# 科学研究費助成事業 研究成果報告書

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研究課題名(和文)極低温単粒子解析による哺乳動物のF-ATP合成酵素の原子レベルの構造機能研究

研究課題名(英文)Single particle cryoEM of mammalian FoF1 ATP synthase

#### 研究代表者

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研究成果の概要(和文): ミトコンドリア内膜のプロトン駆動力によるATP合成は哺乳動物の最も重要なエネルギー変換過程であり、それはF-ATP合成酵素により駆動されている。この極めて基盤的な生理過程を理解するためにはこの酵素の複雑な構造を原子レベルで解明することが必須である。我々はATP合成酵素の新い精製方法などの確立に成功してF-ATP合成酵素の新しい機能を発見した。これでこれからの新しい構造機能解析研究の基礎を作った。

研究成果の学術的意義や社会的意義 哺乳類のF-ATP合成酵素は、ミトコンドリア疾患の鍵を握る存在として現在注目されている。今回のプロジェクトで得られた新しい調製法、膜の再構成、透過性遷移孔としての機能の発見は、F-ATP合成酵素の研究を前進させる上で重要な役割を果たすことが期待される。

研究成果の概要(英文): All the cells in our body rely on the sufficient and well balanced energy supply in the form of the molecule ATP. Most ATP is produced by a large membrane supercomplex, the F-ATP synthase. How this very large protein complex achieves its energy transformation function is not very well understood yet. In this project we attempted to improve our understanding by inventing novel ways to purify fragile F-ATP synthase from cow heart muscle tissue and analyze its structure and function by cryo electron microscopy and other methods. We succeeded to find new ways to purify the protein complex that preserves it during isolation. All physiological functions of the F-ATP synthase could be preserved. We also invented novel ways to put the F-ATP synthase into artificial membranes, which was important to confirm new channel functions of F-ATP synthase. In summary, we have now established a firm basis for structure and function analysis in solution as well as in artificial membrane systems.

研究分野: 生物化学

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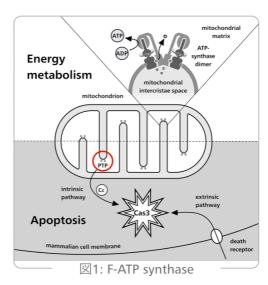
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#### 1. 研究開始当初の背景

The mitochondrial F-ATP synthase is central to energy metabolism and recycles the vast majority of ATP in the mammalian cell. In last two decades its involvement in the architecture of the inner mitochondrial membrane and numerous chronic, age related diseases became apparent. Despite the important cellular role of mitochondrial F-ATP synthase, the atomic structure of its monomeric, dimeric and oligomeric state have remained elusive. Fundamental to this crucial lack of structural insight are the lack of suitable techniques for the isolation of this large membrane complex (MW 1.3 MDa for the F-ATP synthase dimer) from its physiological membrane environment and also the inability to reconstitute its oligomeric form into membranes in vitro.

#### 2. 研究の目的

The aim of this project is to establish novel methods for the purification of the mammalian F-ATP synthase from bovine heart muscle tissue in its monomeric, dimeric and oligomeric state. Crucially, the isolation procedure is aimed at obtaining F-ATP synthase in as pure as possible form in terms of lack of contamination, absence of partial degradation and also in its pure oligomeric form. That is pure also in the sense of absence of self-contamination by the presence of a mix of oligomeric states. Furthermore, novel methods of membrane reconstitution are to be established for the functional characterization of the preparation to a point where physiological relevance of the purified F-ATP synthase can be demonstrated. The



thus obtained preparation is then used for structure determination in solution by single particle cryo-EM and in the membrane by cryo electron tomography. Finally, functional and structural data is combined to arrive at a deeper understanding of how mammalian F-ATP synthase is achieving its energy transforming function, how it as capable to determine inner mitochondrial membrane architecture and how it is connected to chronic and age related diseases such as heart disease, muscular dystrophy, Parkinson and Alzheimer's disease.

#### 3. 研究の方法

# (1) Purification

Despite a 60 years history a suitable purification method for mammalian F-ATP synthase has not been established yet. We use fresh bovine heart muscle tissue to obtain sufficient amounts of mitchondria for large scale purification which is generally favorable for fragile multisubunit protein complexes. Mild detergent solubilized inner mitochondrial membranes are then subjected to density gradient and column extraction of the F-ATP synthase.

