive Polymers”, in press, p.75-, T. Nabeshima, ed., Taylor & Francis, USA, (2014)
5. 小門憲太, 佐田和己, “ゲルテクノロジーハンドブック —機能設計・評価。シミュレーションから製造プロセスまで—” 1-3-5 「有機溶媒高吸収性親油性高分子電解質ゲル」, pp.97-103, 中野義夫他編, エヌ・ティー・エス出版, 東京, (2014)

6. 上西恭平,井上大介,佐田和己,角五彰,“ゲルテクノロジーハンドブック —機能設計・評価。シミュレーションから製造プロセスまで—” 1-5-2 「ATP 駆動型運動素子」, pp.167-172, 中野義夫他編, エヌ・ティー・エス出版, 東京, (2014)
7. S. Amemori, K. Kokado, K. Sada, “Chemo-sensitive Soft Matters Based on Thermo-sensitive Polymers”, Synergy in Supramolecular Chemistry, T. Nabeshima, ed. Taylor & Francis, USA, Chapter 5, pp. 75-90, (2014)

【佐藤宗太】

1. T. Matsuno, S. Sato, H. Isobe, “Curved π-Receptors”, Comprehensive Supramolecular Chemistry II, vol. 3, p. 311-328, Elsevier, (2017)
2. 池本晃喜, 佐藤宗太, 磯部寛之, 「芳香環で「笊」を編む –ナノメートルサイズのボウル分子のボトムアップ合成–」, 化学, vol. 72, No. 7, p. 31-35, (2017)
3. 磯部寛之, 佐藤宗太, ”炭素と水素の構造化学：ありふれた元素・分子からつくる機能性 材料”, パリティ (クローズアップ), vol. 32, No. 3, p. 39-42, (2017)
4. 磯部寛之, 佐藤宗太, ”「防虫剤」からリチウムイオン 2 次電池負電極”, パリティ (特集 物理科学, この 1 年) , vol. 32, No. 1, p. 35-36, (2017)
5. 佐藤宗太, ”サイボーグ超分子により解き明かされる生命現象～生体分子クラスターを人工巨大分子に移植した一義構造の巨大分子”, 分子研レターズ 73, pp32-33 (2016)
6. 佐藤宗太, 加藤晃一, 藤田誠, “生命現象の解明に挑むサイボーグ超分子 – 機能を維持したまま生体分子クラスターを人工分子に移植 –”, 月刊化学, vol. 70, No. 11, p. 31-36, (2015)
7. 佐藤宗太, “巨大な中空球状錯体を骨格として構築した生体分子インターフェースの開発”, Bull. Jpn. Soc. Coord. Chem., 65, 30-37, (Award Accounts), (2015)
8. 藤田大士, 佐藤宗太, 藤田 誠, “長期利用課題報告 放射光 X 線を用いた多成分からなる自己集合性錯体の単結晶構造解析”, SPring-8/SACLA 利用者情報 「最近の研究から」欄, 20, 130-133, (2015)
9. 佐藤宗太, 磯部寛之, “カーボンナノチューブの筒内平滑曲面：炭素性分子ベアリングの構造化学”, 日本結晶学会誌 「最近の研究から」欄, 56, 405-410, (2015)

【鈴木大介】

1. *Daisuke Suzuki, Takuma Kureha and Koji Horigome, “Hydrogel Microspheres”, Encyclopedia of Biocolloid and Biointerface Sciences (Ohshima Ed.), Wiley InterScience, ISBN:9781118485590, pp554-569 (2015)
2. 鈴木大介, 「ゲル微粒子の自己組織化」, 新版ゲルテクノロジーハンドブック, エヌ・ティー・エス出版, 東京, pp186-193 (2014)

【平岡秀一】

1. 平岡秀一, “水の不思議と自己組織化”, 現代化学 No. 555, (6), 34–36, (2017)
2. 平岡秀一, “溶液における分子認識と自己集合の原理：分子間相互作用”, サイエンス社, ISBN 978-4-7819-1403-9, pp1-232 (2017)

【二木史朗】

1. I. Nakase, T. Takeuchi, S. Futaki, Cell Penetrating Peptides for Chemical Biological Studies, In Cell-penetrating Peptides: Methods and Protocols, Ed. by Ülo Langel, Humana Press, Springer, New York, 387-396, (2015)
2. 二木史朗, “膜透過ペプチドを利用する細胞内デリバリー”, 遺伝子医学 MOOK 別冊次世代ペプチド医薬創製 第 2 章 1 節, pp 68-72, 赤路健一編, メディカルドウ, 大阪, (2014)

【芳坂貴弘】

1. 芳坂貴弘, “非天然アミノ酸の導入”, 人工細胞の創製とその応用 (植田充美監修) 1-5 章, シーエムシー出版, pp1-215, ISBN 978-4-7813-1233-0, (2017)
2. 芳坂貴弘, “揺らぎ・ダイナミクスと生体機能: 物理化学的視点から見た生体分子” (DOJIN BIOSCIENCE SERIES) 第 6 章 「変異導入法」, 寺嶋正秀編, 化学同人, 京都, ISBN:9784759815108, (総ページ数 347) , pp91-102 (2013)

【松浦友亮】

1. 松浦友亮, セルフリータンパク質合成系を用いた進化分子工学技術の開発, 日本生物工学会誌, 95, 121-126, (2017)
2. 松浦友亮, ネオバイオ分子創生を目指した配列空間探索, 日本生物工学会誌, 94, 471-472, (2016)

【三宅弘之】

1. 三宅弘之, “CoN₂O₄ 錯体”, “CuN₃OX/CuN₃S 錯体”, AgN₃ 錯体”, LnN₆O₃ 錯体”, “NdN₄O₄ 錯体”, 錯体化合物事典 (大川尚士 他編), 朝倉書店, (2018)

2. *K. Singh, P. Kaur, *H. Miyake, H. Tsukube, "Supramolecular Chemistry Strategies for Naked-eye Detection and Sensing", Synergy in Supramolecular Chemistry, (ed) T. Nabeshima, CRC Press, USA, ISBN 9781466595026, (2014)

A03 : 動的秩序の展開

【内山進】

1. E. Krayukhina, S. Uchiyama, "Analytical Ultracentrifugation", Advanced Methods in Structural Biology, T. Senda and K. Maenaka ed. Springer, Japan, 165-183, 10.1007/978-4-431-56030-2, (2016)
2. M. Noda, K. Fukui, S. Uchiyama, "Mass Spectrometry", Advanced Methods in Structural Biology, T. Senda, K. Maenaka, ed., Springer, Japan, 185-198, 10.1007/978-4-431-56030-2, (2016)
3. S. Uchiyama, F. Arisaka, "Important and Essential Theoretical Aspects of AUC", Analytical Ultracentrifugation, S. Uchiyama, F. Arisaka, W. F. Stafford, T. Laue ed. Springer, Japan, 3-14, 10.1007/978-4-431-55985-6_1, (2016)
4. F. Arisaka, S. Uchiyama, "Experimental Design and Practical Aspect", Analytical Ultracentrifugation, S. Uchiyama, F. Arisaka, W. F. Stafford, T. Laue ed. Springer, Japan, 15-21, 10.1007/978-4-431-55985-6_2, (2016)
5. S. Saito, S. Uchiyama, "Biopharmaceutical Evaluation of Intermolecular Interactions by AUC-SE", Analytical Ultracentrifugation, S. Uchiyama, F. Arisaka, W. F. Stafford, T. Laue ed. Springer, Japan, 419-440, 10.1007/978-4-431-55985-6_21, (2016)

【老木成稔】

1. *Sigetoshi Oiki, Dynamics of the Potassium Channel Pore. Reference Module in Life Sciences, Protein Biophysics, Ion Channels and Transporter Proteins, Elsevier, pp1-43 (2017)
<http://www.sciencedirect.com/science/article/pii/B9780128096338080924>
2. 老木成稔 化学フロンティア 23 “1分子ナノバイオ計測～分子から生命システムを探る革新的技術”, 第 5 章「イオン透過装置：イオンチャネル」, 69-80, 野地博行編, 化学同人, 京都, (2014)

