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研究課題名（和文） 肺血管内皮障害と術後早期の肺血栓塞栓症の発症について

研究課題名（英文） Prevention for the occurrence of the secondary pulmonary embolism related to the endothelial damage of pulmonary vasculature.

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研究成果の概要（和文）：術後翌日の肺血栓塞栓症は術中肺血管内皮障害による二次性の肺血栓塞栓症であることが示された。そのため術中超早期の血小板凝集抑制剤（フルルビプロフェン）の静脈内投与が即効性で効果的な治療方法である可能性を示した。

研究成果の概要（英文）：Pulmonary embolism was related to the endothelial damage of pulmonary vasculature on the day after surgery. Intravenous injection of flurbiprofen at the end of the surgery presents the possibility of the prevention of the secondary pulmonary embolism.

交付決定額

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年度			
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総計	3,500,000	1,050,000	4,550,000

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キーワード：肺血栓塞栓症、血管内皮障害、活性化血小板、経食道心臓超音波検査

1. 研究開始当初の背景

術中および術後の合併症として肺血栓塞栓症はいったん発症すると生命の危険を伴う重症の周術期合併症として認識されてきた。これまで周術期における塞栓物質の検出ならびに術後肺血栓塞栓症の出現頻度、重症度、術中所見との相関関係については詳細な報告はないにもかかわらず、術後肺血栓塞栓症の発症頻度は比較的低いと思われてきた。我々は20名の骨セメントを用いた人工股関節置換術において、術中経食道心臓超音波検査（TEE）による肺血栓塞栓症惹起物質の持続的検出を行った。その結果、骨セメントを使用した群において全例（20例中20例）に明らかな巨大塞栓子陽性所見を認めた。術翌日に肺血流シンチ検査を行ったところ、20例中5例（25%）において術後一日目にすで

に肺塞栓症の診断がなされた（Br J Anesth, 94, 607-12, 2005）。この肺塞栓症の発症頻度は一般の外科手術術後の肺塞栓発症頻度に比べ驚くべき異常に高い値であった。しかしながら、術直後の造影 MDCT では1例も肺血栓塞栓症の発生を確認できなかった。これらの結果より術中 TEE で確認された巨大塞栓子は直接的な肺動脈塞栓物質ではなく、術後第一日目に発生している早期肺血栓塞栓症の重要な惹起因子と考えられる。すなわち TEE で確認された巨大塞栓子は一時的に肺動脈に補足され、骨セメント含有成分などにより同部位の肺血管内皮傷害をもたらし、さらに手術部位で発生する炎症性サイトカインなどによる遠隔臓器への波及や術当日の血液過凝集状態により傷害された肺血管内皮上において二次血栓を形成している

可能性が高いと推測される。この事は、術後超早期における抗血液凝固治療や血小板機能抑制薬投与による肺血栓塞栓症発生の抑制作用を示唆するものである。

2. 研究の目的

今回我々は特に血小板凝集抑制療法を含めた抗血液凝固療法の肺血栓塞栓症の発症に対する治療方針と治療開始時期の確立および治療効果の判定を行った。

研究1

術中フルビプロフェンアキセチル（ロピオン）静脈内投与による血小板機能抑制効果について

研究2

手術侵襲による血小板活性化の経時的変化について

研究3

静脈麻酔薬（プロポフォール）のアナンダマイト誘発性血管内皮症が作用抑制効果について

手術時には凝固能、血小板機能が改善した状態であり、そこに外科侵襲が加わると過凝集状態に陥る可能性もある。そのため、術中の止血操作が済めばなるべく早期に凝集を励起する血小板凝集能を抑制することが求められる。ヘパリンなども使用されるが、その調節性は乏しく過剰な凝集抑制とともに、一次凝集である血小板凝集能は亢進させてしまうなど、血栓形成防止として術中には投与されることは少ない。一方、アスピリンに代表される NSAIDs（非ステロイド性消炎鎮痛薬）は古くから鎮痛薬として臨床に用いられ、また血小板抑制作用を持つため、血栓性疾患や動脈硬化性疾患患者には術前の経口抗血小板薬として頻繁に用いられてきた。しかし、その調節性のなさから術中使用は困難であった。一方、フルビプロフェンは唯一静注できる NSAIDs であり、必要時に静脈内投与により急性の血小板抑制作用を得ることができる。また術後の鎮痛薬として適応もあり頻繁に臨床使用されている。しかしながら、その血小板抑制作用の程度と発現時間など詳細は分かっていない。そこで、今回開腹術患者を対象に術中止血が完成された時点においてフルビプロフェンを投与し、その作用程度と発現時間を確認するために投与前後における血小板機能検査、凝固検査を行った。血小板機能検査は、これまで行われてきた混濁法とともに、外科処置による止血機構にもっとも関与する血小板凝集塊のサイズ別の増加評価をすることを目的とした。

