

令和 6 年 6 月 7 日現在

機関番号：13301

研究種目：若手研究

研究期間：2020～2023

課題番号：20K16262

研究課題名(和文) Real-time observation of native conformations and molecular behaviors of viral fusion proteins using high-speed atomic force microscopy (HS-AFM).

研究課題名(英文) Real-time observation of native conformations and molecular behaviors of viral fusion proteins using high-speed atomic force microscopy (HS-AFM)

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交付決定額(研究期間全体)：(直接経費) 3,200,000円

研究成果の概要(和文)：応募者は、HS-AFMを用いて2つのウイルス融合タンパク質、インフルエンザAヘマグルチニン(HA)およびSARS-CoV-2スパイクタンパク質(Sタンパク質)の分子特性を調査しました。HS-AFMを使用して、HAのネイティブコンフォメーションおよび動的挙動、特に融合遷移、およびHAと脂質層の相互作用が記録されました。同様に、Sタンパク質の構造動態、Sタンパク質とACE2タンパク質の相互作用、Sタンパク質とエクソソームの相互作用、およびSタンパク質の中和相互作用もHS-AFMを使用して可視化および分析されました。その結果は、高インパクトのジャーナルに報告されています。

研究成果の学術的意義や社会的意義

HS-AFM imaging allows the applicant to study structural dynamics of viral proteins and dynamic viral protein-antibody interactions, in which it is difficult to do using conventional methods such as cryo-EM. The applicant's studies have been reported in newspapers and on news television channels.

研究成果の概要(英文)：The applicant has investigated molecular properties of two viral fusion proteins: influenza A hemagglutinin (HA) and SARS-CoV-2 spike protein (S protein) using HS-AFM. Native conformation and dynamic behavior of HA, particularly the fusogenic transition, and the interaction between lipid layer and HA have been recorded using HS-AFM. Likewise, structural dynamics of S protein, S protein-ACE2 protein interaction, S protein-exosome interaction, and S protein-neutralizing interaction, have been visualized and analyzed using HS-AFM. The results have been reported in high impact journals.

研究分野：ウイルス学関連

キーワード：Viral fusion proteins HS-AFM Hemagglutinin Spike protein SARS-CoV-2 Influenza A

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## 様式 C - 19、F - 19 - 1 (共通)

### 1 . 研究開始当初の背景

Viral-cell fusion is a critical event for viral infection. This event is mediated by one or more viral glycoproteins including one in general called fusion protein. Viral fusion proteins are highly dynamic. These proteins share a unified conformer known as a-trimer-of-hairpin. This conformer executes the fusion between viral membrane and host membrane to release viral genomes into host cytoplasm. Conformational changes of viral fusion proteins are triggered by several mechanisms such as low pH and receptor binding. Current conventional techniques including X-ray crystallography, NMR, Cryo-EM, and FRET, are incapable of capturing transient conformations of viral fusion proteins after triggered. In addition, the molecular behavior of viral fusion proteins when interacting with their target membranes remains a mystery. High-speed atomic force microscopy (HS-AFM) allows scientists to directly visualize the conformational changes of biomolecules and the interaction between biomolecules at nanoscopic range with high spatiotemporal resolution. The key scientific question in this proposal is- “Can we use HS-AFM to visualize the native conformations of viral fusion proteins and their conformational dynamics after triggered by low pH with or without the presence of target membrane?”

### 2 . 研究の目的

The purpose of this project was to investigate the native conformation and conformational changes of viral fusion protein triggered by low pH either with or without the presence of membrane (exosomes). Furthermore, this project determined the interaction between neutralizing antibodies with native or triggered form of viral fusion proteins. This project provides significant findings in molecular behavior of viral fusion proteins after triggered by acidic environment, particularly the conformational dynamic of the viral fusion proteins while interacting with exosome at acidic environment. These findings are essential for developing neutralizing antibodies and inhibitors to prevent membrane fusion.

### 3 . 研究の方法

Prior to HS-AFM imaging, the applicant performed in silico analysis to obtain HS-AFM simulated images of viral fusion proteins and their surface charges. The simulated images were used as references by the applicant to identify target proteins during HS-AFM scanning. Protein surface charges information is important for the applicant to select suitable substrates for sample adsorption.

Recombinant viral fusion proteins, respective receptors, and neutralizing antibodies were purchased from companies. Exosomes were isolated by collaborators. Viral fusion proteins were scanned under a near-physiological buffer to obtain their native structures and dynamic behaviors. Manipulations of scanning condition were performed to investigate several aims planned in this research including: fusogenic transition of viral fusion proteins, viral fusion protein-receptors interactions, viral fusion proteins-exosomes interactions, and viral fusion proteins-neutralizing antibodies interaction.

HS-AFM videos were processed and analyzed using ImageJ software. Quantitative analyses and statistical comparisons were performed using SPSS. Graphs were plotted using GraphPad prisms. Adobe Illustrator was used to prepare figures with publication qualities. Movies were edited and compiled using Adobe Premiere Pro software.

#### 4 . 研究成果

First, the applicant elucidated the conformational dynamics of hemagglutinin (HA) of Influenza A virus. He managed to capture fusogenic transition of HA at low pH. Additionally, real-time docking of HA on lipid membrane of exosome was successfully filmed. Distinct interaction patterns between HA and exosomes at neutral pH or acidic pH were found, which HA destabilized exosomal layer only at low pH. At low pH, HA underwent conformational change and released fusion peptide. The fusion peptide inserted into exosomal layer and disrupted its integrity. The results are published in Nano Letters (PMID: 32787163).

Next, during the COVID outbreak, the applicant performed HS-AFM imaging to study molecular properties of SARS-CoV-2 spike protein (S protein). The results revealed rigid trimeric head of S protein (S1 subunit) together with a flexible tail (S2 subunit). Furthermore, HS-AFM captured conformational changes of S protein after interacting with its receptor, ACE2. Exosomes isolated from ACE2-expressing cells were found interacting with S protein. This observation was not found in exosomes isolated from ACE2-deficient cells, suggesting that the S protein-exosome interaction required ACE2. Lastly, the applicant filmed the insertion of S2 subunit (containing fusion peptide) into exosomes in a real-time manner. Like fusion peptide of HA, fusion peptide of S protein also disrupted exosomal layer stability. The results are published in the Journal of Extracellular Vesicles (PMID: 34874124).

Lastly, the applicant conducted HS-AFM imaging to study the interaction between anti-S protein neutralizing antibody (S NAbs) and S protein or S protein-expressing exosomes. This study demonstrates that HS-AFM could function as a nanoscopic platform for assessment of S NAbs against S proteins. The results are reported in Nano Letters (PMID: 36641798).

5. 主な発表論文等

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〔図書〕 計0件

〔産業財産権〕

〔その他〕

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6. 研究組織		
氏名 (ローマ字氏名) (研究者番号)	所属研究機関・部局・職 (機関番号)	備考

7. 科研費を使用して開催した国際研究集会

〔国際研究集会〕 計0件

8. 本研究に関連して実施した国際共同研究の実施状況

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