【岡本祐幸】

1. M. Terazima, M. Kataoka, R. Ueoka, Y. Okamoto (eds.), "Molecular Science of Fluctuations toward Biological Functions", 270 pages, 全体を編集すると共に, 以下の第 9 章 (pp. 183-204) を著す, Structural fluctuations of proteins in folding and ligand docking studied by generalized-ensemble Simulations, (Y. Okamoto), DOI: 10.1007/978-4-431-55840-8, Springer, Tokyo, (2016)
2. Y. Zhang, T. Yamaguchi, T. Satoh, M. Yagi-Utsumi, Y. Kamiya, Y. Sakae, Y. Okamoto, K. Kato, "Conformational dynamics of oligosaccharides characterized by paramagnetism-assisted NMR spectroscopy in conjunction with molecular dynamics simulation", in Advances in Experimental Medicine and Biology 842: Biochemical Roles of Eukaryotic Cell Surface Macromolecules, A. Chakrabarti and A. Surolia.(ed.) pp. 217-230, DOI: 10.1007/978-3-319-11280-0_14, Springer, Heidelberg, (2015)
3. Y. Sakae and Y. Okamoto, "Optimizations of Protein Force Fields", in Computational Methods to Study the Structure and Dynamics of Biomolecules and Biomolecular Processes – from Bioinformatics to Molecular Quantum Mechanics, A.Liwo (ed.) (Springer-Verlag, 2014) pp. 195-247, DOI: 10.1007/978-3-642-28554-7_7, Springer-Verlag, (2014)
4. T. Yoda, Y. Sugita, and Y. Okamoto, "Protein Folding Simulations by Generalized-ensemble Algorithms", in Protein Conformational Dynamics, Advances in Experimental Medicine and Biology, Vol. 805, K.-L. Han, Xin Zhang, and M.-J. Yang (eds.) , pp. 1-27, Springer, (2014)
5. H. Kokubo, T. Tanaka, and Y. Okamoto, "Ligand Docking Simulations by Generalized-ensemble Algorithms", in Advances in Protein Chemistry and Structural Biology, Vol. 92, T. Karabencheva-Christova (ed.), pp. 63-91, DOI: 10.1016/B978-0-12-411636-8.00002-X, Academic Press, (2013)

【奥村久士】

1. 「In Silico 創薬」, 第 4 章第 1 節「高速計算プログラム GEMB による分子動力学シミュレーションの実際」, 技術情報協会 pp. 317-327, ISBN: 978-4-86104-688-9, (2018)

【加藤晃一】

1. K. Kato, H. Yagi, *Y. Yamaguchi, "NMR Characterization of the Dynamic Conformations of Oligosaccharides", Modern Magnetic Resonance, 2nd Edition, (G.A. Webb ed.), Springer International Publishing, in press, 10.1007/978-3-319-28275-6_35-1, (2018)
2. *K. Kato, S. Yanaka, H. Yagi, "Technical basis for nuclear magnetic resonance approach for glycoproteins", Experimental Approaches of NMR Spectroscopy (The Nuclear Magnetic Resonance Society of Japan ed.), Springer International Publishing, 415-438, (2018)

3. Y. Yamaguchi, H. Yagi, and K. Kato, "Stable isotope labeling of glycoproteins for NMR study," NMR in Glycoscience and Glycotechnology , K.Kato and T.Peters ed., RSC Publishing, Cambridge, p194-207, (2017)
4. M.Yagi-Utsumi, T.Yamaguchi, Y.Uekusa, and K.Kato, "NMR characterization of the conformations, dynamics, and interactions of glycosphingolipids," NMR in Glycoscience and Glycotechnology, RSC Publishing, Cambridge, pp161-178, 10.1039/9781782623946-00161, (2017)
5. 粂原顕輔,“分子が関わる人工細胞から生命を考える”季刊 生命誌 88, (2016)
6. T. Itoh, L. Sheng, *K. Kurihara, “Life Emerged from Oil”, Atlas of Sci. October 28, pp1-3 (2016)
7. G. Mandal, H. Yagi, K. Kato, and B. P. Chatterjee, “Structural heterogeneity of glycoform of alpha-1 acid glycoprotein in alcoholic cirrhosis patient”, Advances in Experimental Medicine and Biology (A.Chakrabarti and A.Surolia ed.), Springer, Switzerland, 842, 389-401, (2015)
8. Y. Zhang, T. Yamaguchi, T. Satoh, M. Yagi-Utsumi, Y. Kamiya, Y. Sakae, Y. Okamoto, *K. Kato, “Conformational dynamics of oligosaccharides characterized by paramagnetism-assisted NMR spectroscopy in conjunction with molecular dynamics simulation” Biochemical Roles of Eukaryotic Cell Surface Macromolecules, Advances in Experimental Medicine and Biology, A. Chakrabarti and A. Surolia ed. Springer, Volume 842, 217-230, 10.1007/978-3-319-11280-0, pp14, (2015)
9. T. Yamaguchi, K. Kato, “Paramagnetism-assisted nuclear magnetic resonance analysis of dynamic conformations and interactions of oligosaccharides”, Glycoscience: Biology and Medicine, N .Taniguchi, T. Endo, G.W. Hart, P. Seeberger, and C.-H. Wong ed. Springer (Japan), Vol.1, 137-145, 10.1007/978-4-431-54841-6_101, (2015)
10. Y. Zhang, T. Yamaguchi, T. Satoh, M.Yagi-Utsumi, Y. Kamiya, Y. Sakae, Y. Okamoto, K. Kato, “Conformational Dynamics of Oligosaccharides Characterized by Paramagnetism-assisted NMR Spectroscopy in Conjunction with Molecular Dynamics Simulation”, Advances in Experimental Medicine and Biology, 842, 217-230, 10.1007/978-3-319-11280-0_14, (2015),
11. Y. Yamaguchi, T. Yamaguchi, K. Kato, “Structural analysis of oligosaccharides and glycoconjugates using NMR”, Glycobiology of the Nervous System, R. K. Yu and C.-L. Schengrund ed. Springer New York, pp.165-183, 10.1007/978-1-4939-1154-7_8, (2014),
12. 加藤晃一, 山口拓実, “NMR 原理”, 摺らぎ・ダイナミクスと生体機能, (寺嶋正秀編), 化学同人, pp. 69-79, (2013)

【佐甲靖志】

1. 柳川正隆,佐甲靖志, “細胞膜受容体の1分子イメージング”, 生体の科学 **68**, 386-387, (2017)
2. 廣島通夫,佐甲靖志, “蛍光1分子可視化技術と超局在化顕微鏡法” 実験医学別冊「初めてでもできる！超解像イメージング」岡田康志編,羊土社, 日本, pp. 213-218, (2016)
3. C.-G. Pack, M.-K. Jung, M.-R. Song, J.-S. Kim, S.-S. Han, Y. Sako, “Use of engineered nanoparticle-based fluorescence methods for live-cell phenomena”, Fluorescence Microscopy: Super-Resolution and Other Novel Techniques, pp.153-170, Cornea, A. and Conn, P. M. eds., Elsevier, the Netherlands, (2014)
4. 佐甲靖志, 廣島通夫, 日比野佳代, “細胞内情報処理反応の1分子計測:蛋白質ダイナミクスと分子認識”, 「1分子ナノバイオ計測,分子から生命システムを探る革新的技術」,pp.180-189, 野地博行編, 化学同人, 京都, (2014)

【申惠媛】

1. *申惠媛,中山和久, 8章 Arf ファミリーによるメンブレントラフィックの調節. Dojin BioScience Series 24 メンブレントラフィック 福田光則・吉森保編, 化学同人, pp. 114-129, (2016)

【真行寺千佳子】

1. Chikako Shingyoji, “Regulation of Dynein-driven Ciliary and Flagellar Movement”, In Dyneins – Structure, Biology and Disease (2nd ed.), S. M. King ed., Chapter11. (Academic Press, N.Y.), in press, 10.1016/B978-0-12-809471-6.00011-5, (2017)
2. H. Higuchi and C. Shingyoji, “Measuring the Motile Properties of Single Dynein Molecules,” In Handbook of Dynein, K. Hirose ed., Chapter 5 (Pan Stanford Publishing Pte. Ltd.), in press, (2017)
3. Chikako Shingyoji, “Regulation of Dynein Activity in Oscillatory Movement of Sperm Flagella”, Muscle Contraction and Cell Motility, H. Sugi ed., Pan Stanford Publishing, Chap. 15, pp.371 -386, 10.1201/9781315364674-16, (2016)

【水野健作】

1. S. Fujiwara, *K. Mizuno, “Role of Intermediate Filaments in Cell Locomotion,” In: eLS (Encyclopedia of Life Sciences), A26365, John Wiley & Sons, Ltd.: Chichester, 10.1002/9780470015902.a0026365, (2017)