3. 研究の方法

対象：19名の外科手術患者を対象とした。
方法：止血が確認された時点でフルビプロフェンを50mg（1A）緩徐に静注した。
測定：投与前、投与15分後に動脈モニタ路

から血液を採取した。

測定項目：血算：Hb、Ht、全血血小板数（Plt）、濃厚血小板血漿中血小板数（PRP-Plt）、凝固系検査：PT、APTT、フィブリノーゲン、血小板機能：透過度（1分、3分、5分、7分）、コラーゲン励起7分後の粒子径比率

4. 研究成果

術中止血確認後のフルビプロフェンアキセチル（ロピオン）静脈内投与による血小板機能抑制効果は投与15分後には血小板凝集抑制作用が発現された。また、その持続時間は6時間を超えた。フローサイトメトリーによる検討ではフルビプロフェンアキセチルによる血小板凝集抑制作用は血小板凝集作用には影響がなく、血小板凝集塊の増大を抑制することがわかった。またその作用程度はチエノピリジン系抗血小板剤（チクロピジン、クロピドグレル）とほぼ同等であることがわかった。これらにより、不安定狭心症、冠動脈ステント挿入患者など、血小板凝集亢進による凝集塊が危機の状態につながる場合、術中止血が確認直後からフルビプロフェンアキセチル静脈内投与による血小板凝集抑制作用をもたらすことが危機の状態の発生抑制につながる可能性を示すことができた。

研究1結果

性別 (M/F)	年齢 (歳)	身長 (cm)	体重 (kg)	手術時間
17/2	62 ± 14	166 ± 7	63 ± 9	4:48

血算

	Hb	Ht	Plt	PRP-Plt
投与前	9.4 ± 1.6	28.5 ± 4.8	17.8 ± 5.8	16.3 ± 5.9
投与後	9.4 ± 1.7	28.2 ± 4.9	17.8 ± 7.6	16.6 ± 5.7

Hb；ヘモグロビン(g/dl)、Ht；ヘマトクリット%、Plt；血小板数、PRP-Plt；濃厚血小板血漿中血小板数(x104/ml)

凝固系

	PT (sec)	PT (INR)	APTT (sec)	Fibrinogen
投与前	12.4 ± 0.8	1.05 ± 0.07	40.6 ± 11.2	263 ± 74
投与後	12.9 ± 1.0	1.10 ± 0.09	41.4 ± 11.0	236 ± 69

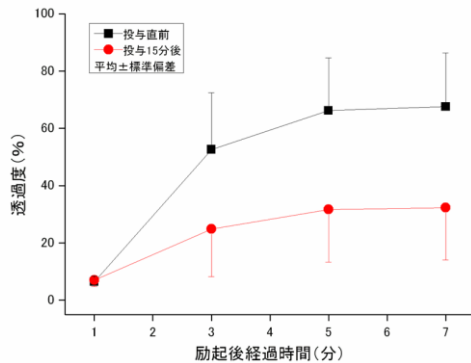
血小板凝集能（PA-200）検査結果

Collagen 20ug/ml 刺激

	透過度 (%)			
	1min	3min	5min	7min
投与前	6.4 ± 2.0	52.6 ± 19.9	66.2 ± 18.4	67.6 ± 18.7
投与後	7.0 ± 2.3	24.9 ± 16.7	31.7 ± 18.4	32.3 ± 18.2

透過度の経時的変化

フルルビプロフェン投与15分後には透過度を用いた血小板凝集能検査では投与前に比べ50%以下に抑制された。

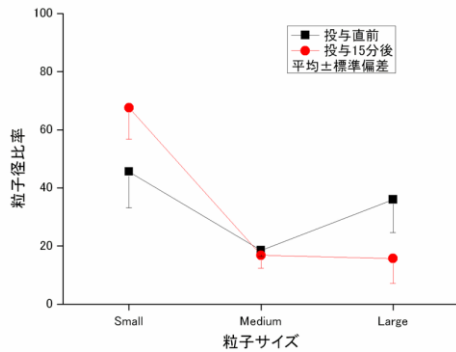


透過度抑制の要因を考察するために、血小板サイズ別比率をコラーゲン励起後7分値において評価した。

励起7分時の粒子径比率

	Small	Medium	Large
投与前	45.6 ± 12.4	18.5 ± 2.1	36.0 ± 11.3
投与後	67.6 ± 10.8	16.8 ± 4.5	15.7 ± 8.6