#### (2) Liposome reconstitution

For a full functional characterization of the F-ATP synthase preparation it is crucial to test the complexes functionallity in closed membrane systems that mimic the physiological environment of the inner mitochondrial membrane. GraDeR mediated liposome reconstitution was employed to measure proton pumping and novel channel functions.

#### (3) Native Mass Spectrometry

With our collaborators in Oxford new, direct native mass spectrometry was developed and tested on our F-ATP synthase, inner mitochondrial membrane and mitochondria preparation.

#### (4) Cryo Electron Tomography

Liposome reconstitution was monitored using cryo electron tomography to examine the presence or absence of dimeric F-ATP synthase after liposome reconstitution.

#### (5) Single Particle cryo-EM

For structure determination single particle cryo-EM is used first for screening of grid freezing conditions and then for image data collection.

#### 4. 研究成果

#### (1) F-ATP synthase purification

We succeed to isolate large amounts of mammalian F-ATP synthase from bovine heart muscle tissue (>100 mg) at a very low level of contamination (<5%) exhibiting a high oligomycin inhibition rate (>98%) using a density gradient and Poros column based approach. That using this method it is possible to isolate tetrameric F-ATP synthase was demonstrated as well.

# (2) Oligomer Pure F-ATP synthase purification

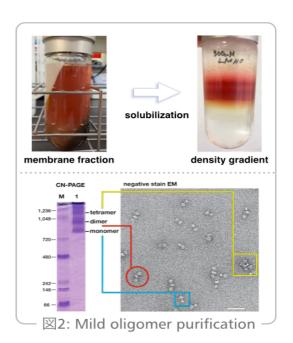
A column free purification approach is being established that allows the clean separation of monomeric, dimeric and oligomeric F-ATP synthase. Progress has been made, but further work is necessary.

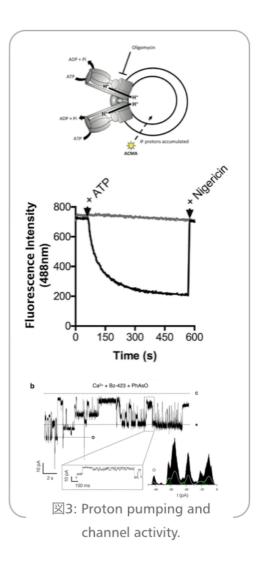
## (3) Liposome Reconstitution

Reconstitution techniques that allow dimeric or oligomeric mammalian F-ATP svnthase in vitro lipid bilaver reconstitution were established demonstrated for the first time. This success allowed to probe Opal - F-ATP synthase interaction in (Quintana-Cabrera et al., Nat. Comm., 2018) and to measure proton pumping efficiency with high accuracy (Urbani et al. Nat. Comm., 2019). In the same study the new, GraDeR based reconstitution technique could be used to unequivocally establish that mammalian F-ATP synthase has a thus far unrecognized channel function as the molecular identity of the mitochondrial permeability transition pore (mPTP) (Urbani et al., Nat. Comm., 2019).

## (4) Native Mass Spectrometry

High resolution structures rarely reveal and identify all structural lipids associated with membrane protein complexes. Our collaborators in Oxford of the Robinson lab provided proof of native principle data that spectrometry on isolated bovine F-ATP synthase, inner mitochondrial membranes and even whole mitochondrial is feasible (Chorev et al., Science, 2018).





#### (5) Cryo Electron Tomography

Among the structures of desire for mammalian F-ATP synthase are structures determined in their physiological environment of the lipid bilayer. Cryo electron tomograms of bovine F-ATP synthase proteoliposomes successfully visualized dimeric complexes confirming the ability to reconstitue oligomeric forms.

#### (6) Single Particle Cryo-EM

Cryo-grid freezing conditions were screened and fragility of the complex to blotting and freezing established. The mix of rotational states and oligomeric forms in combination with the fragility of mammalian F-ATP synthase proved to be a challenge. The publication of a 6 Å single particle cryo-EM structure of tetrameric porcine by a team from Tsinghua University in 2019 shows that the IF1 inhibited form could be a suitable target for single particle cryo-EM. The same work which used two months of data collection time and highly impure preparation (<20% target complex) also demonstrated the need for new purification approaches. Further work is needed to establish the ability to obtain reliable atomic models of mammalian F-ATP synthase.

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