2. *K. Ohashi, K. Mizuno, "A novel pair of split venus fragments to detect protein-protein interactions by in vitro and in vivo bimolecular fluorescence complementation assays" in "Methods in Molecular Biology, Exocytosis and Endocytosis II", Ed. by A. I. Ivanov, 1174, 247-262, [10.1007/978-1-4939-0944-5_17](https://doi.org/10.1007/978-1-4939-0944-5_17), Humana Press, New York, USA, (2014)
3. 永井友朗, 水野健作, 「哺乳類 NDR キナーゼの細胞機能：もうひとつの Hippo 下流キナーゼ」, 医学のあゆみ, 251 (5), pp. 365-370, 医歯薬出版, 東京, (2014)

【村田和義】

1. 岡本健太, *村田和義「巨大ウイルスの構造解析：クライオ電子顕微鏡の新たな挑戦,」 特集「クライオ電子顕微鏡で見えた生命のかたちとしくみ」, 実験医学 Vol.36, No.8, P.1344-1348, 羊土社, (2018)
2. 村田和義「超高压電子顕微鏡」, ライフサイエンス顕微鏡学ハンドブック, 山科正平・高田邦昭 責任編集, pp.150-154, 朝倉書店, 東京, (2018)
3. 片山和彦, 芳賀 慧, 藤本 陽, 戸田玲子, 村上耕介, 村田和義, 中西 章「ノロウイルス研究の最新知見」 感染制御と予防衛生 Vol.1, No.1, P.4-11, メディカルレビュー社, (2017)
4. 村田和義 (共著), 「超高压電子顕微鏡による分析」, マイクロビーム アナリシス・ハンドブック 5.1.3, P.459-464, オーム社, 東京, (2014)
5. 村田和義, 「電子顕微鏡によるバイオイメージング」, 画像ラボ Vol.25, No.4, P.6-13, 日本工業出版, 東京, (2014)

【山本量一】

1. 山本量一, 大山倫弘, John J. Molina, Simon K. Schyder, 「ソフトマターのモデリング：非平衡系・生物系への挑戦」, 化学工学, 81, 282-285, (2017)
2. John. J. Molina, Ryoichi Yamamoto, Chapter 8 of "Computer Simulation of Polymeric Materials: Applications of the OCTA System", (Springer, ISBN:978-9811008146), pp 149-167, [10.1007/978-981-10-0815-3](https://doi.org/10.1007/978-981-10-0815-3), (2016)

班友

【重田育照】

1. Y. Shigeta* R. Harada* H. Matsumura, "Identification of the Key Interactions in Structural Transition Pathway of FtsZ from *Staphylococcus aureus*", 物性研究所Activity Report 2017, リサーチハイライト (invited letter), ISSN:2189-6070, pp40 (2018)
2. R. Harada* Y. Shigeta* "Recent Extensions and Applications of Parallel Cascade Selection Molecular Dynamics Simulations", 物性研究所スーパーコンピューターセンターActivity Report 2017 (invited review), pp31-38 (2018)
3. R. Harada, *Y. Shigeta, "Parallel Cascade Selection Molecular Dynamics Simulations for Transition Pathway Sampling of Biomolecules", Advances in Quantum Chemistry Vol. 78, 129-147, ISSN 0065-3276 (2018),
4. Y. Shigeta, Y. Kitagawa (分担共著) , Book Chapter (Chap. 4-1) of "Quantum and Computational Chemistry for Transition Metal Complexes", Ed. K. Yamaguchi, S. Sakaki, A Series Books of Transition Metal Complexes 10, Sankyo Publishing, ISBN-13: 978-4782707098, pp1-540 (2014)

【塚崎智也】

1. 塚崎 智也 “タンパク質膜透過装置の構造生物学” 生物物理 58, 78-82, (2018)
2. 塚崎 智也 “タンパク質膜透過駆動モーター SecDF の分子機構“ 医学のあゆみ 262, 367-372, (2017)
3. 塚崎 智也, “タンパク質を透過させる分子装置の活写” 生化学 88, 114-118, 10.14952/SEIKAGAKU.2016.880114, (2016)
4. 塚崎智也, “実験医学 (増刊「構造生命科学で何がわかるのか, 何ができるのか」)” 第2章, 第6節「2つの Sec モーター蛋白質による蛋白質膜透過のしくみ」 p113-117, 田中啓二, 若槻壮市編, 羊土社, 東京, 日本, (2014)

【前田大光】

1. 前田大光, “[ZnN₄]_n: Bis(dipyrrolylphenylethynyl)benzenes Zn^{II}-bridged polymers” 錯体化合物事典 (大川尚士他 編) , 朝倉書店, in press, (2018)
2. 羽毛田洋平・前田大光, 「イオンペアリングπ電子系集合体の創製」 超分子研究会アニュアルレビュー, 高分子学会 (Japan), 38, 10-11, (2018)
3. 前田大光, 「π電子系の合成を基盤としたイオンペア集合体の創製」 液晶 (日本液晶学会設立20周年特集記事) , 日本液晶学会, (Japan), 21 (3), 317-320, (2017)

4. Y. Haketa, H. Maeda, “ π -Electronic Ion-Pairing Assemblies Providing Nanostructured Materials”, Functional Organic and Hybrid Nanostructured Materials: Fabrication, Properties, and Applications; Li, Q. Ed.; Wiley-VCH, in press, ISBN: 9783527342549, (2017)
5. Y. Haketa, R. Yamakado, H. Maeda, “Supramolecular Assemblies of π -Electronic Charged Species”, Conjugated Objects: Developments, Synthesis, and Application, Nagai, A.; Takagi, K. Eds., Pan Stanford, in press, ISBN: 9789814774031, (2017)
6. *前田大光, 「イオンペアリング π 電子系超分子集合体」自己組織化マテリアルのフロンティア (エキゾチック自己組織化材料研究グループ 編) フロンティア出版, pp 126-136, ISBN: 978-4-902410-26-6, (2015)
7. H. Maeda, “Ion-Based Liquid Crystals: From Well-Defined Self-Organized Nanostructures to Applications”, In Nanoscience with Liquid Crystals: From Self-Organized Nanostructures to Applications; Li, Q. Ed.; Springer, Ch. 9, 281-299, [10.1007/978-3-319-04867-3_9](https://doi.org/10.1007/978-3-319-04867-3_9), ISBN: 978-3-319-04866-6, Springer International Publishing, Switzerland, (2014)
8. H. Maeda, “Supramolecular Assemblies Based on Interionic Interactions” In Synergy in Supramolecular Chemistry; T. Nabeshima, Ed.; 57-74, ISBN: 978-1466595026, CRC Press, UK, (2014)
9. 前田大光, 「刺激応答性円偏光発光を示す π 電子系」光化学 (レビュー), 45 (2), 58–63, (2014)
10. 前田大光, 「電荷を有する π 電子系の規則配列による次元制御型集合体の創製」高分子, 63, (12), 858-859, (2014)

研究成果による産業財産権の出願・取得状況

A01：動的秩序の探査

【松森信明】

1. 「新規蛍光標識スフィンゴミエリン及びその利用」, 発明者: 村田道雄、松森信明、木下祥尚、楠見明弘、鈴木健一, 権利者: 国立大学法人京都大学 国立大学法人大阪大学, 特許番号: 特許第 6398055 号, 登録日: 2018 年 9 月 14 日

A02：動的秩序の創生

【平岡秀一】

1. 「ヘキサフェニルベンゼン骨格の位置選択的交互型修飾方法」、発明者: 平岡秀一、小島達央、権利者: 国立大学法人東京大学, 番号: 特願 2014-45233, 出願日: 2014 年 3 月 7 日, 国内外の別: 国内

【二木史郎】

1. 「細胞質送達ペプチド」、発明者: 二木史朗、秋柴美沙穂、川口祥正、武内敏秀, 権利者: 国立大学法人京都大学, 番号: 特願 2016-552026, 出願日: 2015 年 9 月 28 日, 国内外の別: 国内

A03：動的秩序の展開

【菊地和也】

1. 「メチル化 DNA を蛍光標識する方法」, 権利者: 国立大学法人大阪大学、発明者: 菊地 和也、堀 雄一郎、乙村 法道, 特許番号: 特許第 6274632 号, 登録日: 2018 年 1 月 19 日
2. 「タンパク質を蛍光標識する方法」, 権利者: 国立大学法人大阪大学, 発明者: 水上進, 渡辺修二, 秋元悠里, 菊地和也, 特許番号: 特許第 5686385 号, 登録日: 2015 年 1 月 30 日