Range : Small 2 ~ 20, Medium 21 ~ 27, Large 28 ~ 32



研究2結果

麻酔導入後からフルルビプロフェン投与直前までの血液凝固能と血小板凝集能変化血算

	Hb	Ht	Plt	PRP-Plt
麻酔導入後	10.4 ± 2.3	31.3 ± 6.2	25.5 ± 11.1	23.5 ± 7.0
投与直前	9.7 ± 2.1	29.8 ± 5.9	19.6 ± 10.6	19.3 ± 9.4

Hb:ヘモグロビン(g/dl)、Ht:ヘマトクリット%、Plt:血小板数、PRP-Plt:濃厚血小板血漿中血小板数(x104/ml)

凝固系

	PT (sec)	PT (INR)	APTT (sec)	Fibrinogen
麻酔導入後	12.4 ± 0.7	1.1 ± 0.1	37.2 ± 1.2	336.7 ± 116.9
投与直前	13.1 ± 1.2	1.1 ± 0.1	41.4 ± 13.3	241.3 ± 93.5

血小板凝集能 (PA-200) 検査結果

Collagen 20ug/ml 刺激

	透過度 (%)			
	1min	3min	5min	7min
麻酔導入後	4.9 ± 2.2	56.1 ± 20.5	70.0 ± 14.6	69.4 ± 13.9
投与直前	5.9 ± 2.2	58.0 ± 19.3	68.8 ± 17.2	71.5 ± 18.0

透過度の経時的変化

麻酔導入後からフルルビプロフェン投与直前における透過度を用いた血小板凝集能検査では抑制は見られなかった。

透過度抑制の要因を考察するために、血小板サイズ別比率をコラーゲン励起後7分値において評価した。

励起7分時の粒子径比率

	Small	Medium	Large
麻酔導入後	33.8 ± 7.1	20.1 ± 2.4	46.4 ± 7.7
投与直前	41.4 ± 14.2	19.4 ± 2.1	39.3 ± 13.3

研究3

Propofol Protects against Anandamide - Induced Injury in Human Umbilical Vein Endothelial Cells

The endocannabinoid anandamide, (i.e., arachidonylethanolamine (AEA)) has been isolated from porcine brain lipid extract as an endogenous ligand for cannabinoid receptors in the central nervous and immune systems. AEA is synthesized from N-arachidonoyl phosphatidylethanolamine in depolarized neurons, macrophages, endothelial cells and platelets, and quickly degraded by the fatty amide acid hydrolase (FAAH) into arachidonic acid and ethanolamine. In normal humans, AEA exists at low levels in blood and cerebrospinal fluid. However, it has been demonstrated that the serum levels of AEA increase dramatically during the shock caused by either hemorrhage or sepsis, and play a crucial role in the pathogenesis of hypotension. Furthermore, in several types of cells, elevated levels of AEA can induce

apoptosis by producing free radicals such as reactive oxygen species (ROS), and the production of ROS is exacerbated by the inhibition of FAAH.

Vascular endothelial cells have important physiologic functions as barriers, and in maintaining cardio-vascular homeostasis and vascular stability. However, this function may be impaired in septic shock and ischemia-reperfusion injury, resulting in cellular necrosis and apoptosis. On the other hand, propofol (2,6-diisopropylphenol), an intravenous general anesthetic, possesses an antioxidant property because it contains a phenolic hydroxyl group that gives electrons to the free radicals generated during ischemia and reperfusion. The purpose of the present study was to examine the protective effect of propofol against AEA-induced cellular injury using human umbilical vein endothelial cells (HUVECs).

MATERIALS AND METHODS

Cell culture

The initial batch of HUVECs was purchased from Lonza, Inc. (Basel, Switzerland), and cultured in endothelial growth medium-2 (EGM-2), consisting of ascorbic acid, fibroblast growth factor, hydrocortisone, insulin-like growth factor-1, vascular endothelial growth factor, gentamicin, amphotericin-B, and 10% fetal bovine serum (FBS). The cells were grown in a humidified incubator at 37°C containing 95% air and 5% of carbon dioxide with media replenishment every 3 days. Following growth to 90% confluence, the cells were split and grown to confluence again. Before the experimental intervention, confluent HUVECs with 2-5 passages were incubated in a starved medium supplemented with 1% of FBS for 4 h. In the ROS production assay and the caspase-3 activity assay, HUVECs were divided into three experimental groups characterized by culture medium conditions as 1) control, 2) cultured with AEA alone, or 3) pretreatment with propofol for 30 min, then co-incubated with AEA.