班友

【重田育照】

1. 「情報処理、シミュレーションプログラムおよびシミュレーション方法」、発明者: 中村朋健、重田育照、原田隆平、権利者: 富士通:筑波大=9:1、特許番号: 特許第 6407761 号, 登録日: 2018 年 9 月 28 日

研究成果

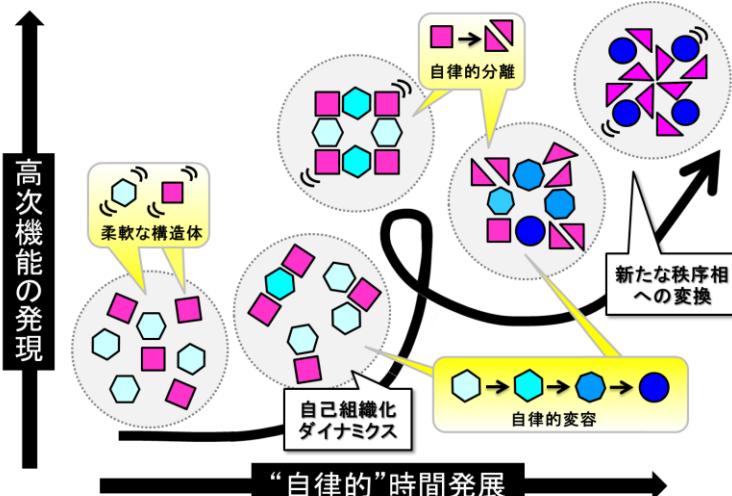
研究領域の目的及び概要

研究の学術的背景 着想に至った経緯

生命現象の特徴は、システムを構成する複数の分子素子がダイナミックな集合離散を通じて秩序構造を形成し、それが自律的に時間発展していくことがある。生命体を構成する個々の分子素子は、それぞれが複雑な3次元構造と独特の柔構造を有している。すなわち、内的複雑性を秘めた分子素子が多数集積して協奏的に高次秩序系を体現していることが生命体の特質であり、その形成原理を解明することが生命現象の本質的理解につながるはずである。

生命体の秩序は、歴史的には Schrödinger の思索において negentropy という概念で表出され、Prigogine らによって散逸構造の一形態として取り扱われてきた。当然のことながら、こうした概念は今日明らかとなっている生命分子の多様性や複雑性を念頭に置くものではない。前世紀末期に勃興したゲノムサイエンスの潮流は、「omics」と称されるアプローチによる生命情報の網羅的集積を推進してきた。実際、今世紀に入りタンパク質の立体構造データは爆発的な勢いで蓄積されており、生命素子間の相互作用ネットワークを基盤とするシステム生物学も急速に発展してきた。しかしながら、それら生命素子がダイナミックな相互作用を通じて時空間的秩序を形成し機能発現する仕組みを分子科学に立脚して理解することを目指した取り組みは驚くほど少ない状況であった。

一方、1990年代より分子の自己組織化を主題とする超分子化学の研究領域が目覚ましい発展を遂げてきた。これは、巧妙にデザインされた低分子が自発的に集積する性質を利用して一定の空間的秩序を有するナノ構造体の構築を目指すものである。こうした研究は、これまでかならずしも生命システムとの積極的な関連付けを企図して行われてきたわけではないが、自己集積を通じて個々の要素が持つえない高次機能を創発するという点において生命システムと通底する。但し、いうまでもなく、生命分子の自己組織化は、非生体系の場合と比して遙かに複雑である。それは、上述のように個々の要素分子が複雑な動的高次構造を有するということもさることながら、多種類の分子が弱い相互作用を通じて自己集積し、非対称性と可塑性を有する動的秩序構造を形成するということによる。こうした性質は、生命機能の発現と密接に関係している。さらに、生命超分子集合体は、外部環境の変動や超分子集合体間の自律的なコミュニケーションを通じて時空間的発展を遂げる。そのスケールは空間的にはサブミクロン、時間的には日(day)のオーダーに及ぶこともある。このような生命分子集団の振る舞いについての統合的理理解に向けた分子科学的取り組みはポストゲノムがうたわれて久しいにも関わらず実現しておらず、とりわけ生体分子研究者と非生体系研究者との体系的な知的連携はなされていなかった。



以上の学術的背景のもとに、生命分子が動的な秩序を形成して高次機能を発現する仕組みを分子科学の観点から解き明かし、それを通じた生命現象の本質的理解を目指して本研究を提案した。さらに、先端生命分子科学と超分子化学のアプローチを融合することによって、生命分子システムの特質を具現化した動的秩序系を人工構築することを目指した。

領域申請の理由 本領域の発展による学術水準の向上

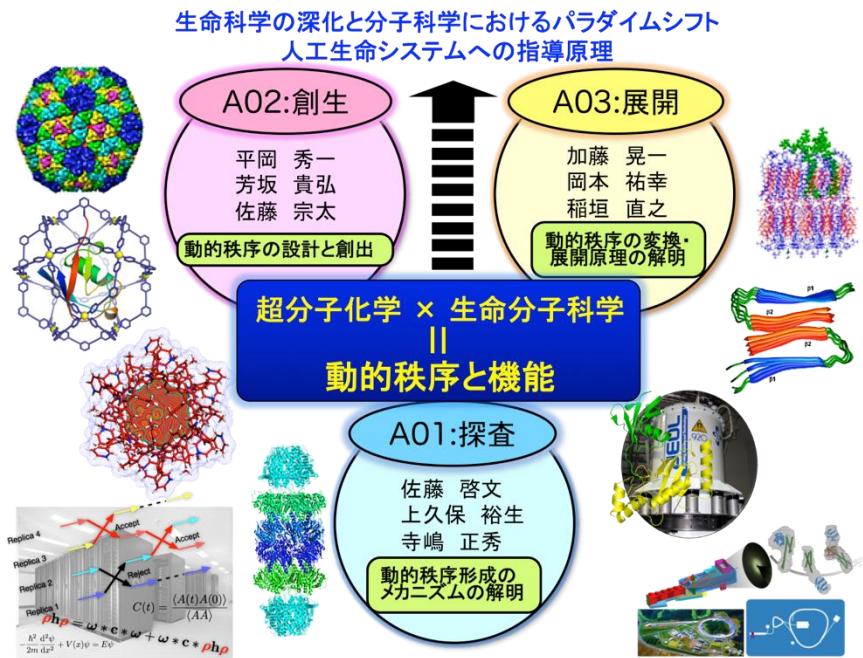
以上の目的を実現するためには、生命超分子系という巨大な森の相の変動を見つめつつ、それを構成する木々の個々の枝葉の振る舞いを捉え、両者を相關づけるアプローチが必要である。そのために、生命分子科学を基軸に、生物物理学、理論・計算科学、合成化学、構造・システム生物学、さらには医学・薬学・工学・環境科学等への応用を見据えた研究体制を構築する。現在、国際的にも生命分子の構造・機能データベースが充実し、我が国においても計算科学と実験科学を支援する大型施設が拡充している。こうした時機を鑑みても、多様なバックグラウンドの研究者が明確な問題意識を共有して叡智を結集し、生命分子の動的秩序形成の理解に向けた分野横断的な研究体制を組むことの意義は大きい。

本研究領域は、生命分子科学と自己組織化の化学の融合により、生命現象の諸相にみられる分子の秩序形成の原理を統合的に理解するとともに、その人工構築を目指すものである。その成果は、創薬をはじめとする応用研究の進展に資するばかりでなく、生命科学一般の深化と分子科学におけるパラダイムシフトをもたらし、人工的な生命システムを創生するための指導原理を導き出すことが期待される。したがって、本領域が成功裡に進展することにより、他の研究領域の発展にも極めて大きな波及効果がもたらされる。

研究の概要

本領域では、分子が自律的に集合するプロセスについて精密に探査することを可能とする実験と理論の融合研究を実施した（研究項目 A01「動的秩序の探査」）。また、生命分子科学と超分子化学のアプローチを統合することを通じて、生命分子システムの特質を具現化した動的秩序系を人工構築した（研究項目 A02「動的秩序の創生」）。さらに、生命分子の自己組織化系の設計原理を明らかにするとともに、外的擾動に対するシステムの不安定性とロバストネスを解明することを通じて、高次機能発現に至る時空間的展開の原理を理解することに取り組んだ（研究項目 A03「動的秩序の展開」）。

上記各項目において、各メンバーが独自の生命分子システムを対象に研究を進めるとともに、いくつかの特定の対象をターゲットとして、項目横断的な研究連携も推進した。その主な対象としては、細胞内のタンパク質の運命（立体構造形成・輸送・分解）にかかわる超分子マシンリー、細胞表層における糖鎖・脂質・タンパク質からなる超分子系、アルツハイマー病をはじめとする神経変性疾患にかかわるタンパク質の会合体形成、概日リズムを司る時計タンパク質複合体などを取り扱った。また、非生体分子を基盤に構築された人工的な分子集合システムについても、生体分子系と同様に精密な実験計測と理論的アプローチを展開してそのダイナミクスを明らかにし、時空間的非対称性と散逸性を有する動的秩序形成に向けての分子設計上の指針を得た。



本領域の目的を達成するために、生命分子システムにおける動的秩序の探査（A01）・創生（A02）・展開（A03）の3つの研究項目を研究の柱として設定し、それぞれの項目について化学・物理学・生物学の分野横断的な連携研究と、新規方法論の開発を含めた実験と理論の融合研究を実施し、下記の通り順調に成果を上げた。

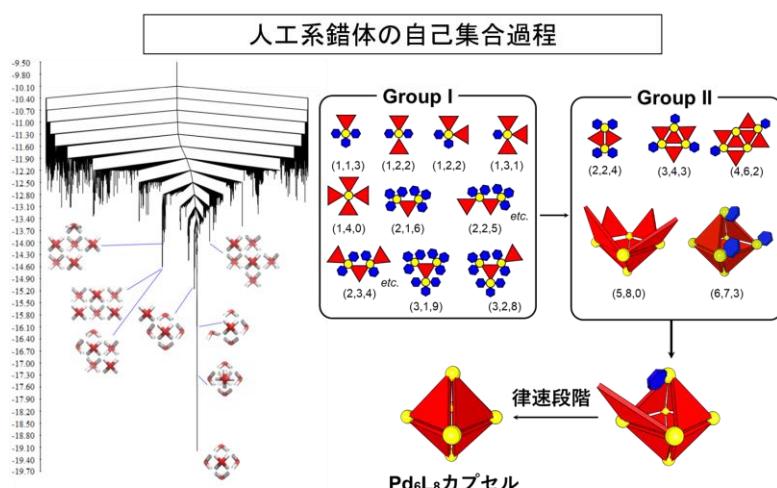
- A01** 生命システムが現有する動的秩序構造を捉えるための計測技術の開発・高度化を通じて、その形成過程の精密観測を達成し、理論と実験の両面から動的秩序形成のメカニズムを解き明かした。
- A02** 実験・理論の両面から生命システムの動的秩序を再構築するとともに、自己集積能を有する非生体系分子に生命分子の特質を賦与することにより、人工的な動的秩序形成系を創成した。
- A03** 生命分子の自己組織化系の設計指針を明らかにし、外的摂動に対するシステムの不安定性とロバストネスを解明することを通じて離合集散を経て高次機能発現に至る時空間的展開の仕組みを理解した。

このように、研究領域として設定した目的は十分に達成されたが、特筆すべきことは、各研究項目の枠組みを超えた共同研究が活発に行われたことで、期待以上に多くの成果を収めたことである。

1. 人工系と生体分子系に底通する自己集合の普遍性の解明

分子自己集合も化学反応の1つであるが、多くの中間体が生成する上に、これらを実験的に観測することが難しい。そのため、本研究領域が発足した当時、分子自己集合過程に関する分子論的解明は全くと言って良いほど進んでいなかった。分野融合的な研究を推進することにより、人工系および生命分子系の自己集合機構を分子論的に解明するためのアプローチ法を確立し、両者に通底する自己集合の普遍性を明らかにすることができた。この成果は、人工的な生命システムを設計・創生するための指導原理を確立する基盤を築くものである。

平岡（A02 計画）は、分子自己集合過程で生成する中間種を検出できないという問題を解決し、自己集合過程を解明する解析法（QASAP: Quantitative Analysis of Self-Assembly Process）を開発した。これは、自己集合の原系と生成系の全成分を定量することで、全中間種の平均組成を間接的にあぶり出す手法であり、原理的に様々な分子自己集合に応用可能である。QASAP を自己集合性錯体



生命系におけるタンパク質の集合メカニズム

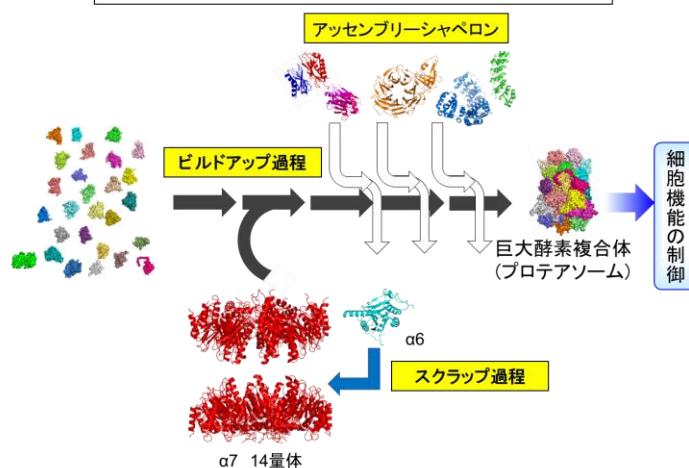


図1. 人工系および生命系における一義自己集合体の形成機構の解明。(上)理論的なアプローチと実験的アプローチを合わせることで、自己集合過程で生成する観測不可能な中間種の同定に成功。(下)プロテアソームの自己集合過程におけるアッセンブリーシャペロンの役割を解明するとともに、スクラップ・アンド・ビルト過程の存在を見出した。

に適用し、14種類に及ぶ自己集合体の形成機構を解明することで、分子自己集合がエネルギーランドスケープ上を様々な経路を経て進行することを明らかにした。そして、佐藤(啓)（A01 計画）はマスター方程式を使った自己集合過程に迫る新しい理論手法を開発し、これらの自己集合性錯体のより詳細な形成機構

を理論的に解明した。このように、実験と理論の協働により、人工分子の自己集合過程がタンパク質のフォールディングと相同性をもつことを明らかにした（図 1）。さらに、佐藤（啓）は量子化学計算や REST (replica exchange with solute tempering) 法を使って、平岡が開発した箱型六量体「ナノキューブ」の自己集合過程を理論的に解明することにも成功した。ナノキューブは疎水効果やファンデルワールス力などの生命系で頻繁に利用されている弱い因子により集合化しているため、この自己集合体の形成過程の理解は、生命系の自己集合過程の理解に繋がるだけなく、生命分子の特性を兼ね備えた人工分子のデザイン指針の確立にも欠かせない。

一方、加藤（A03 計画）は、複雑な生命系におけるタンパク質分子の自己集合過程として、多数のサブユニットから構成されるタンパク質分解装置プロテアソーム複合体の動的秩序形成機構の解明に向けて、各ペース分子の動的秩序形成におけるミクロ・マクロ相関を探査した。プロテアソームの分子集合に一過的に関わる一連のアッセンブリーシャペロンがサブユニットの適切な配置を定める“チェックポイント”や“分子マッチメーカー”として機能していることを解明した。さらに、プロテアソームの 4 次構造形成にスクランブル・アンド・ビルトのプロセスが織り込まれることにより、精密な分子集合が可能になっていることを明らかにした（図 1）。

これに呼応するかのように、平岡は、人工分子の自己集合過程で一過的に生成する中間種の中には、最終構造の 50 倍以上も大きい種が形成することや、非対称化された準安定種が存在することを突き止めた。さらに、構成成分では無い分子の介在が自己集合に必須である系も見出し、生命系におけるシャペロンに近い機構が人工系にも存在することを明らかにした。これにより、人工系と生体分子系における自己集合の普遍性がより明確に浮き彫りとなった。

2. 分子集合のエネルギー・ランドスケープにおける速度論支配と熱力学支配の制御

分子自己集合には上記のような一義自己集合体の他に、線維やシートなど、ある次元について無限に広がる自己集合体もある。これらは、動的秩序形成の普遍性を論ずるのに格好の題材であり、その形成機構についても人工系および生命系の実験と理論研究者が共同で問題に取り組むことで重要な知見が得られた（図 2）。杉安（A02 公募）は、ポルフィリン環に置換基を導入した分子が、はじめに微粒子を速度論的に形成し、これが単量体への解離平衡を経て熱力学的に安定な線維構造へ変換することを発見し、さらに導入する置換基をわずかに変化させるだけで、シート構造を生成することに成功した。これらの成果は、分子集合のエネルギー・ランドスケープにおける速度論支配と熱力学支配を制御することで超分子形成の時間発展のプログラミングが可能であることを示した。