Evaluation of HUVECs viability exposed to AEA

As shown in Figure 1A, the HUVECs were coincubated with AEA (0, 0.01, 0.1, 1, 2.5, 5, 7.5 and 10 μ M) at 37°C for 24 h. Then, the cell viability was evaluated using a 3-(4,

5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay. In brief, 10 μ L of MTT (Sigma Chemical, St. Louis, USA) solution (5 mg/L) was administered to each incubating well, and the wells were incubated at 37°C for 5 h. The formazan in each well, produced by the MTT assay, was dissolved in 100 μ L of dimethyl sulfoxide. The absorbance of this colored solution was measured at 570 nm by a spectrophotometer. According to the results of this experiment, 10 μ M of AEA was utilized as a positive control for AEA-induced cell injury in all subsequent experiments.

Effect of propofol on HUVEC viability

Propofol was prepared by diluting Diprivan (AstraZeneca, London, UK) with EGM-2 to a concentration of 10, 50, or 100 μ M. After pretreatment of the culture medium with propofol for 30 min at the above concentrations, the HUVECs were stimulated without or with 10 μ M of AEA and incubated at 37°C for 24 h. Thereafter, the cell viability was observed employing the MTT assay as mentioned above.

Analysis of ROS production

The Image-iT live green reactive oxygen species detection system (Molecular Probes, Eugene, OR), counterstained with Hoechst 33342 for nuclei, was used to visualize reactive oxygen species in live HUVECs under a microscope (Olympus FV1000, Tokyo, Japan), using fluorescein filter sets.

Intracellular ROS formation was detected using 5-(and-6)-carboxy-2',7'-dichlorodihydrofluorescein diacetate (carboxyl H2DCFDA) as previously reported. Briefly, starved HUVECs, seeded at a density of 2.0×10^4 cells/well, were loaded with the redox sensitive dye carboxyl H2DCFDA for 45 min, washed and pretreated with propofol (50 μ M), and then stimulated with AEA alone (10 μ M) or in combination with propofol (50 μ M) at 37°C for 1 h (Fig. 1C). Then, ROS levels were measured with a multiwell fluorescence plate reader (Tecan, Mannedorf, Switzerland), using excitation and emission filters of 485 nm and 535 nm, respectively.

Caspase-3 activity assay

The caspase-3 activity in the incubated HUVECs was colorimetrically assayed 8 h following treatment with AEA alone (10 μ M) or in combination with 50 μ M of propofol. The cells were washed, lysed, and incubated with the caspase-3 specific

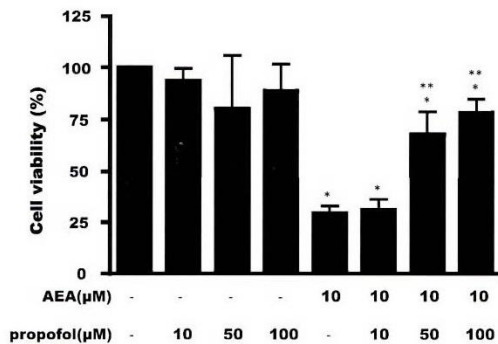
labeled substrate, N-acetyl-Asp- Glu-Val- Asp-p-nitroanilide (Ac-DEVD- pNA), on ice for 15 min. The chromophore p-nitroanilide (pNA), which was released from Ac-DEVD- pNA upon cleavage by caspase-3, could be quantified using a microtiter plate reader at 405nm. The relative increase in caspase-3 activity was determined by comparing the absorbance of pNA from an apoptotic sample to an uninduced control.