生命系において、茶谷（A03 公募）はタンパク質の線維状集合体の形成過程で線維前駆中間体が一過的に生成することを見出し、この中間体の同定にも成功した。さらに、線維前駆中間体に先駆けて生成する粒子状集合体の検出にも成功し、特定の中間体についてのみ超音波照射により線維形成を引き起こす活性種へ変換できることを明らかにした（図 2）。また、奥村（A03 公募）はハミルトン置換分子動力学シ

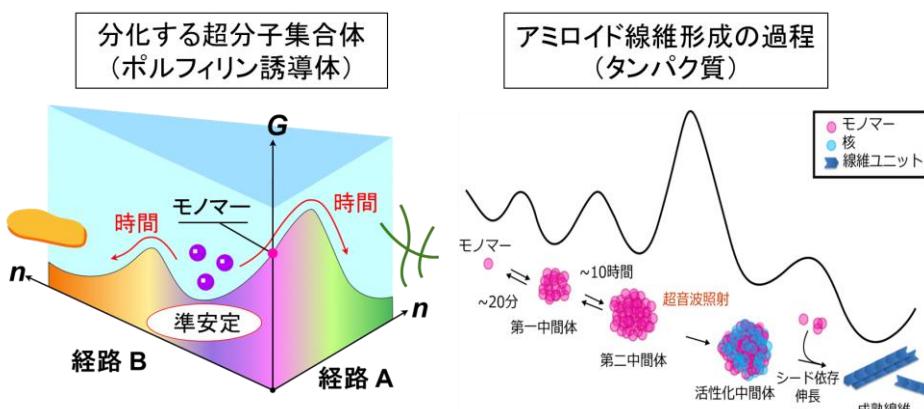


図 2. 人工系および生命系における線維やシートの自己集合過程の解明。
「人工分子の自己集合におけるエネルギー・ランドスケープのコントロール(左)」と「タンパク質の集合化による線維状自己集合体の形成機構(右)」には高い類似性があり、人工系と生体分子系に普遍な自己集合の原理が見出された。

ミュレーションによって、タンパク質の線維形成のシミュレーションに成功し、単量体が接近することでヘアピン構造が増加し、分子間のシート構造が増えることを見出した。さらに、この2量体に単量体が結合することで三量体、四量体と成長し、核へ単量体が結合することで集合化が進む機構で線維形成が起ることを突き止めた。このように、これまで異なるものとして認識されていた現象が、本領域の活動を通じて体系的に理解できるようになり、その根底に共通する機構に基づいて分子集合をプログラムすることが可能となった。

3. 非平衡系における分子の離合集散による秩序形成の理解

生命系の特徴は、一度形成された秩序構造が安定に維持されるだけでなく、集合・解離を繰り返すことで、動的に秩序を生み出していることである。本研究領域では、このような系についてもミクロとマクロのダイナミクスをつなぐ理解を促した（図3）。

時計タンパク質は24時間という長い周期で離合集散を繰り返し、概日リズムを司っている。秋山(修)(A01公募)はシアノバクテリアの生物時計を構成するKai Cタンパク質に24時間の周期を司る機構が存在することを突

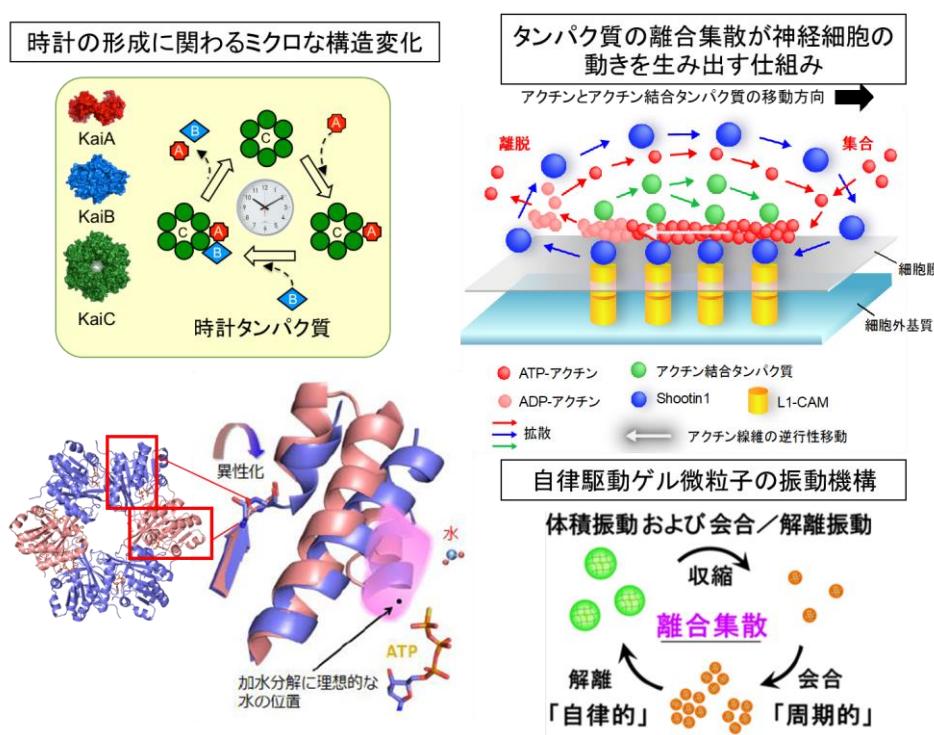


図3. 離合集散を繰り返す動的な秩序形成の機構の解明。時計タンパク質のうち、KaiCが24時間のリズムを司り、この長い周期をコントロールする分子機構が明らかになった。また、神経軸索の形成におけるアクチン波の新規機構を解明した。さらに、温度とpHという2種類の外的因子に独立して応答する自立駆動ゲル微粒子の創出が実現した。いずれも離合集散により同調的に周期的律動をする分子システムの理解につながる成果である。

き止め、長い周期を刻む分子機構が、立体的な要因とペプチド鎖の異性化という2つの因子がATPの加水分解を極端に遅らせていることに起因することを明らかにした。稻垣(A03計画)は神経細胞における軸索の形成機構の解明に取り組み、アクチン線維が神経細胞の伸長方向へ重合し、後方で脱重合するアクチン波が起こる際に、細胞接着タンパク質を介して細胞外基質に連結されるという従来のモデルを覆す新しい機構であることを明らかにした。さらに、神経軸索が誘引分子を検知して軸索を伸長させるための力を生み出す仕組みも解明した。このように、生命系における離合集散の分子論的理解が飛躍的に進んだが、併せて、人工系においても周期を刻み律動する分子を開発することに成功した。鈴木(A02公募)はBelousov-Zhabotinsky(BZ)反応を温度応答性の高分子と複合化することで、周期的に自律駆動するゲル微粒子を開発した。また、その作用機構を解明するとともに、高田(A01公募)との共同研究を通じて、ゲル微粒子の架橋点にロタキサン構造を導入することで、pHと温度といった複数の刺激に対してそれぞれ応答できるようになり、生命系に迫る動秩序系を構築することにも成功した。

本領域は、人工系と生命分子系の動的秩序形成に本質的な相同性を見出し、両者に通底する自己集合の原理を浮き彫りにすることことができた。さらに、非平衡系における生体分子の離合集散過程について、実験と理論の両面から詳細に解き明かした。それらの特質を備えた人工分子の創生や、同調的に周期的律動をする人工高分子ゲルや生体分子と人工超分子のハイブリッド化による高次機能の創出にも成果を収めた。このように、提案当初に掲げた目標を十二分に達成することができた。

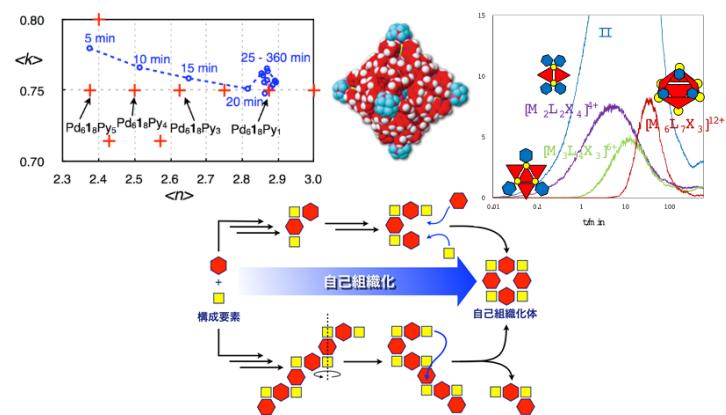
主な研究成果

研究項目 A01 「動的秩序の探査」

動的秩序形成を探査する分子理論のアプローチ法を開拓し、超分子の自己組織化メカニズムを解明

主な発表論文：*Phys. Chem. Chem. Phys.* 2018【A01 佐藤(啓)、A02 平岡の共同】、*Phys. Chem. Chem. Phys.* 2018【A01 立川、A01 重田、A02 平岡の共同】、*Chem. Eur. J.* 2017【A01 立川、A02 平岡の共同】

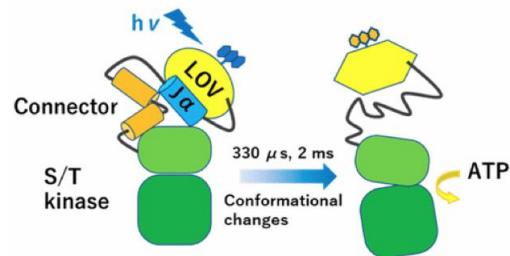
理論を担当する佐藤(啓) (A01 計画) は、平岡 (A02 計画) と協力しながら、自己集合過程の物理化学的原理の解明に取り組み、秩序形成過程をマスター方程式や量子化学計算によって追跡する方法を提案した。これにより、自己集合過程の実験的観測を実現した平岡の成果をサポートするだけでなく、中間体の生成・分解の経時変化や、自己集合律速段階をコントロールする因子を解析し、秩序形成のメカニズムを明らかにした。また、立川や重田(ともに A01 公募)は、量子化学計算や MD 法を駆使し、溶媒効果が会合体の安定性や動態に与える影響や、分子自己集合過程における速度定数や活性化エネルギーを求める成功に成功した。



タンパク質の過渡的相互作用を捉え、マルチドメインタンパク質の作動原理を解明

主な発表論文：*J. Phys. Chem. B* 2017【A01 寺嶋】、*Sci. Rep.* 2017【A01 上久保、A01 内橋、A03 加藤、A03 村田の共同】、*J. Am. Chem. Soc.* 2016【A01 寺嶋、A02 神谷の共同】

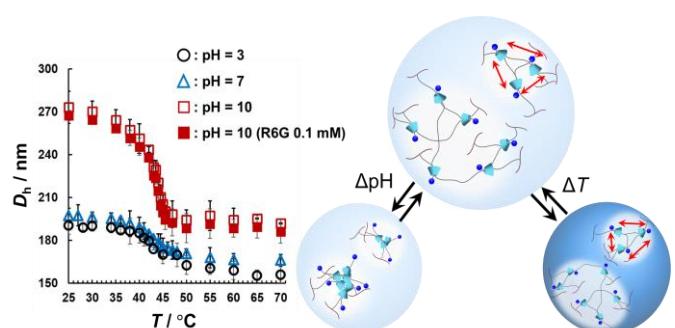
タンパク質が機能を発揮する際には、内部のミクロ構造変化のみならず、様々な機能をもったモジュール構造のマクロな時空間的变化が重要な鍵となる。寺嶋 (A01 計画) は、独自に開発した過渡回折格子法を武器に秩序構造ダイナミクスを追跡し、巨大なマルチドメイン構造をもつ光センサータンパク質が局所構造を維持しつつドメイン間の空間配置を変化させる様子の時間分解測定に成功した。さらに本手法を神谷 (A02 公募) が開発した光応答性人工核酸システムと組み合わせ、DNA2 重差構造の解離過程の詳細を解明することにも成功した。上久保 (A01 計画) は、多成分系の溶液散乱データの系統的収集を実現する微量自動サンプリングシステムを開発した。領域内の 4 グループ (上久保、内橋 (A01 公募)、加藤 (A03 計画)、村田 (A03 公募)) が連携することで、溶液散乱計測や高速 AFM 観察による動的構造解析と、電子顕微鏡、結晶構造解析の静的観察結果を統合し、細胞内秩序維持に関わる小胞体フォールディングセンサー酵素の、柔軟なモジュール構造を巧みに用いた作動機序を明らかにした。



“お互いに干渉しない”多刺激応答性ゲル微粒子の実現

主な発表論文：*Angew. Chem. Int. Ed.* 2017【A01 高田、A02 鈴木の共同】、

生命活動の維持には、複数の刺激に適切に応答し、構造や機能の変化を行うことが不可欠である。こうした動的秩序システムの人工系での実現を目指し、高田 (A01 公募) は鈴木 (A02 公募) と協力して多刺激応答性を有するソフトマテリアルを創出した。人工系での分子設計においては、1 つの刺激に対応すると、他の刺激応答性が著しく消失してしまうという問題が生じやすい。そこでロタキサン構造に由来する環動性をゲル微粒子に導入することで問題を克服し、温度と pH の変化に対し相互不干渉に応答する分子システムを実現した。

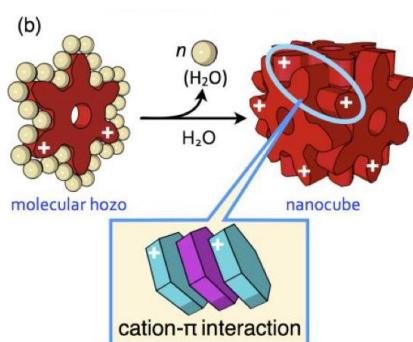


研究項目 A02 「動的秩序の創生」

弱い相互作用を積み上げて高次構造体を創生

主な発表論文： *Commun. Chem.* 2018 【A01 立川、A02 平岡、A03 内山の共同】、*Chem. Comm.* 2017 【A01 岩田、A01 松森、A02 大谷の共同】、*Nature* 2016 【A02 佐藤(宗)】

生命分子システムの動的秩序化においては、疎水効果やファンデルワールス力などが無数に集積することで強大な束縛力を産み出す。人工系においてもこうした弱い相互作用を制御することで高次構造体を創出するとともに、物理化学計測を通して秩序形成のメカニズム理解の深化を図った。平岡（A02 計画）は、立川（A01 公募）とともに分子同士の嗜み合わせを評価する手法を確立した。ここから得られた知見をもとに精密設計した歯車状分子の自己組織化を通して、超好熱菌由来タンパク質の熱耐性をも上回る安定性をもった超分子を開発することに成功した。佐藤(宗)（A02 計画）は配位結合によって 140 を超える構成成分からなる人工分子を創出し、X 線結晶構造解析を通して、構成成分数の増加に伴い従来の安定な幾何学的制約を打ち破る秩序が新たに創出されることを発見した。また大谷（A02 公募）は、金属錯体を含む人工脂質からなるリポソームを設計するとともに、岩田や松森（ともに A01 公募）との連携による脂質ダイナミクス解析を通して、表面は硬く内部は柔らかいという膜構造の不均一性を明らかにした。

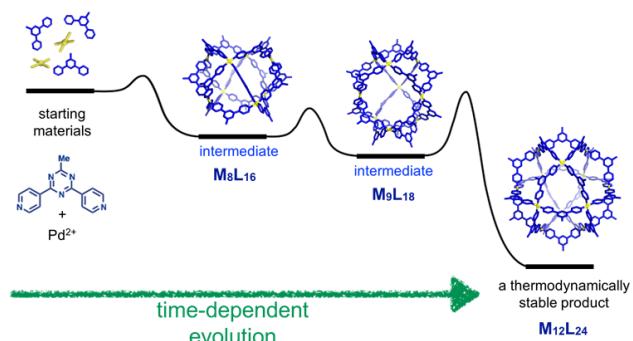


超分子集合体形成の時間展開の発見と制御

主な発表論文： *Nature Chem.* 2017 【A02 杉安】、*Angew. Chem. Int. Ed.* 2015 【A02 佐藤(宗)】

佐藤(宗)（A02 計画）は、時間展開する動的な秩序化システムの過程を、中間体構造の実験的な構造決定も含めて解明し、36 成分からなる金属錯体分子の自己組織化の過程が、実は、幾何学的に制約されることで簡素化されて、たった 2 種類の準安定な中間体構造を経て、単一の生成物に落ち着く、明瞭なシステムであることを明らかにした。