RESULTS

Reduction in HUVECs viability due to AEA

Evaluating cell viability by an MTT assay 24 h after the AEA exposure showed that 5 μ M, 7.5 μ M and 10 μ M of AEA caused a significant deterioration of cultured HUVECs viability to 79.2 \pm 2.3%, 58.3 \pm 1.2% and 38.5 \pm 2.1%, respectively, compared with the control. Protective effect of propofol on HUVECs viability

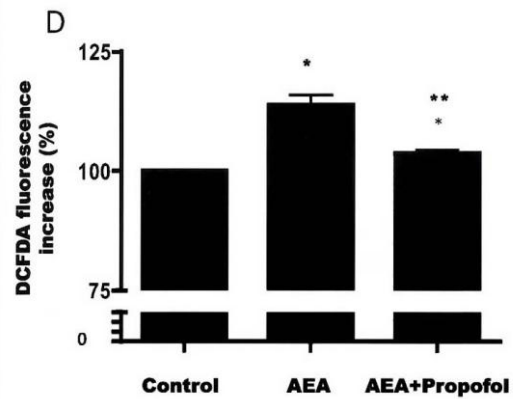
Propofol itself did not affect the cell viability at the concentrations of 10-100 μ M. Of great importance was that pretreatment with 50 and 100 μ M of propofol significantly increased viability in cells exposed to 10 μ M of AEA from 29.7 \pm 3.2%, to 67.9 \pm 11.0% and 78.4 \pm 6.5%, respectively, although 10 μ M of propofol had no protective effect.



Decrease in ROS production by propofol

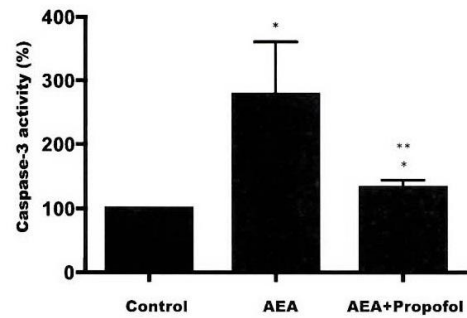
HUVECs stimulated with AEA alone were ROS-positive (green), whereas the pretreatment of cells with propofol significantly attenuated the green ROS signal, and almost no green ROS signal was observed in controls.

ROS production in the cultured HUVECs 1h after exposure to 10 μ M of AEA was significantly attenuated by 50 μ M of propofol falling from 113.8 \pm 2.0% in the AEA-exposed cells to 103.7 \pm 1.0% in the propofol treated cells.



Protected caspase-3 activity by propofol

Caspase-3 activity 8 h after the exposure to 10 μ M of AEA in the cultured HUVECs was significantly ameliorated by 50 μ M of propofol, dropping from 277.6 \pm 83.3% in the AEA-exposed cells, to 132.1 \pm 12.0% in the propofol-treated cells.



DISCUSSION

This study demonstrated the protective effects of propofol against AEA-induced cell injury. One of the crucial mechanisms of cell injury due to anandamide is surmised to be ROS production followed by apoptosis. Siegmund et al. reported that glutathione, an antioxidant, attenuated the AEA-derived ROS formation and effectively suppressed the death of primary hepatic stellate cells. Propofol is also known to have antioxidant activity in scavenging ROS and suppressing apoptosis, coinciding with our results.

Lipid rafts, localized in gamma-aminobutyric acid A receptors, are supposed to play an important role in AEA-induced cell death. One possible protective mechanism of propofol might be its action on these receptors.

As mentioned above, AEA is metabolized by FAAH and the inhibition of FAAH could enhance AEA-induced cellular toxicity. Meanwhile, propofol is thought to be an FAAH inhibitor. However, Schelling et al.

observed no remarkable increase in blood level of AEA during general anesthesia using propofol. It is not clear whether the mechanism by which propofol protected HUVECs against AEA-induced injury in this study involved the inhibition of FAAH. It is notable that in the present study propofol exerted a protective effect at a blood concentration level used clinically to achieve general anesthesia, (i.e., 10-60 μ M). This finding suggests the clinical usefulness of propofol as a general anesthetic for patients with shock caused by hemorrhage, endotoxin, and so forth. There have been a number of reports on the blood concentration of AEA in normal humans, most of which were conducted on the nM-level. However, this could increase to the μ M-level in endotoxic shock, and a level of over 10 μ M, at which the death of cells was observed, could be considered a pathologic state. Concerning anandamide-induced cell death, the involvement of nitric oxide and several receptors of cannabinoid and capsaicin, and the differences in pathway leading to cell death depending on the types of cell have been suggested. Further investigations are needed to clarify both the mechanism of and the therapeutic strategy for anandamide-induced cell death.

In conclusion, our results suggest that propofol at clinically used blood concentrations protects HUVECs against AEA-induced injury, in part by suppressing ROS production and subsequent apoptosis.

5. 主な発表論文等

(研究代表者、研究分担者及び連携研究者には下線)

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○出願状況 (計 0 件)

○取得状況 (計 0 件)

[その他]

ホームページ等

6. 研究組織

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