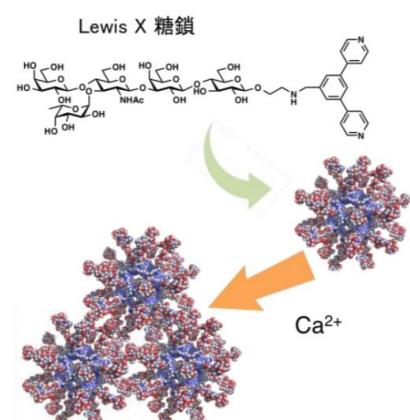
また杉安（A02 公募）は、分子デザインを通して熱力学的な準安定状態を利用して秩序化を制御することに成功し、1 つの初期状態から全く異なる 2 つの終状態へ分化する超分子の創生や、超分子集合体形成の時間発展プログラミングを実現した。



サイボーグ超分子を創生して糖鎖クラスターの相互作用の観測に成功

主な発表論文： *Chem. Asian J.* 2017 【A02 佐藤(宗)、A03 加藤の共同】、*Angew. Chem. Int. Ed.* 2015 【A02 佐藤(宗)、A03 加藤の共同】

佐藤(宗)（A02 計画）と加藤（A03 計画）は、生体分子と人工超分子のハイブリッドによるサイボーグ超分子を創生し、アルツハイマー病の発症にかかるアミロイド β タンパク質と糖鎖クラスターとの相互作用解析に応用した。さらに、機能性糖鎖を組み込んだ人工分子クラスターのデザインを発展させ、外部環境に応答して高次の離合集散を行う性質を賦与した超分子複合体を創生した。外部環境に応じた離合集散の過程においては糖鎖-糖鎖間の分子認識が重要な役割を担うことを明らかとした。



人工細胞システムの創出と応用

主な発表論文： *Proc. Natl. Acad. Sci. USA* 2017 【A02 松浦】、*Chem. Commun.* 2016 【A03 栗原】、*ChemBioChem* 2015 【A02 芳坂、A02 松浦の共同】

生体分子や人工分子を組み合わせて構成した人工細胞の創出と応用も進んだ。芳坂（A02 計画）と松浦（A02 公募）は、人工細胞システムを用いた進化分子工学手法によりタンパク質を改良・進化させ、非天然アミノ酸を導入するための高機能型 tRNA 合成酵素を作り出すことに成功した。さらに松浦は、細胞モデル中の 241 成分の濃度変化を計算し、タンパク質合成反応をシミュレーションするプログラムを開発し web 上に公開した。また栗原（A03 計画分担）は人工細胞モデルの化学的進化を進め、生命の必須要素である代謝のための触媒が内在し、自己生産するベシクルを合成した。

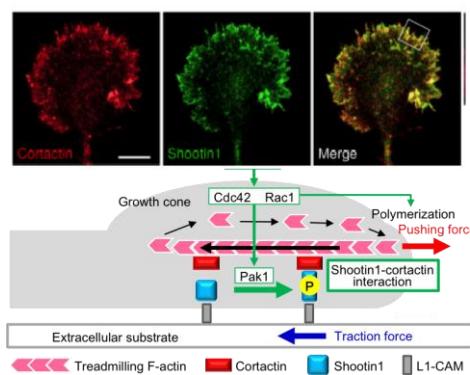
研究項目 A03 「動的秩序の展開」

力と分子の滑りを利用した神経ネットワーク形成の新しい仕組みを解明

主な発表論文：*Proc. Natl. Acad. Sci. USA*, 2018【A03 稲垣】、

Cell Reports 2015【A03 稲垣、A03 水野の共同】

神経細胞は、脳内の分子の濃度勾配に導かれて軸索を伸ばし、適切な神経細胞と結合することで脳の活動に必要な神経ネットワークを作る。稻垣（A03 計画）は、細胞接着分子 L1-CAM が細胞膜上で軸索を伸ばすための推進力を伝えるタイヤのような働きを見出しつつ、細胞と細胞外環境の間に生じる力と分子の滑りを巧妙に利用した新たな走触性の仕組みを明らかとした。稻垣と水野（A03 公募）は、既知のモータータンパク質による輸送とは異なる新しい細胞内分子輸送機構を明らかとした。



糖鎖の3次元構造ダイナミクスが展開する仕組みを解明

主な発表論文：*Sci. Rep.* 2017【A03 岡本、A03 加藤の共同】、*Adv. Exp. Med. Biol.* 2015【A02 神谷、A03 岡本、A03 加藤の共同】、*J. Cell Biol.* 2015【A02 神谷、A03 加藤の共同】

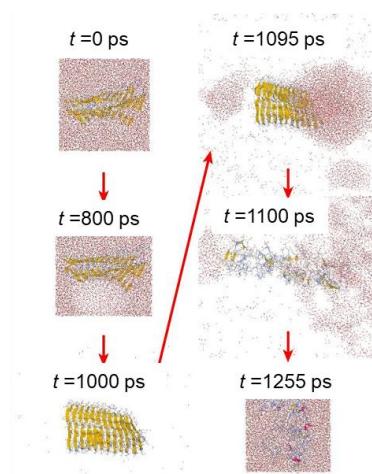
細胞内で新たに作られたタンパク質が小胞体において立体構造を形成する際や、固有の立体構造獲得後にゴルジ体へと移る際、あるいは不要となったタンパク質が分解処理されるプロセスには、糖鎖をはじめとする多数の生命分子素子が関与している。岡本（A03 計画）は、レプリカ交換分子動力学シミュレーションによって、複雑な分岐構造を有するオリゴ糖鎖の配座空間を探査した。こうした理論的アプローチに、加藤（A03 計画）が開発してきた常磁性 NMR 法による精密実験データを組み合わせることにより、輸送や分解といったタンパク質の運命を規定する糖鎖の立体構造のダイナミクスや、タンパク質によって糖鎖が認識されるプロセスを明らかにすることに成功した。

アミロイド線維の形成と崩壊の過程を捉えた

主な発表論文：*J. Phys. Chem. B* 2016【A03 奥村】、*Sci. Rep.* 2015

【A03 茶谷、A03 杉山の共同】

奥村（A03 公募）は、独自開発したハミルトニアンレプリカ置換分子動力学計算手法により、アミロイド β ペプチドの線維化初期段階におけるシミュレーションを行い、分子間 β シート構造形成による 2 量体形成過程を観測することに成功した。さらに、外部刺激による線維破壊のメカニズムを明らかにするとともに、線維伸長の方向が制御されるメカニズムを解明した。さらに、茶谷と杉山（ともに A03 公募）の共同研究では、線維形成反応の開始に同期させた時分割小角 X 線散乱測定を実施し、タンパク質の初期集合体の形状や、徐々に肥大成長しながら成熟線維へと構造発達する様子を捉えることに成功した。



時計じかけの秩序形成の仕組みを解明

主な発表論文：*Sci. Rep.* 2016【A03 加藤、A03 内山、A03 杉山、A03 寺内の共同】、*Science* 2015【A01 秋山(修)】

ATP の結合と加水分解に共役してダイナミックな離合集散を伴うシアノバクテリア時計タンパク質の作動原理も明らかとなりつつある。秋山(修)（A01 公募）は、ATP の加水分解にはペプチド主鎖の異性化反応を伴った大規模な構造転移、24 時間周期の遅いリズムの根源が KaiC の構造の中にプログラミングされていることを明らかとした。A03 班の複数のグループの共同研究（寺内、杉山、内山、加藤）では、未変性質量分析および中性子小角散乱を利用して KaiB-KaiC タンパク質複合体の化学量論数決定と複合体を構成するサブユニットの空間配置情報を明らかにした。

生体膜の動的秩序と膜タンパク質の離合集散動態の解明

主な発表論文：*Nature Commun.* 2017【A03 申】、*Sci. Rep.* 2017【A01 松森、A03 老木の共同】、*J. Phys. Chem. Lett.* 2017【A01 内橋、A03 老木の共同】

申（A03 公募）は、シグナル依存的なフリッパーゼの活性調節メカニズムを解明し、細胞膜脂質 2 重層でのリン脂質非対称性を維持するシステムが解明された。脂質 2 重層間のリン脂質の移動（フリップ・フロップ）によって細胞膜の曲率形成を駆動することを明らかにした。老木（A03 公募）は、松森や内橋（ともに A01 公募）との連携によりイオンチャネルのゲーティングに伴って膜中で集合離散する動態を初めて明らかにした。

